

Translational research in breast cancer

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Outline

- Introduction to the clinical classification
- Molecular characterization and intertumor heterogeneity
 - Examples of clinical applications
- The challenge of intratumor heterogeneity
- The role of the tumor immune microenvironment
- Novel translational research tools and future perspectives

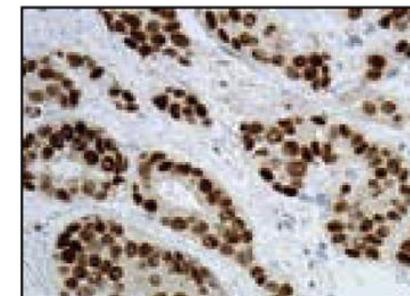
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- Introduction to the *clinical* classification
- Molecular characterization and intertumor heterogeneity
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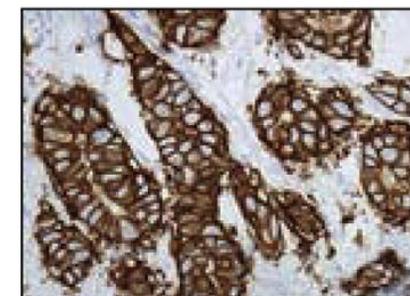
Clinical classification of breast cancer (BC) – markers

Biomarker	Method and threshold	Use	LOE
ER	IHC; positive if $\geq 1\%$	<ul style="list-style-type: none"> Essential for the characterization of the IHC luminal group Poor prognostic marker if negative Predictive marker for endocrine treatment Mandatory for endocrine treatment prescription 	I
PR	IHC; positive if $\geq 1\%$	<ul style="list-style-type: none"> If negative, tumour classified as IHC luminal B Strong poor prognostic marker if negative Predictive marker for endocrine treatment 	I
HER2	<ul style="list-style-type: none"> IHC; positive if $>10\%$ complete membrane staining (3+) Single-probe ISH; positive if HER2 ≥ 6 copies Dual-probe ISH; positive if HER2 and CEP17 ≥ 2 and HER2 ≥ 4 copies, or HER2 and CEP17 < 2 and HER2 ≥ 6 copies 	<ul style="list-style-type: none"> Essential to characterize HER2-enriched (ER-negative) disease and luminal B, HER2-positive Prognostic marker Predictive marker for anti-HER2 treatment Mandatory for anti-HER2 therapy 	I (IHC) and I (ISH)
Ki67	IHC; no final consensus on cut-off value but values $<10\%$ are considered low and $>30\%$ are considered high ^a	Absence of international consensus for scoring and threshold	I
		Prognostic value in ER-positive, HER2-negative tumours (primary tumours and post-neoadjuvant tumour residues)	I
		Absence of prognostic value in HER2-positive disease or TNBC	I
		Predictive of response to neoadjuvant endocrine therapy ^a	I
		Predictive of response to neoadjuvant chemotherapy	Expert opinion
		If elevated, chemotherapy is often prescribed in ER-positive, HER2-negative tumours	Expert opinion
		Part of the IHC definition of luminal tumours whereby when Ki67 is low, luminal A tumour likely and when Ki67 high, luminal B tumour likely	Expert opinion

A



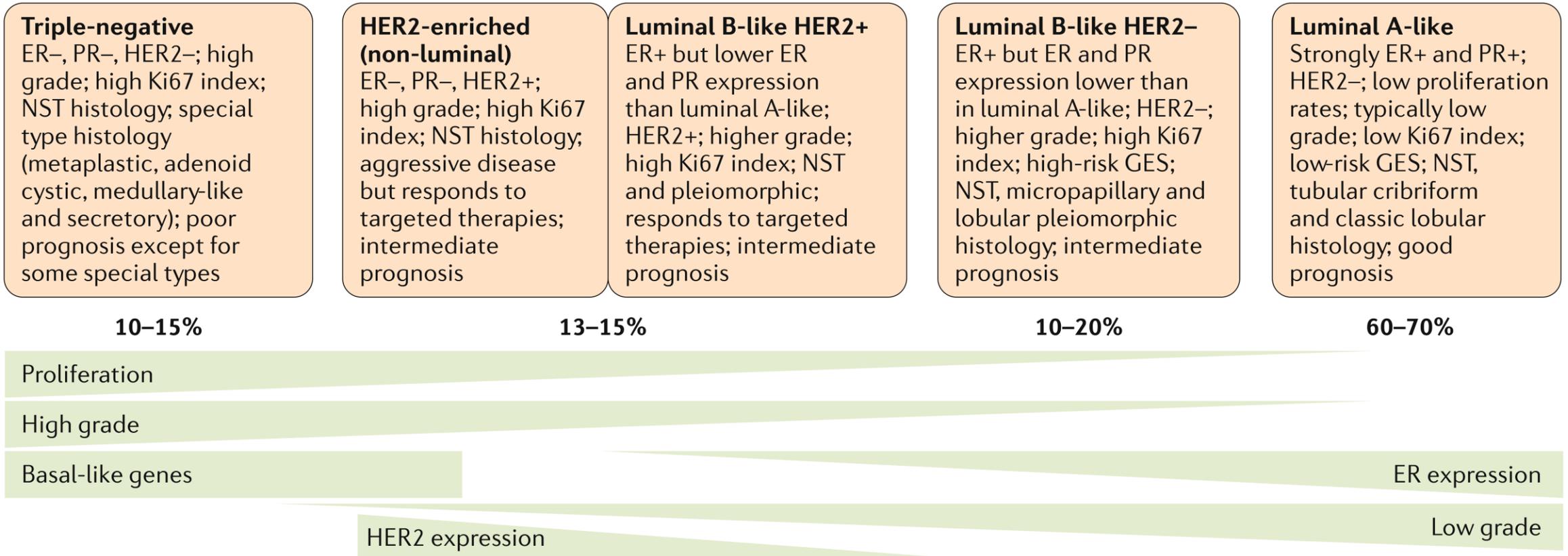
B



Expression of (A) ER and (B) HER2 assessed by IHC

ER = estrogen receptor; PR = progesteron receptor; HER2 = human epidermal growth factor receptor 2; IHC = immunohistochemistry; ISH = *in situ* hybridization; LOE = level of evidence.

Clinical classification – *subtypes* based on immunohistochemistry



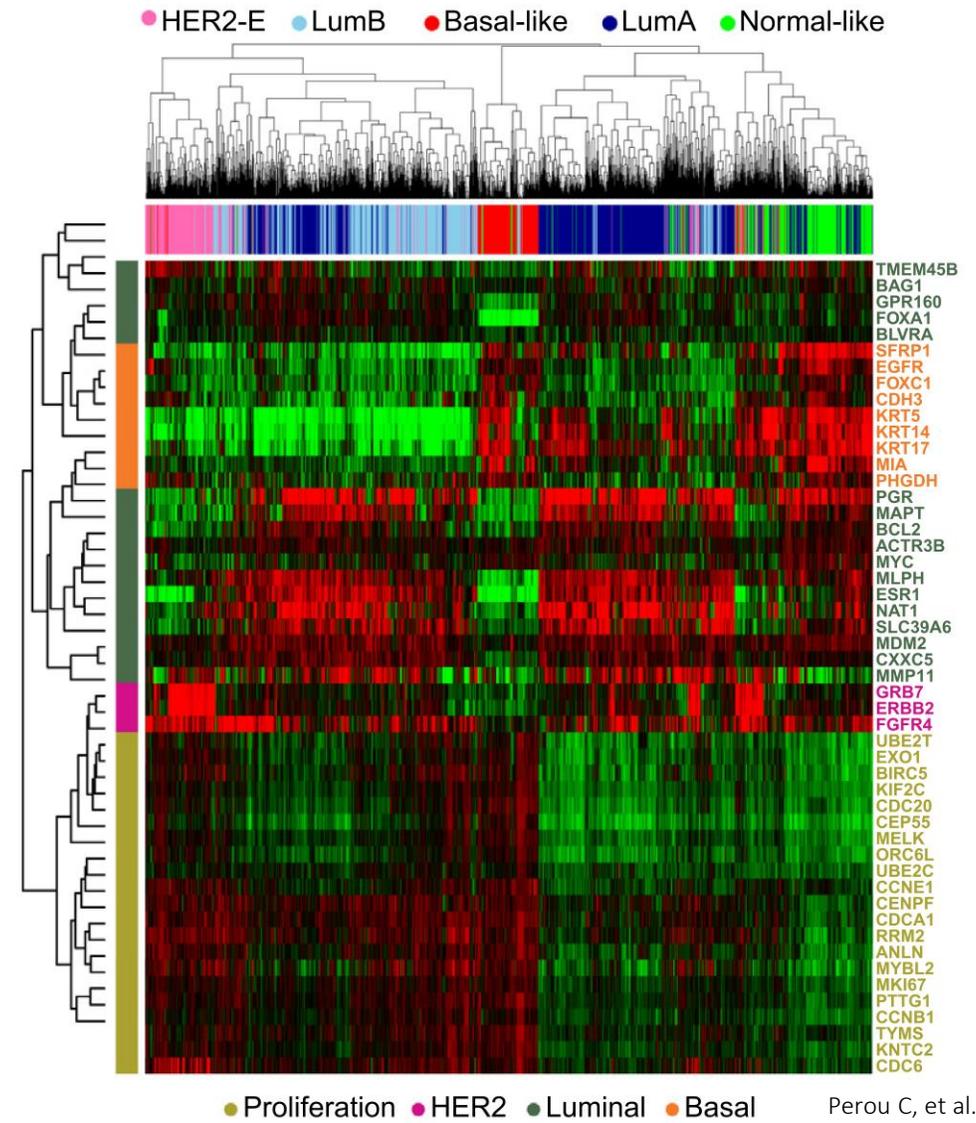
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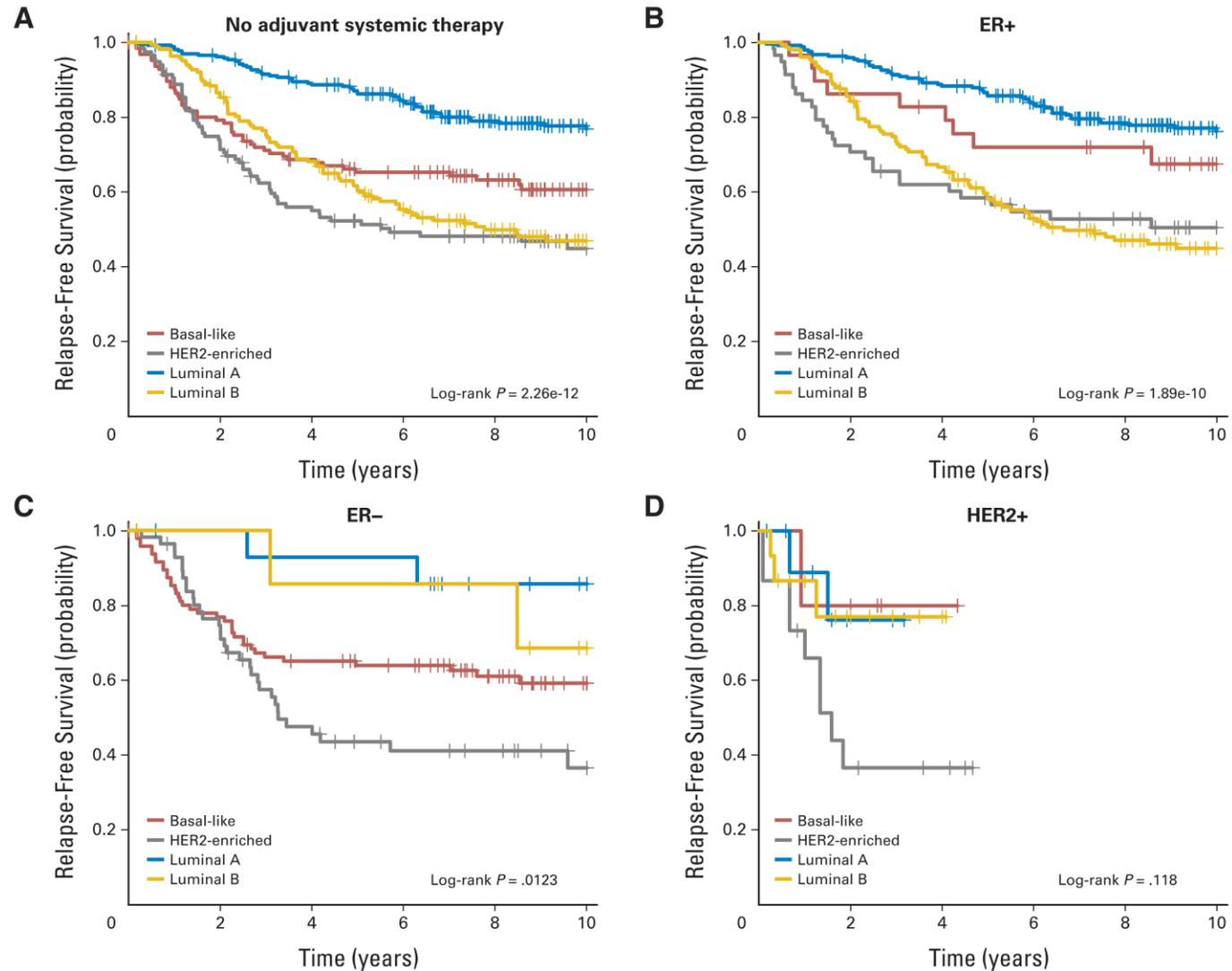
Gene expression-based “intrinsic” subtypes – PAM50

- 2000: Gene expression studies initially identified 4 “intrinsic” subtypes
- Subsequent studies **refined** the classification and demonstrated **prognostic significance**
- Prediction Analysis of Microarray (PAM) 50 subtypes (based on 50 genes):
 - Luminal A
 - Luminal B
 - HER2-enriched
 - Basal-like
 - Normal-like (may represent non-cancer cells “contaminating” bulk tissue samples)

→ Differences in biological processes



PAM50 subtypes and *prognosis*



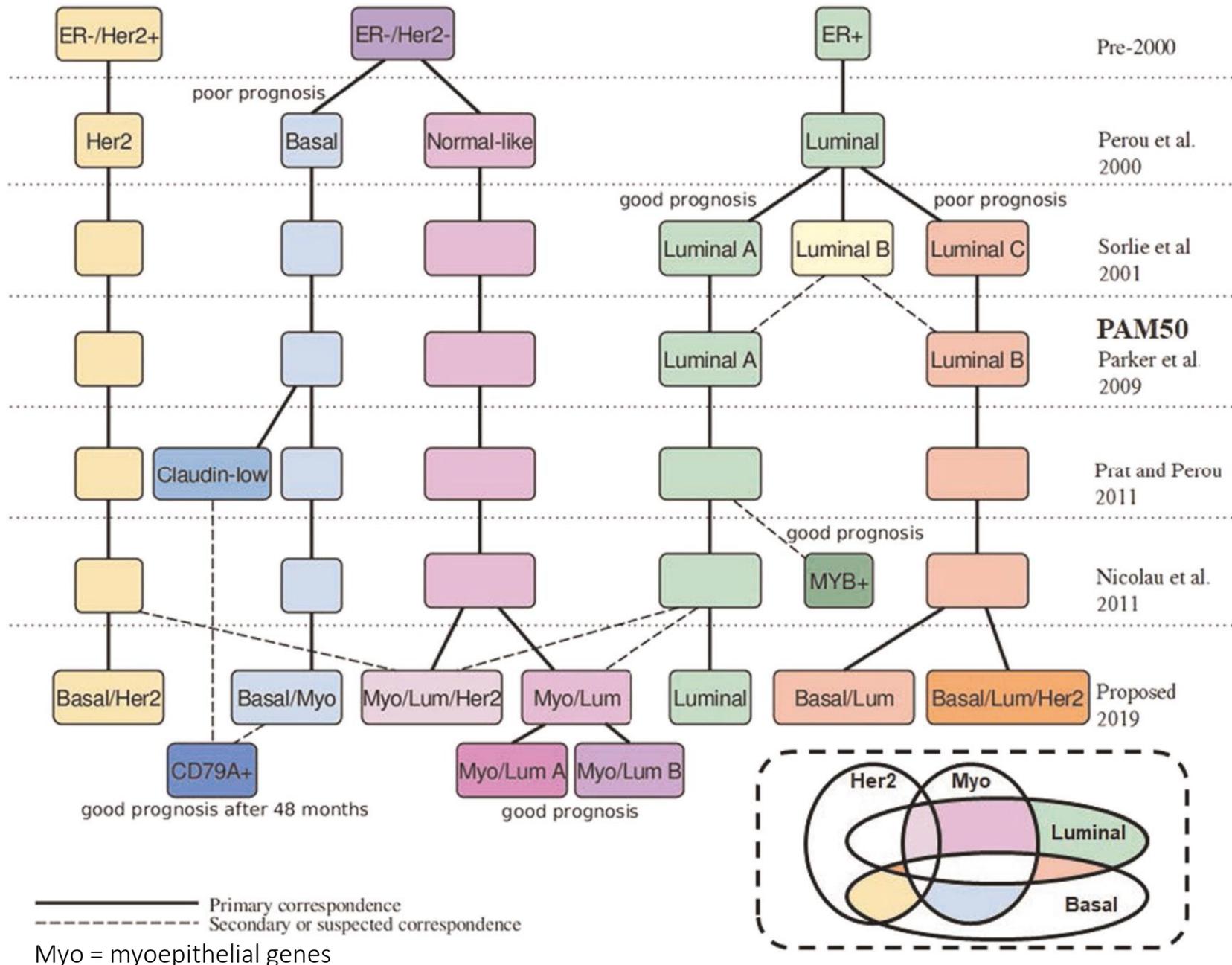
PAM50 vs “clinical” classification

Distribution of the PAM50 intrinsic subtypes within the pathology-based groups.^a

IHC-based group	References	N	PAM50 intrinsic subtype distribution			
			Luminal A	Luminal B	HER2-enriched	Basal-like
HR+/HER2–	[10,14,16–22]	4295	60.3%	31.9%	6.6%	1.2%
Luminal A	[10,14,17,21]	637	62.2%	27.0%	10.2%	0.6%
Luminal B	[10,14,17,21]	317	34.1%	51.1%	11.0%	3.8%
HER2+	[6,23–26]	831	17.6%	26.8%	44.6%	11.0%
HER2+/HR+	[25,26]	182	33.0%	46.2%	18.7%	2.2%
HER2+/HR–	[25,26]	168	19.0%	4.2%	66.1%	10.7%
TNBC	[12–15]	868	1.6%	3.2%	9.1%	86.1%

^a The data has been obtained from the different publications. Several studies have performed a standardized version of the PAM50 assay (RT-qPCR-based or nCounter-based) from formalin-fixed paraffin-embedded tumour tissues [10,14,17,19–22], while others have performed the microarray-based version of the PAM50 assay [6,16,18,23–26].

- Combined the data from studies for a total of 5994 independent samples
- **Overall discordance of ~30%**
- The **two methods** to identify intrinsic biology should **not be considered the same**
- 3 or 4 biomarkers do not fully recapitulate the intrinsic subtypes of breast cancer

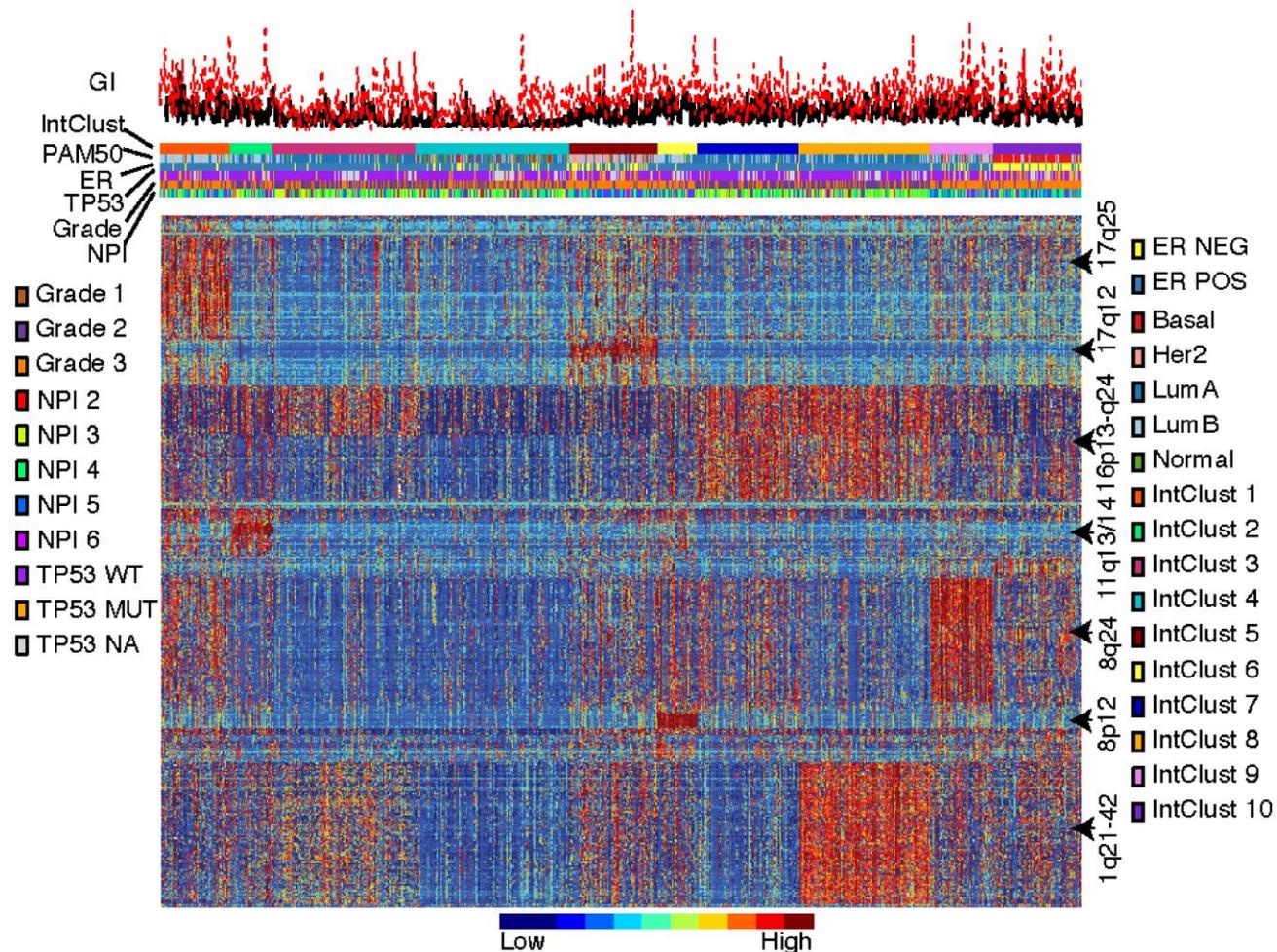


The METABRIC* study

Somatically acquired copy number aberrations (CNAs) are the **dominant feature** of BC

- Collection of ~2000 primary BC samples
- **None** of the HER2-positive patients **received** trastuzumab (!)
- Integrated analysis of *copy number* and *gene expression*

→ CNAs influencing gene expression in *cis* likely to be enriched in driver genes

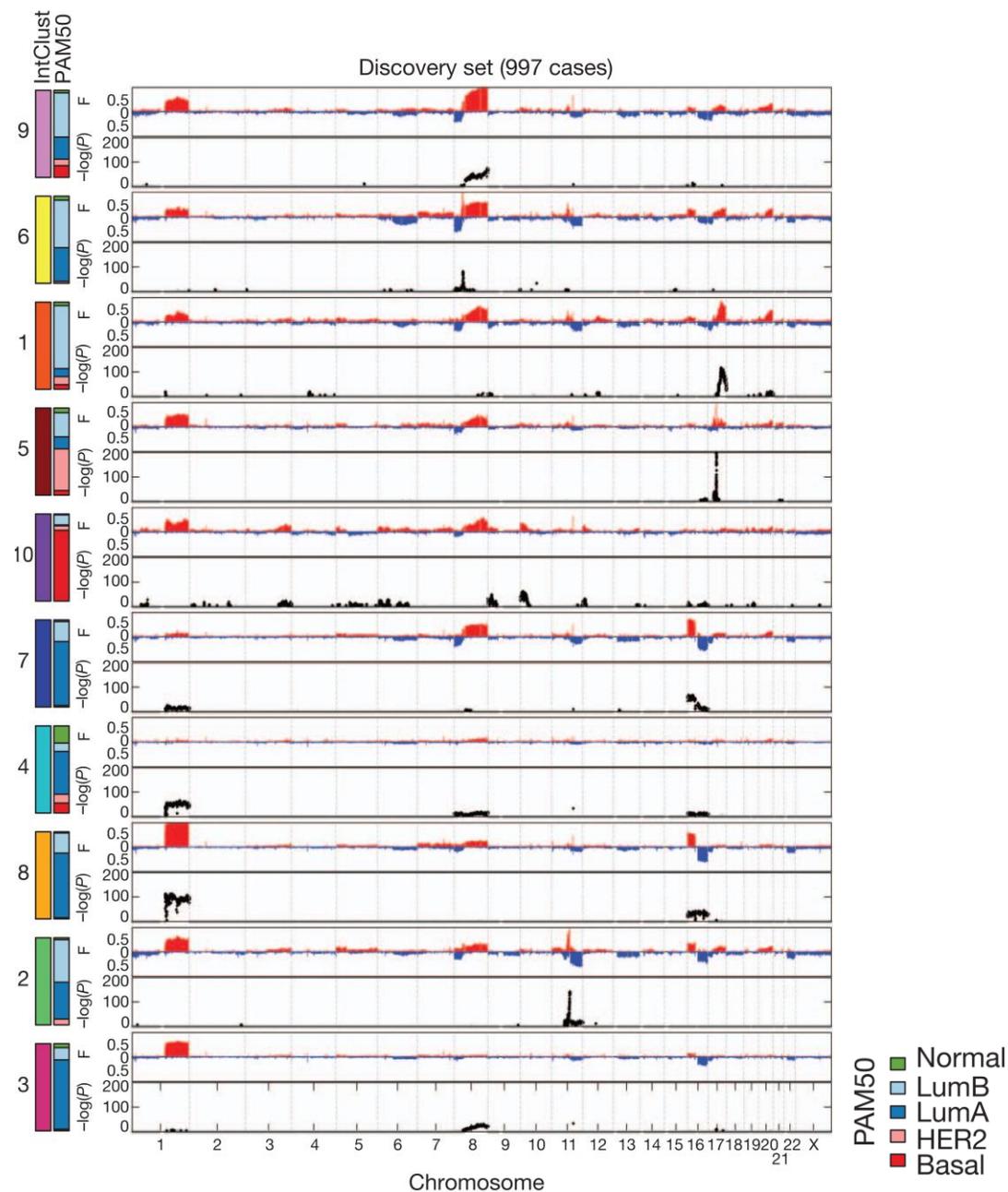


Unsupervised analysis of paired DNA–RNA profiles revealed **10 novel subgroups (Integrative Clusters - IntClusts)**

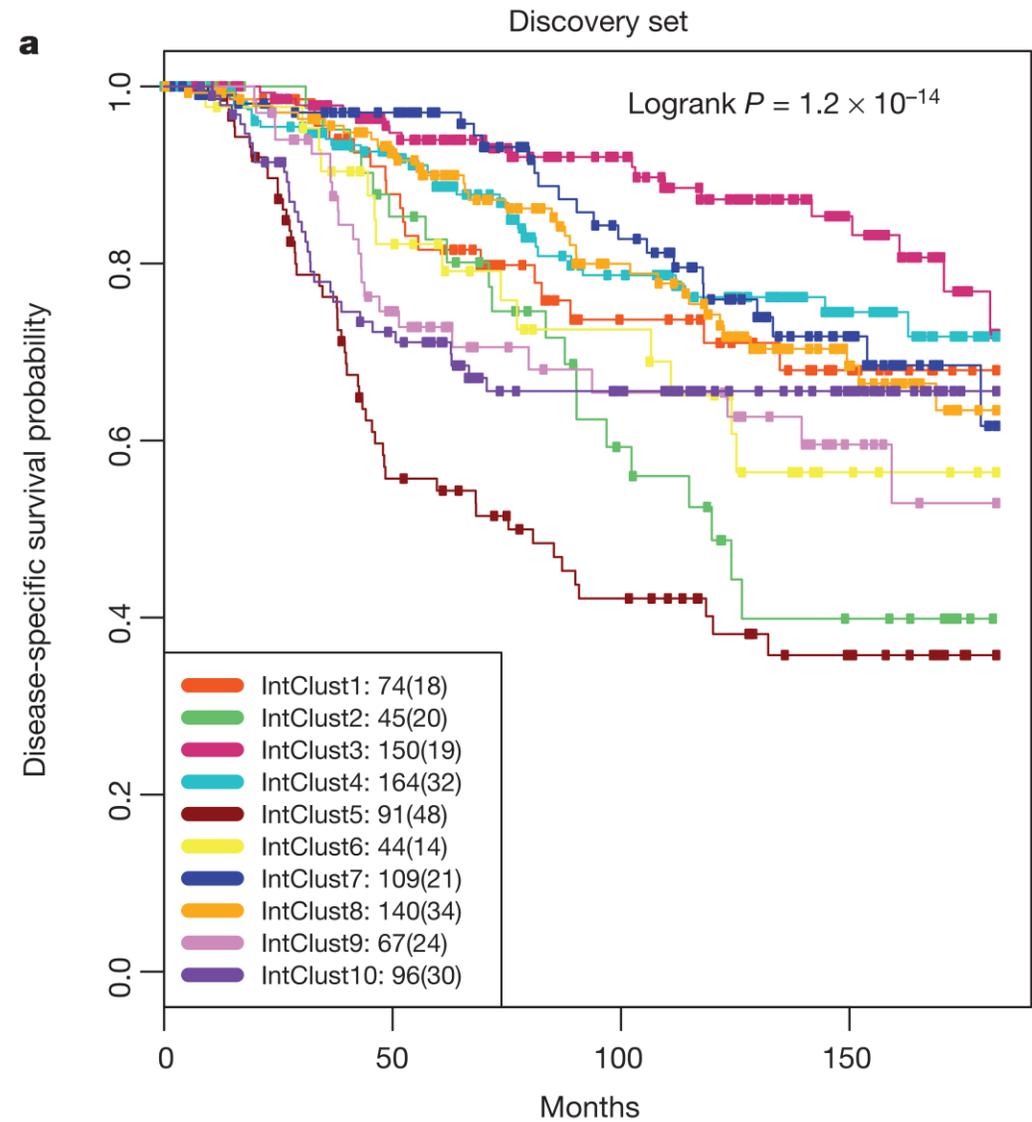
GI = genomic instability; NPI = Nottingham prognostic index

* Molecular Taxonomy of Breast Cancer International Consortium

Cis = Variant at a locus has an impact on its own expression \neq *trans* when it is associated with genes in other sites in the genome



Regions of copy number **gain** are indicated in red and regions of **loss** in blue in the frequency plot



The Cancer Genome Atlas (TCGA)

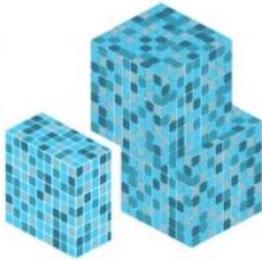
NATIONAL CANCER INSTITUTE THE CANCER GENOME ATLAS

TCGA BY THE NUMBERS

TCGA produced over

2.5

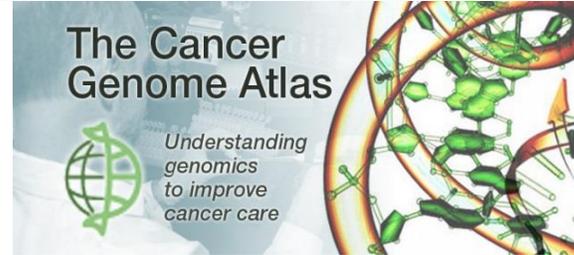
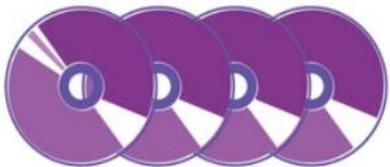
PETABYTES
of data



To put this into perspective, **1 petabyte** of data is equal to

212,000

DVDs



TCGA data describes



33

DIFFERENT
TUMOR TYPES

...including

10

RARE
CANCERS

...based on paired tumor and normal tissue sets collected from



11,000

PATIENTS

...using

7

DIFFERENT
DATA TYPES



10,000 Tumors
33 Cancer Types

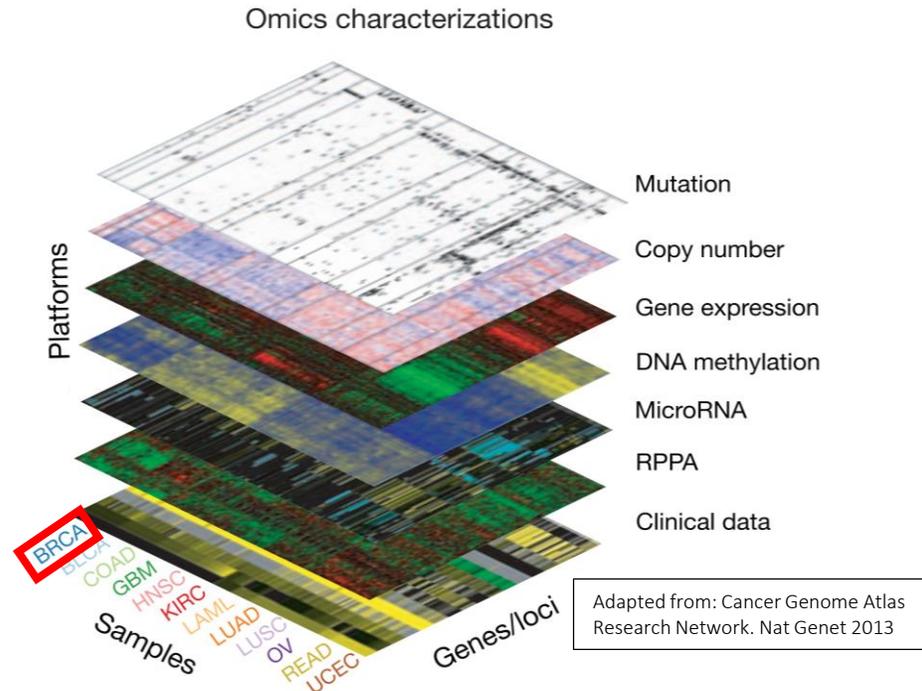
Clinical Data
Copy Number
Exome/Mutation
DNA Methylation
mRNA-Seq
microRNA-Seq
RPPA Protein

28 iClusters

Adapted from: Hoadley KA, et al. Cell 2019

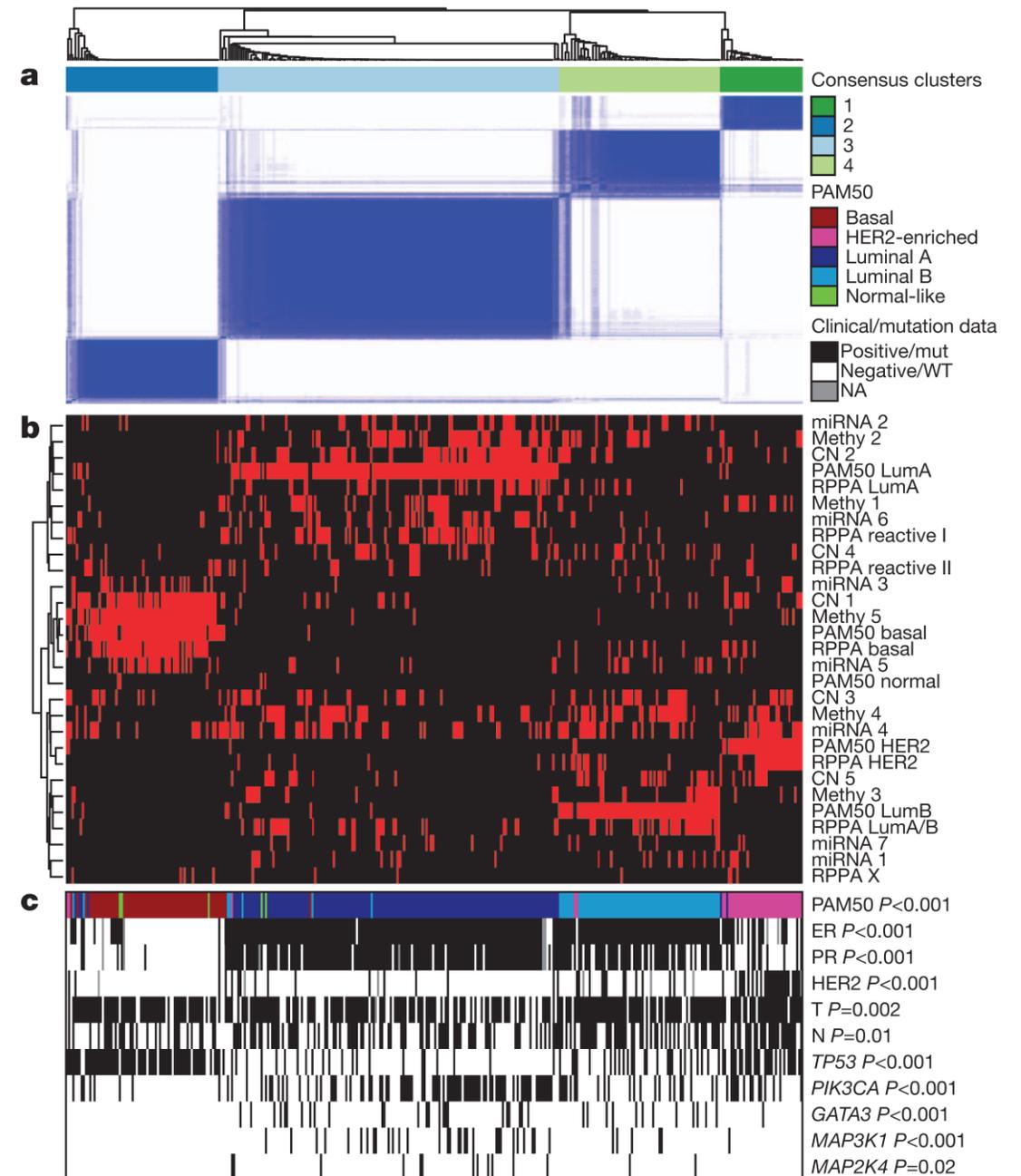
The Cancer Genome Atlas

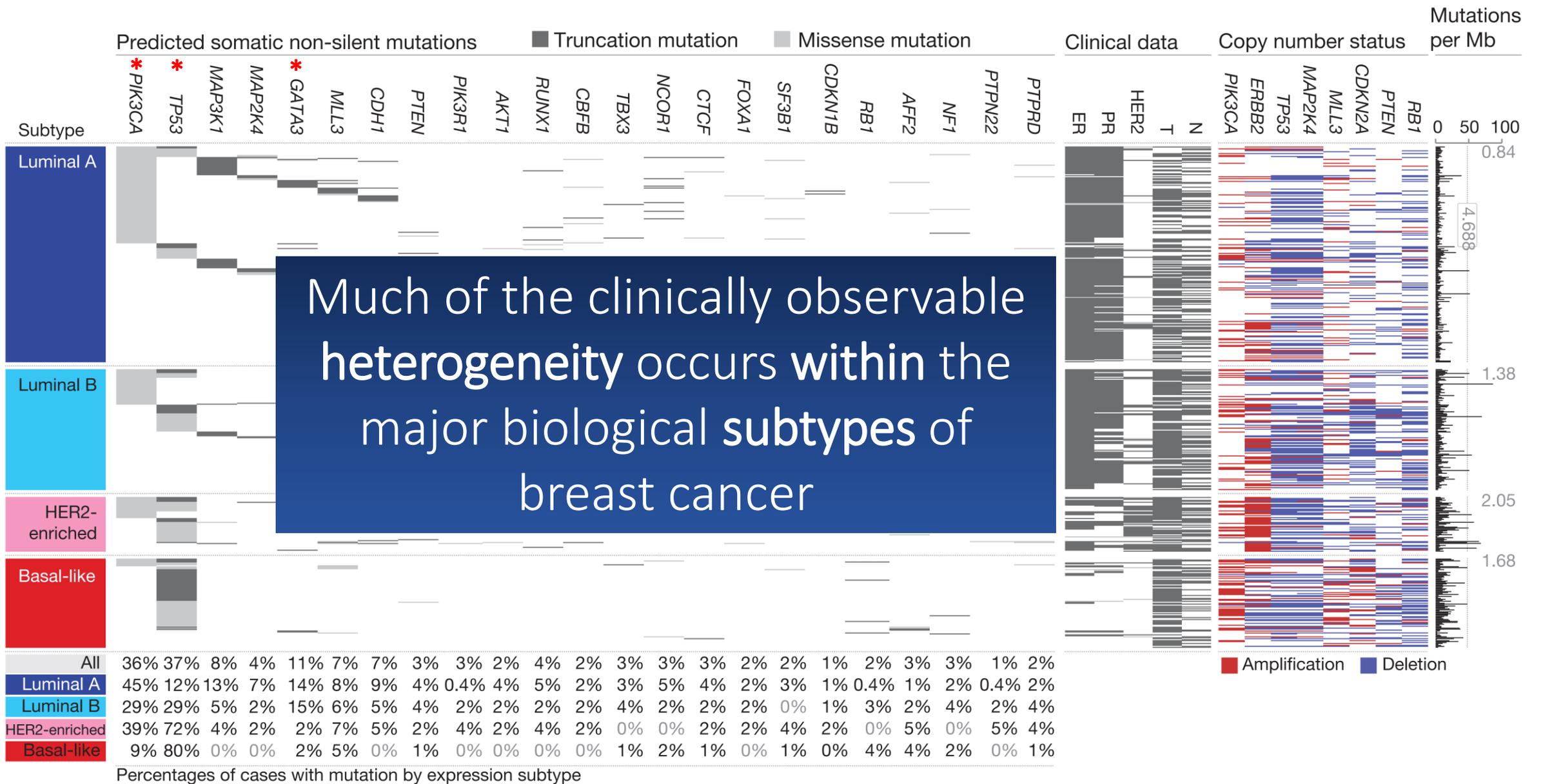
- Primary breast cancers from 825 patients



- Unsupervised clustering on data from five molecular platforms (N=348, not including WES) and *integration* of results

→ 4 main BC consensus clusters





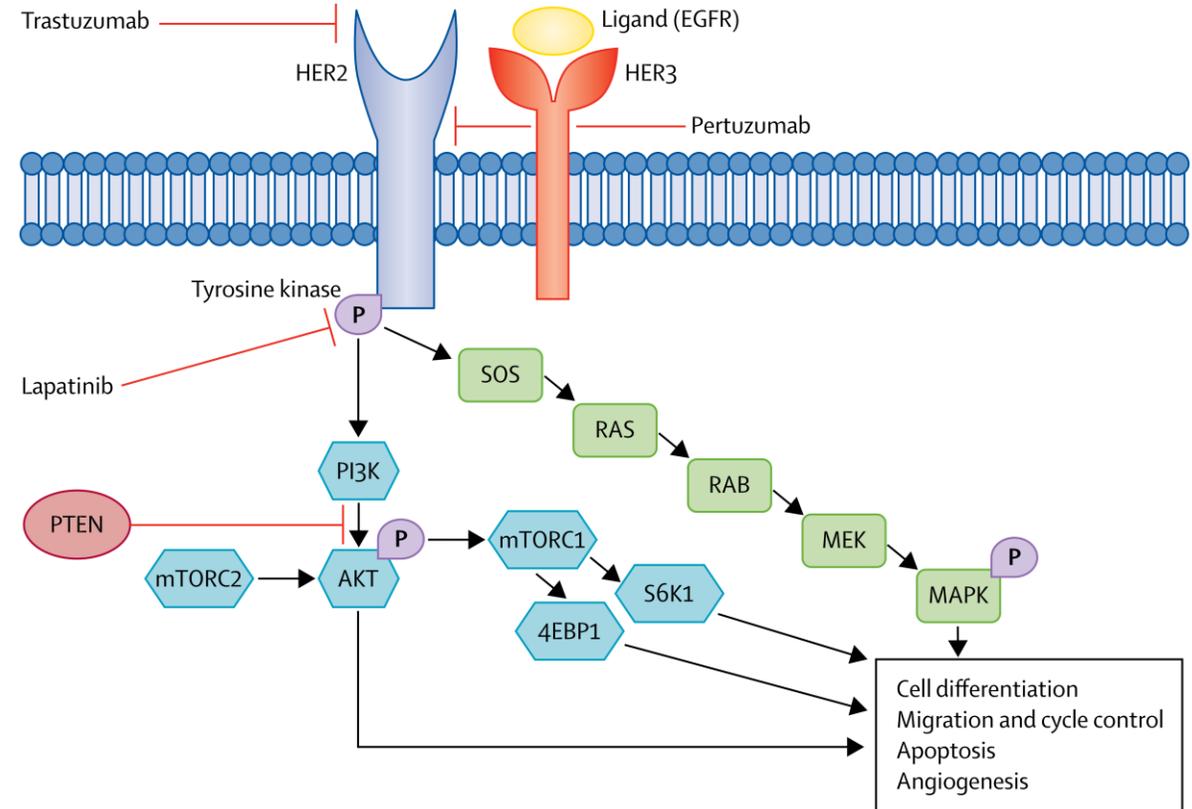
* >10% incidence across all BCs

Outline

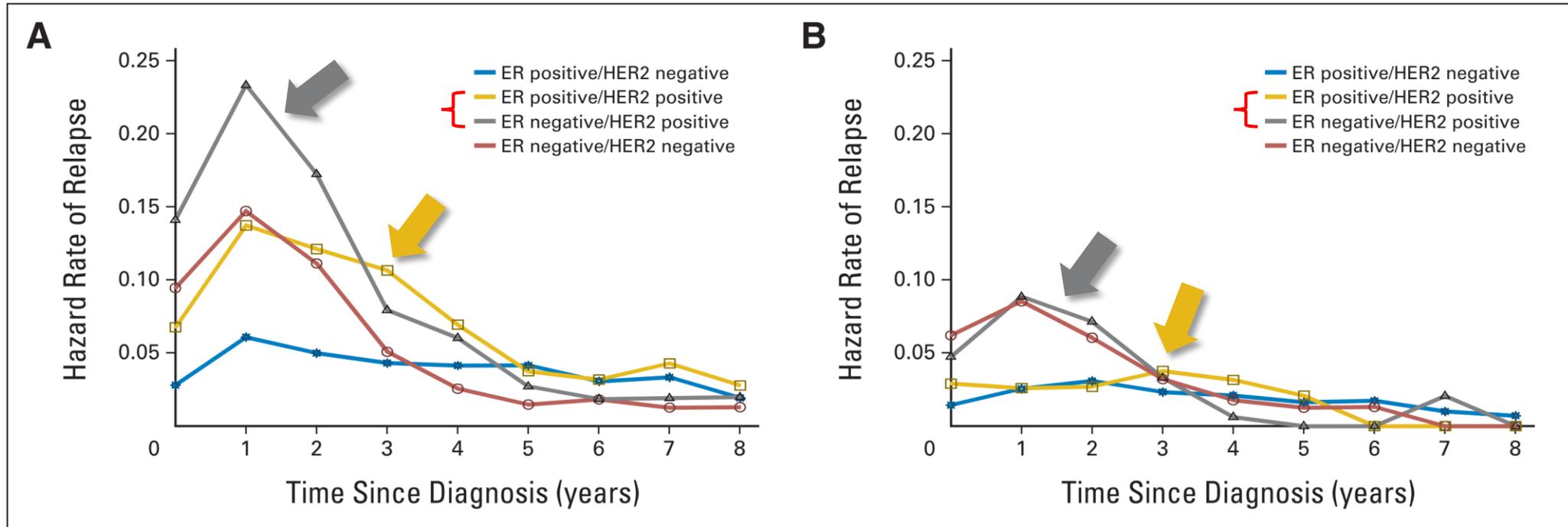
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HER2-positive breast cancer

- 1980s: **discovery** of the oncogene Human Epidermal growth factor Receptor 2 (HER2)
- 1987: **amplification** of HER2 in BC is associated with *poor prognosis*
- Development of the anti-HER2 monoclonal antibody **trastuzumab**
- 1998: FDA approved trastuzumab for **metastatic** HER2+ BC
- 2005: Results of **adjuvant** trastuzumab trials
- 2013: FDA approved trastuzumab + pertuzumab + docetaxel as **neoadjuvant** treatment



Prognosis *before* and *after* trastuzumab

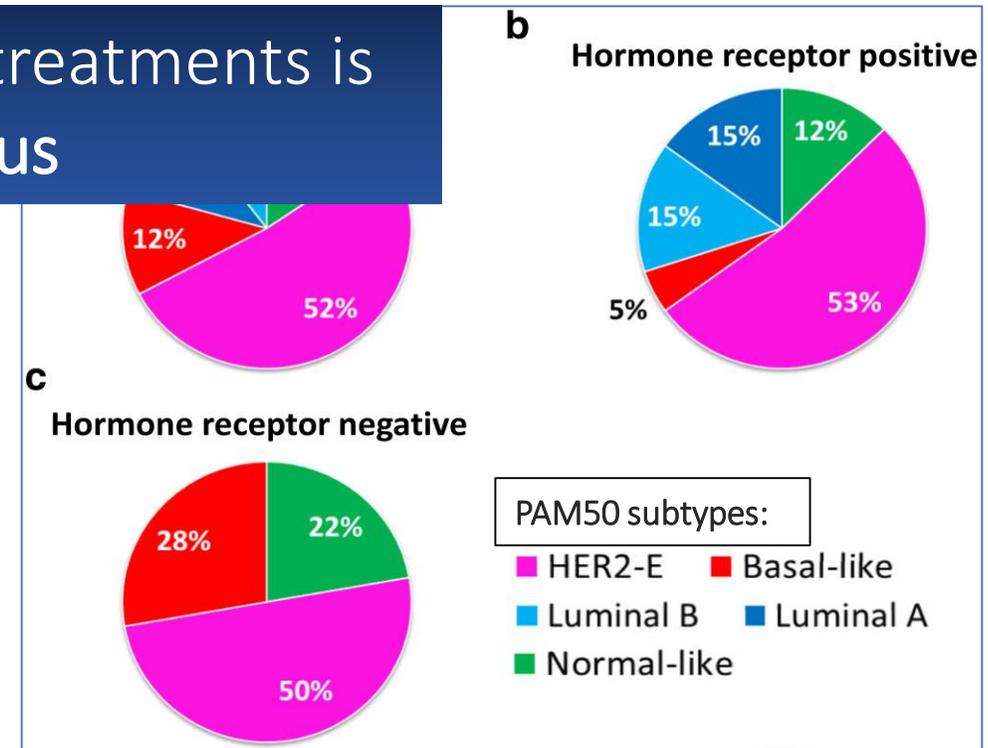


Hazard rate of relapse according to tumor subtype in (A) cohort 1 (1986-1992) and (B) cohort 2 (2004-2008)

HER2+ BC *heterogeneity*

- Histopathology (ductal vs lobular)
- HER2 positivity (3+ vs 2+ with FISH amplified) and HER2 IHC expression levels
- Hormone receptor status
- PAM50 intrinsic subtypes¹
- Mutational/copy number/epigenetic profiles^{2,3}
- Tumor-Infiltrating Lymphocytes (TIL) levels⁴
- ...

Response to anti-HER2 treatments is heterogeneous



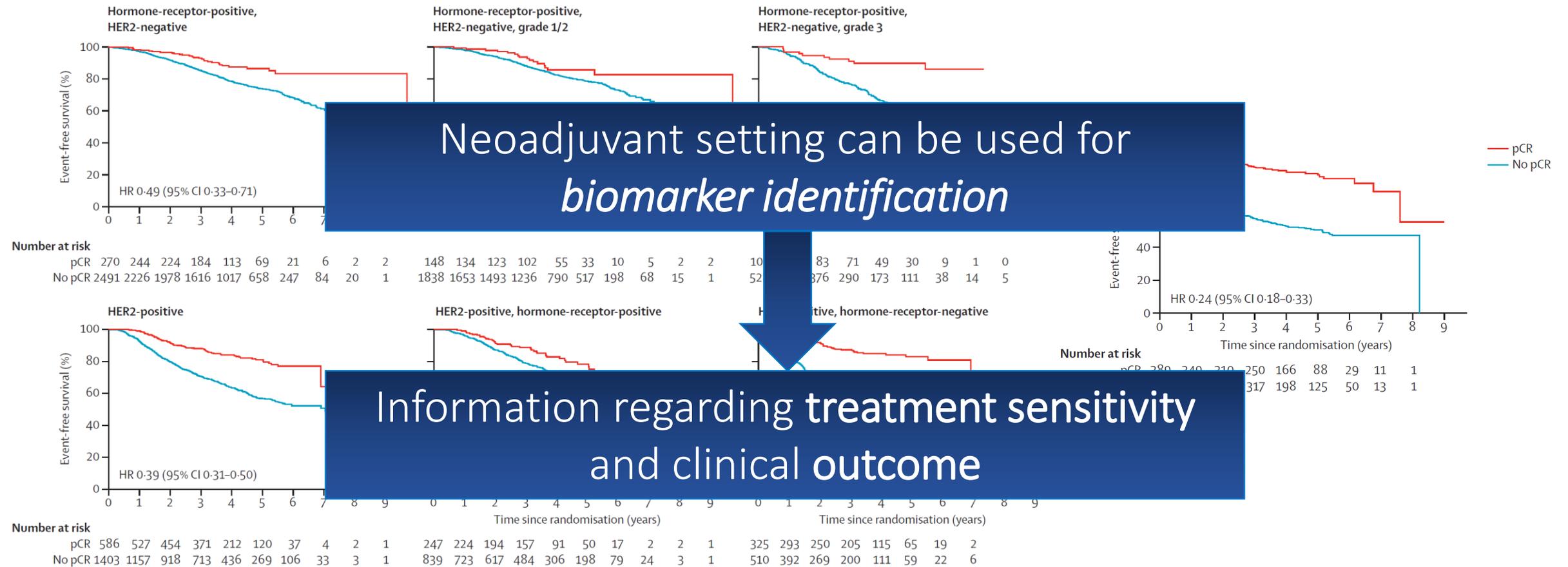
¹ Gavilà J, et al. BMC Med 2019

² Cancer Genome atlas network, Nature 2012

³ Pereira B, et al. Nat Commun 2016

⁴ Solinas C, et al. Breast 2017

Neoadjuvant therapy – association between *pCR* and *long-term outcome*



pCR = pathological complete response

The NeoALTTO trial

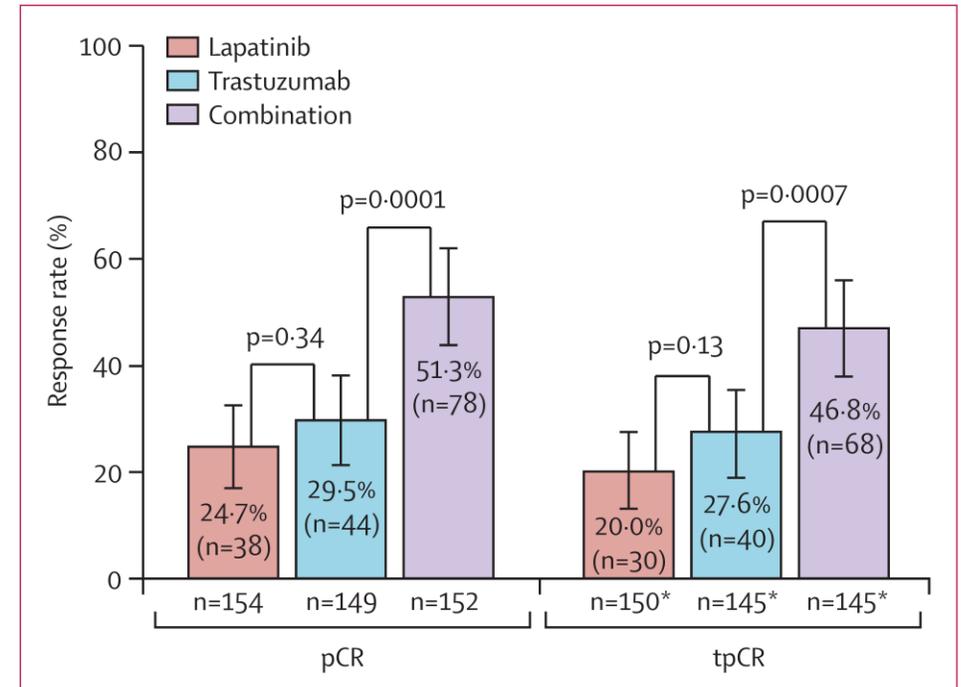
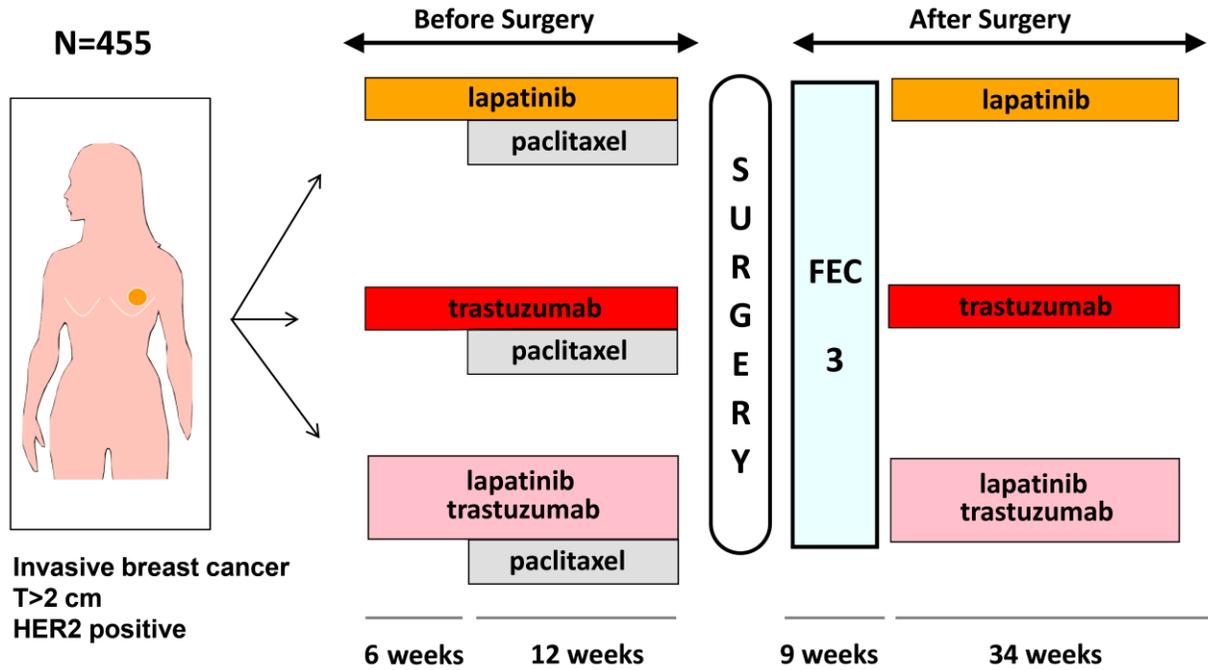
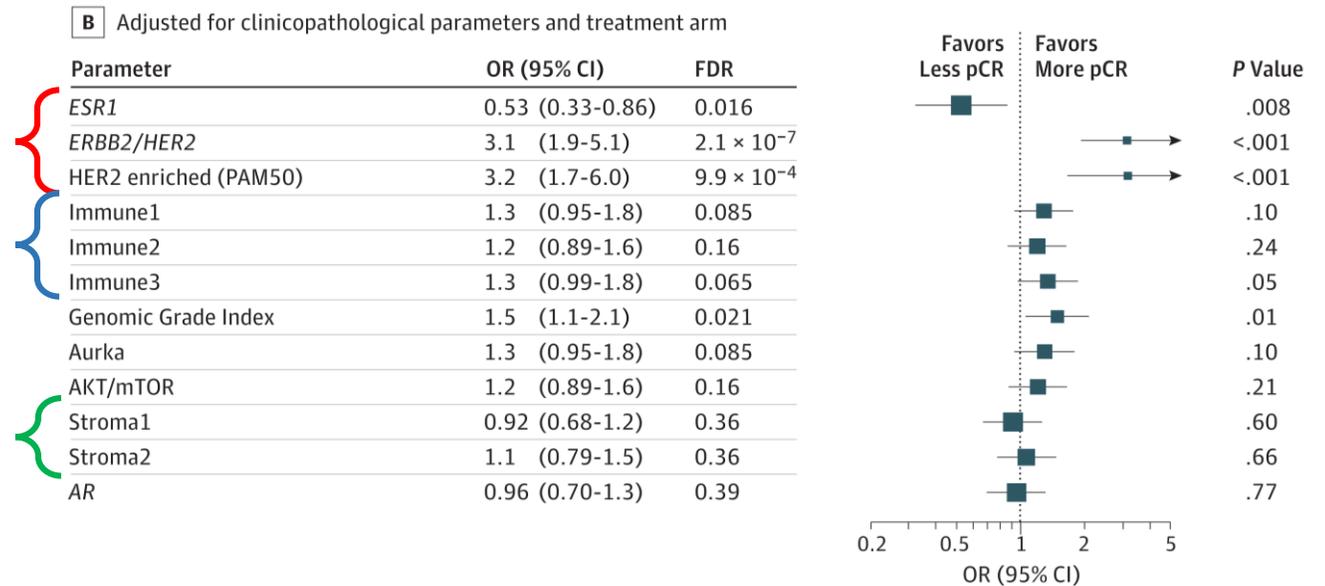


Figure 2: Rates of pCR and of locoregional total pCR in the three treatment groups

RNA-sequencing to predict pCR in the NeoALTTO trial

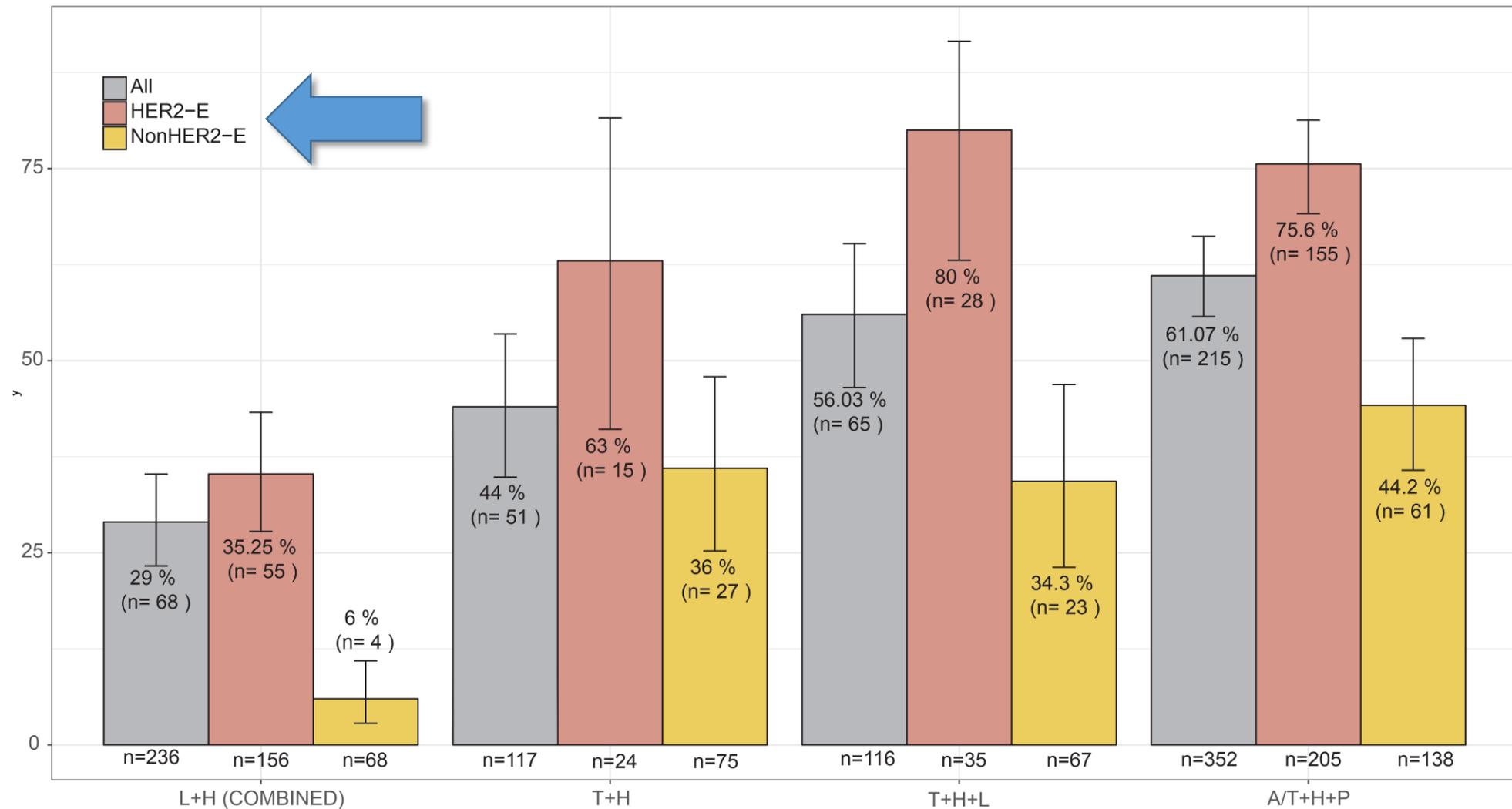
- RNA-sequencing to determine **gene expression levels, PAM50 subtypes, gene signatures (GSs)***
- In **all treatment arms**:
 - High expression levels of **ERBB2/HER2**: \uparrow pCR
 - Low levels of **ESR1**: \uparrow pCR
 - **HER2-enriched** PAM50 subtype: \uparrow pCR
- In the **combination arm**:
 - High expression of **immune GSs**: \uparrow pCR
 - High expression of **stroma GSs**: \downarrow pCR



Effect of single genes and gene expression signatures on pCR adjusting for clinicopathological parameters and treatment arms.

- * **Gene expression signature**: group of genes with expression pattern characterizing biological processes
 \rightarrow pathway activation, prognostic/predictive biomarkers, gene sets associated to specific function, disease subgroups

pCR rates according to *HER2-E PAM50 subtype*

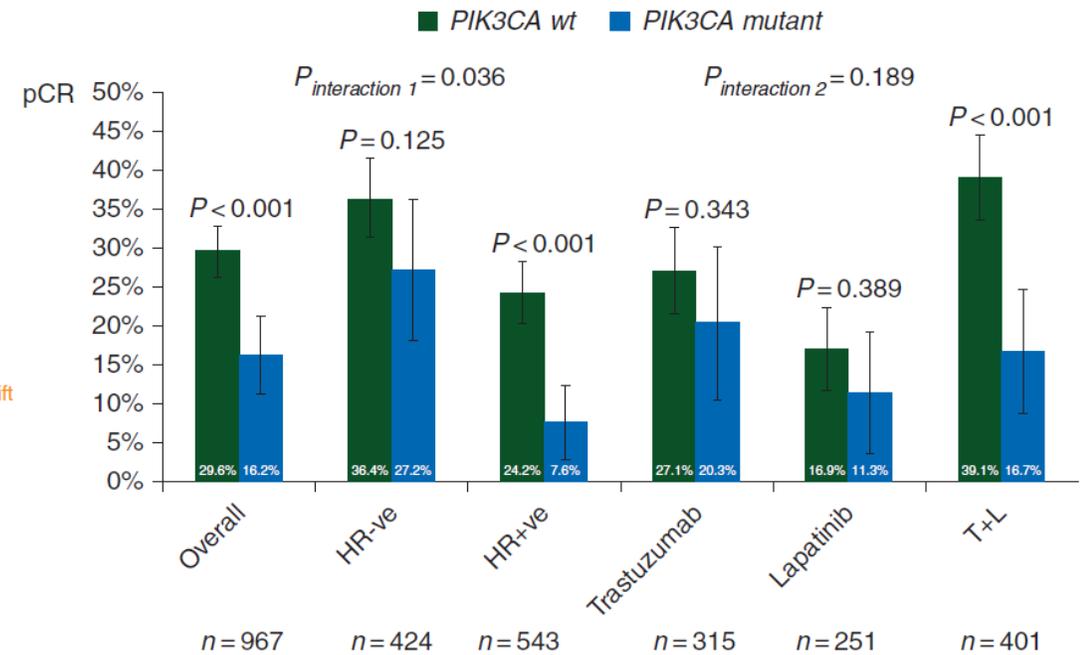
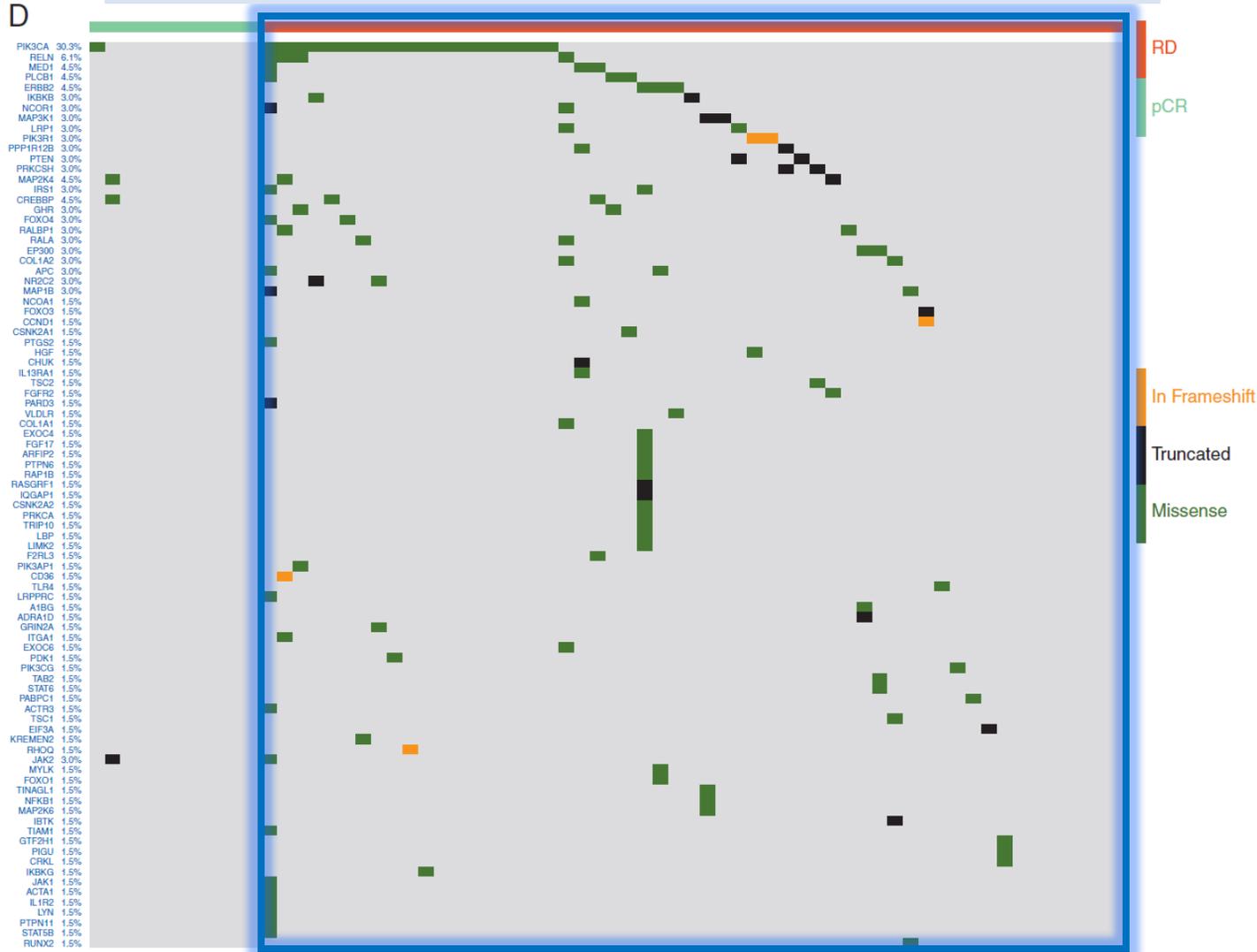


Rates of pCR according to the type of chemotherapy and anti-HER2 therapy using data from 8 neoadjuvant clinical trials in HER2+ breast cancer
 H = trastuzumab; L = lapatinib; P = pertuzumab; T = taxane; A = anthracycline

PIK3CA mutations to predict pCR

NeoALTTO trial → mutations in PIK3CA network genes

Pooled analysis from 5 prospective trials



Triple-negative breast cancer

BL1: Basal-like 1

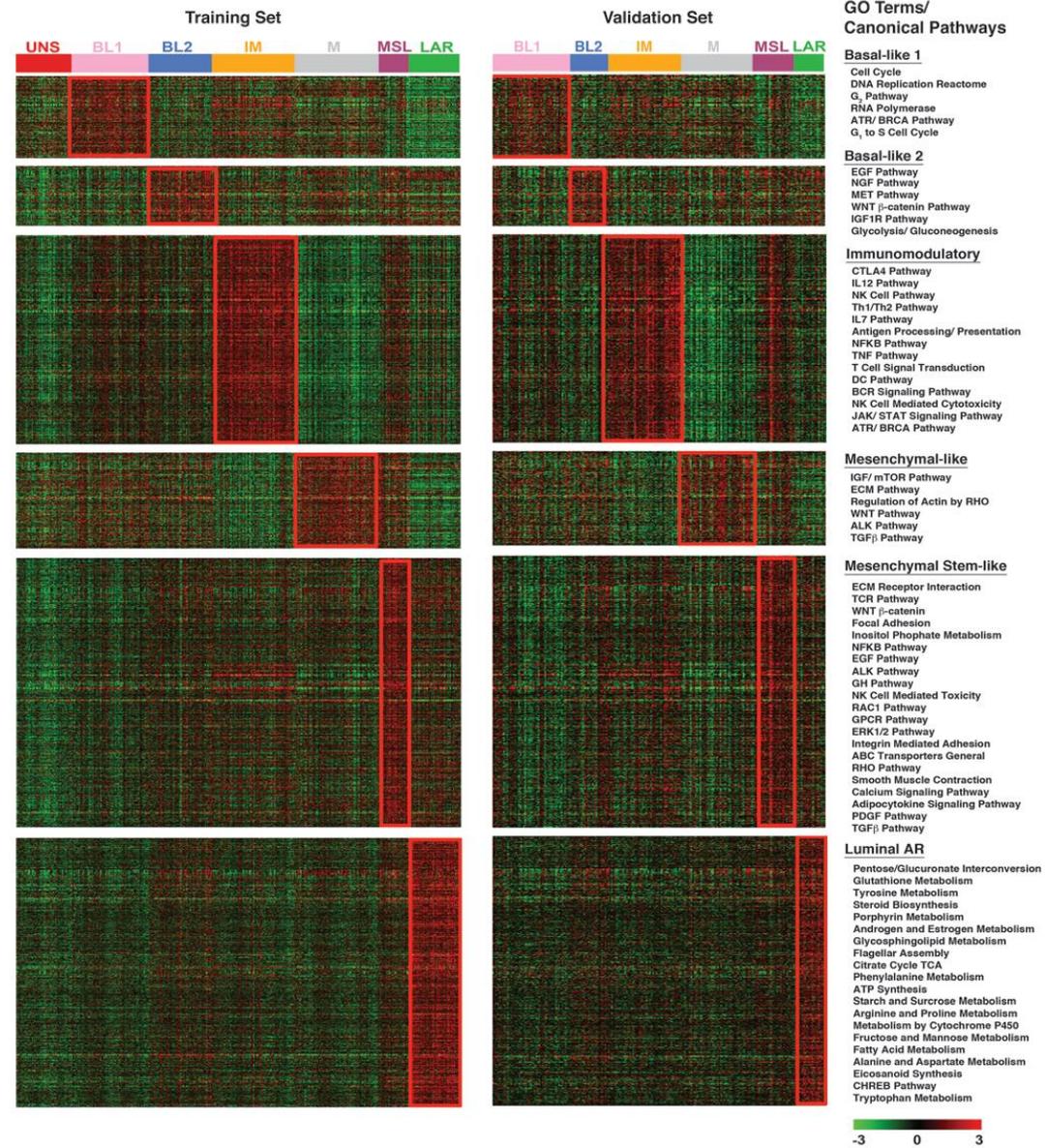
BL2: Basal-like 2

IM: Immunomodulatory

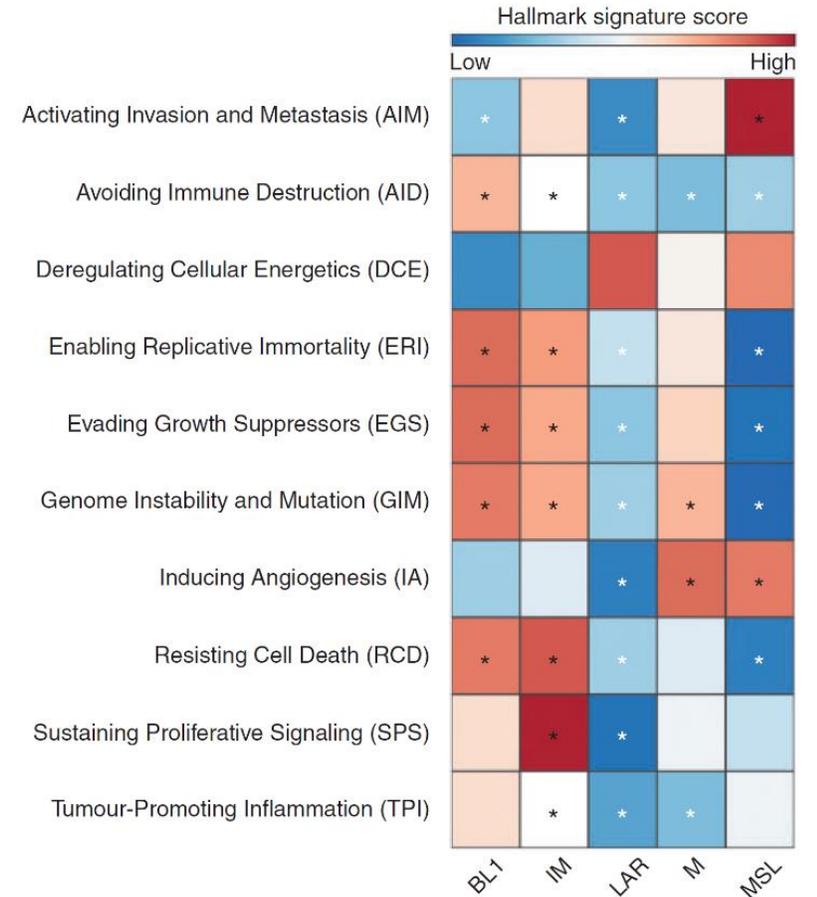
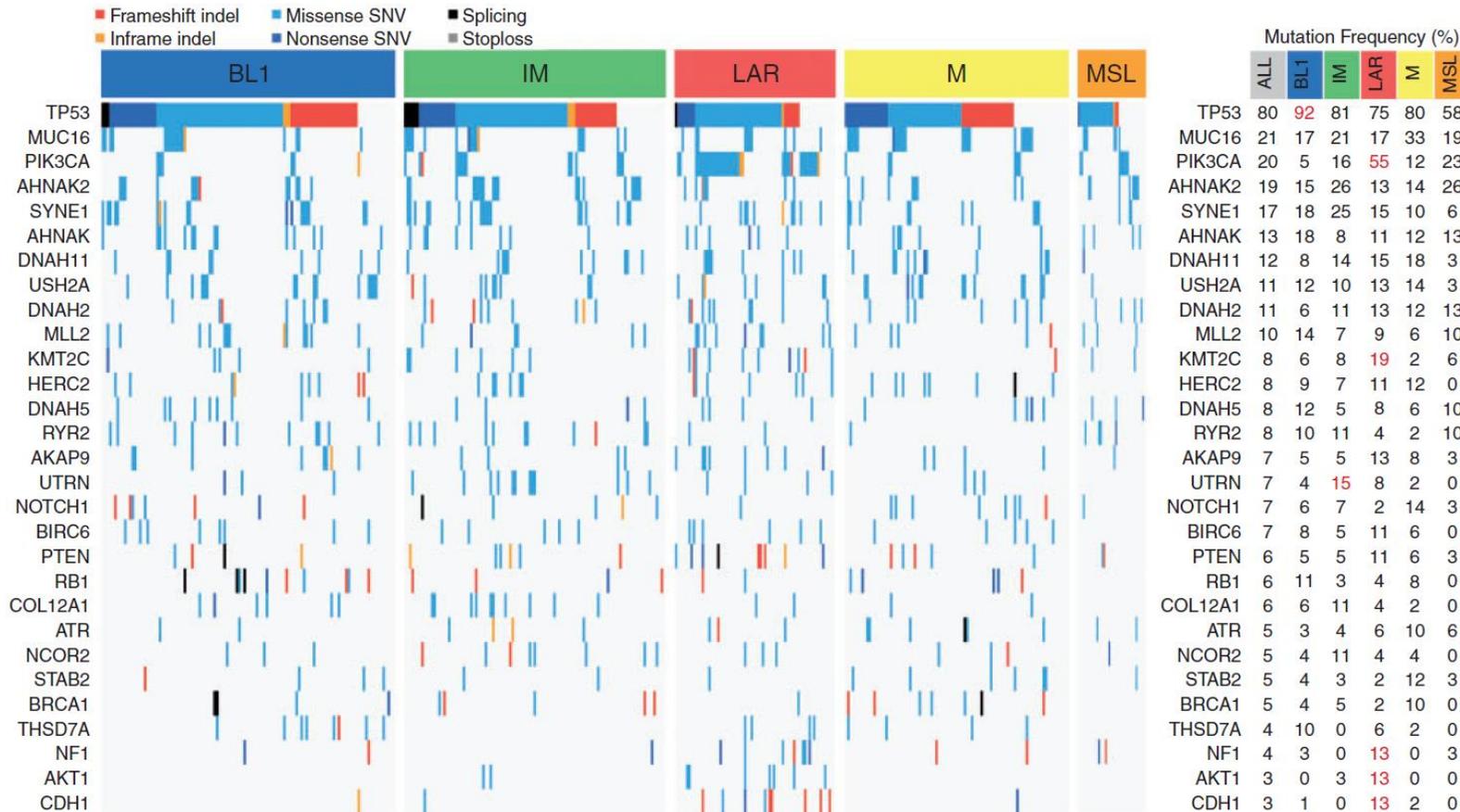
M: Mesenchymal

MSL: Mesenchymal Stem-like

LAR: Luminal Androgen receptor

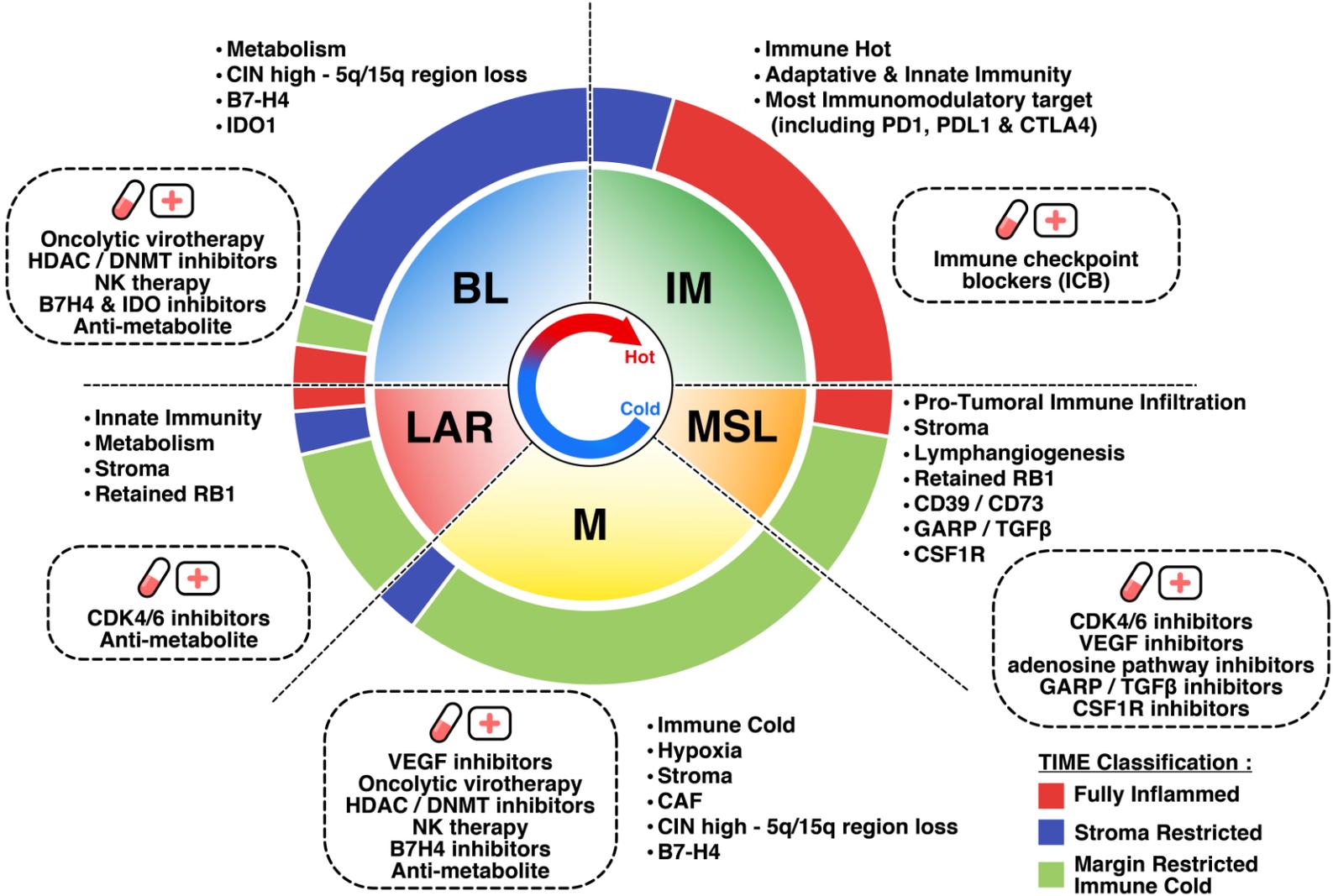


TNBC subtypes – multi-omic analysis



Significant **up-regulation** displayed in black, **down-regulation** in white

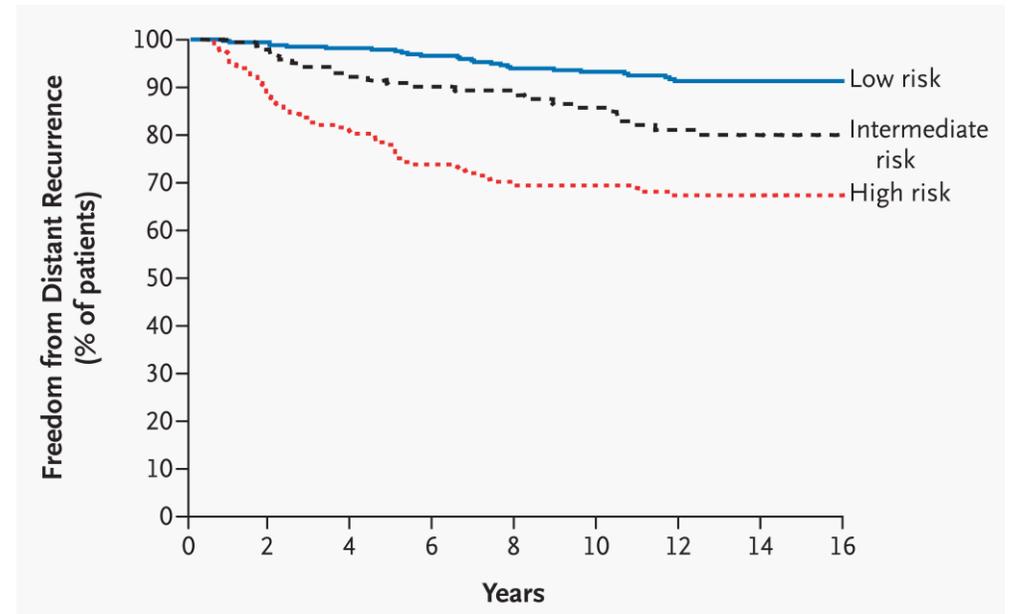
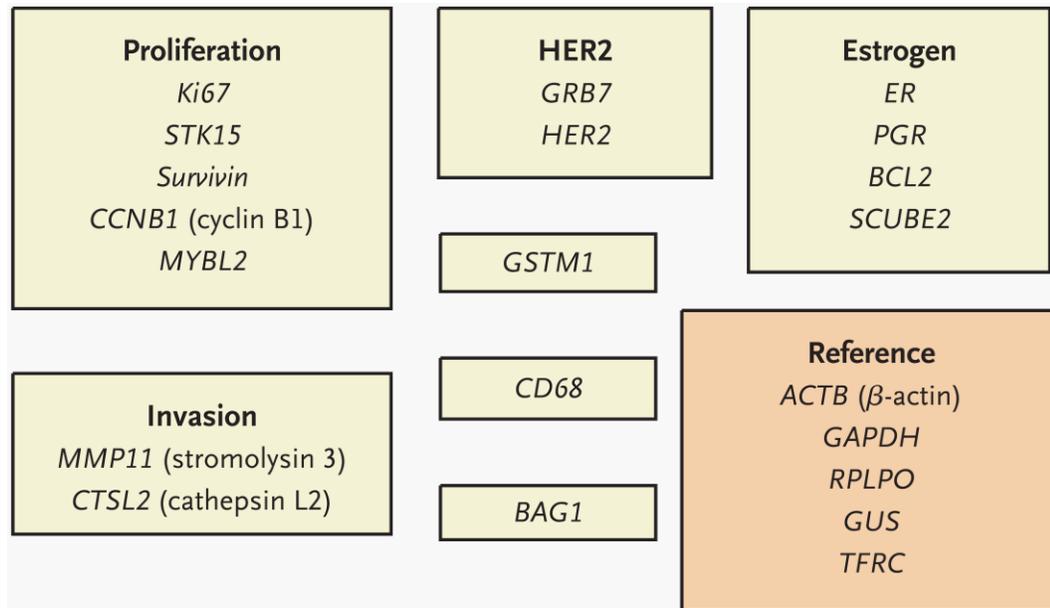
Heterogeneity of TNBC – opportunities for personalized treatment



Luminal breast cancer

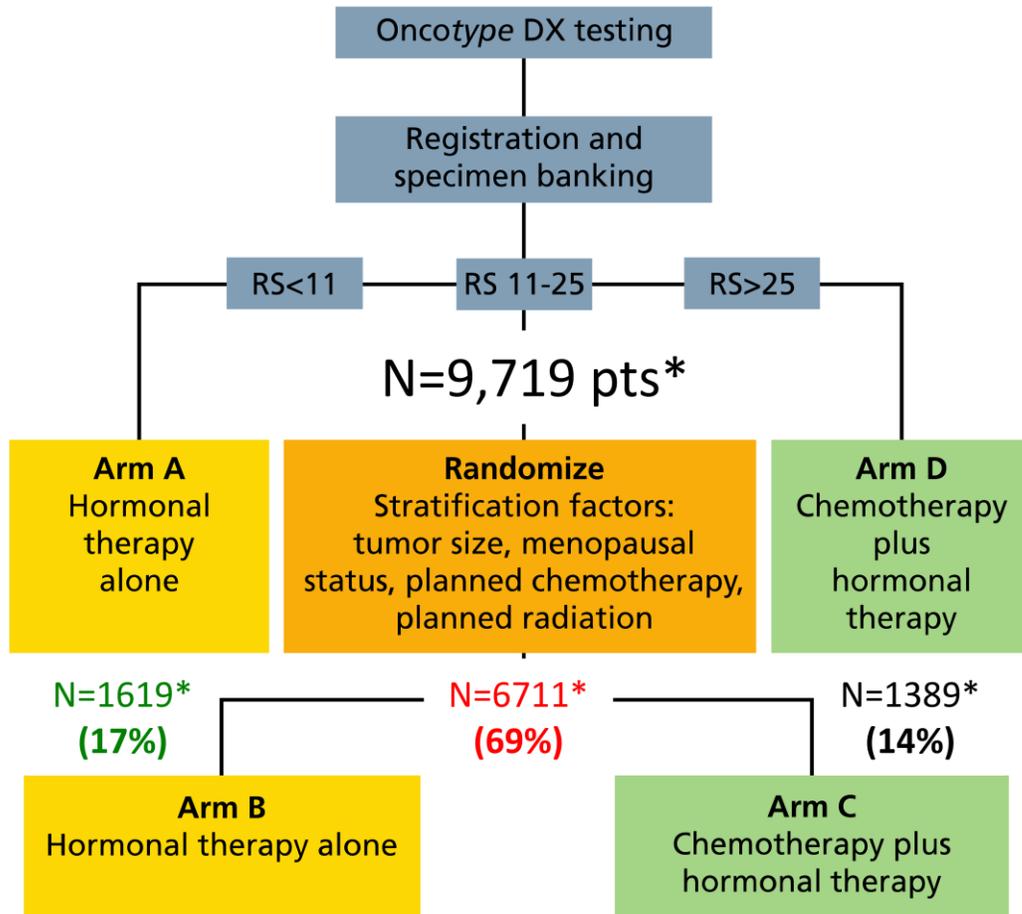
Oncotype DX assay: 21 genes selected after three independent preliminary studies involving 447 patients and 250 candidate genes

Range of recurrence scores from 0 to 100 based on *gene expression levels*

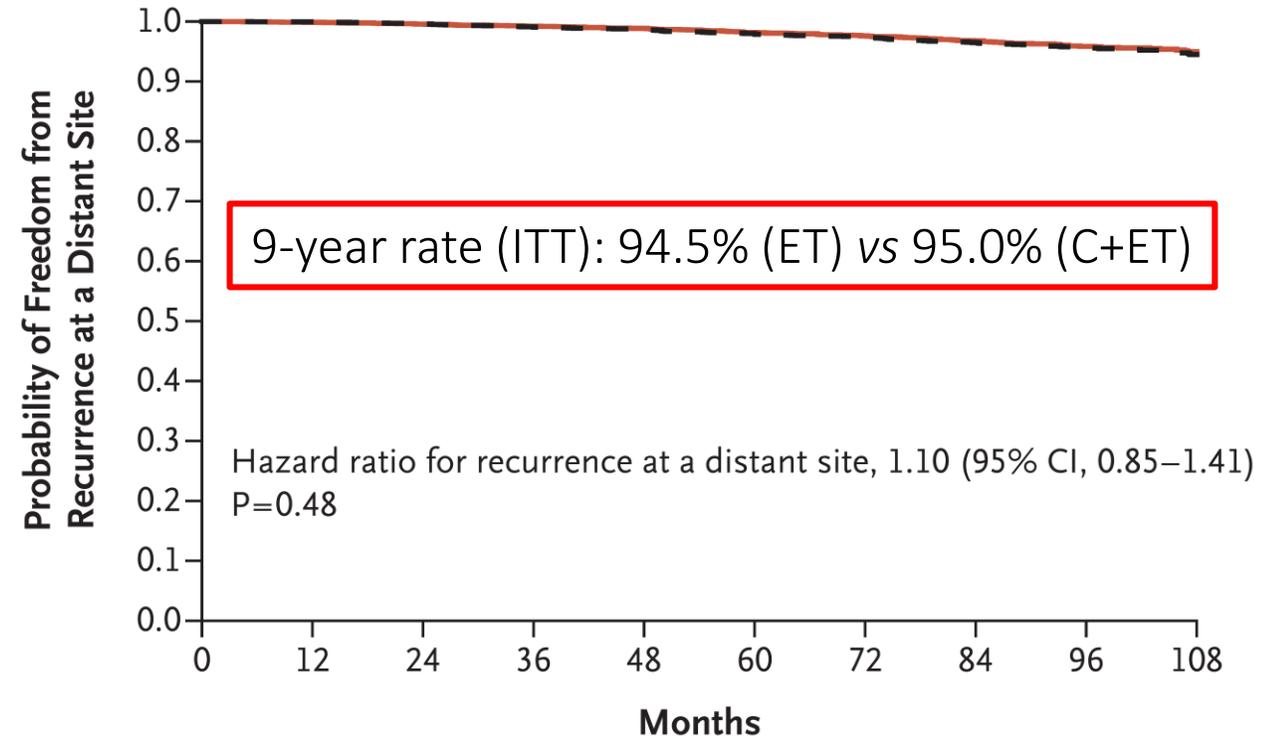


Likelihood of distant recurrence, according to Recurrence-Score Categories

The TAILORx trial – adjuvant setting



* Included in the main analysis (eligible patients with FU information)



Some **benefit of chemotherapy** found in women **50 years of age or younger** with a recurrence score of **16 to 25**

Table 1 | Summary of studies used in testing of different gene signatures

Gene-signature test	Training set	Initial validation set	Proportion (%) of patients assigned to the 'low-risk' category	Clinical application
Oncotype DX ²⁵	447 ER+/- tumour samples from patients with LN+/- disease enrolled in three separate clinical trials, including from the tamoxifen only arm of NSABP B-20	668 ER+ and tumour samples from patients with LN- disease in the tamoxifen only arm of NSABP B-14 (including samples from the training set)	51.0	Prediction of 10-year recurrence risk in patients with ER+ and LN- disease
Prosigna ^{37,41}	189 ER+/- tumour samples from patients with LN+/- disease and 29 nonmalignant breast tissue biopsy samples	786 ER+/- tumour samples from patients with and LN+/- disease	28.2	Determining the prognosis of postmenopausal women with ER+ and LN+/- disease of stages 1 or 2
MammaPrint ^{43,44}	78 ER+/- tumour samples with a diameter <5 cm from patients <55 years of age with LN-negative disease	295 ER+/- tumour samples <5 cm in diameter from patients <53 years of age with and LN+/- disease (including samples from the training set)	40.0	Determining the prognosis of women with ER+/- and LN- disease of stages 1 or 2
Breast cancer index ^{59,60}	60 ER+ tumour samples from patients previously treated with tamoxifen	588 ER+ tumour samples from patients with LN- disease enrolled in the Stockholm trial	53.0	Determining the prognosis of women with ER+ and LN- disease, prediction of benefit from extended endocrine therapy
EndoPredict ⁶⁵	964 ER+ tumour samples from patients with LN+/- disease treated with tamoxifen	378 ER+ tumour samples from patients with LN+/- disease from the ABCSG-6 trial (tamoxifen-only arm) and 1,324 patients from the ABCSG-8 trial	62.6	Determining the prognosis of women with ER+ and LN+/- disease
Genomic Grade Index ⁷⁴	64 ER+ tumour samples of histological grades 1-3	<ul style="list-style-type: none"> • 125 ER+/- • Tumour samples of histological grades 1-3 from patients with and LN- disease 	59.7	Prognosis and risk stratification based on histological grade

ER: oestrogen receptor; LN: lymph node

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Tumor *evolution* and *heterogeneity*

Experimental data in breast cancer → *Mixed model*

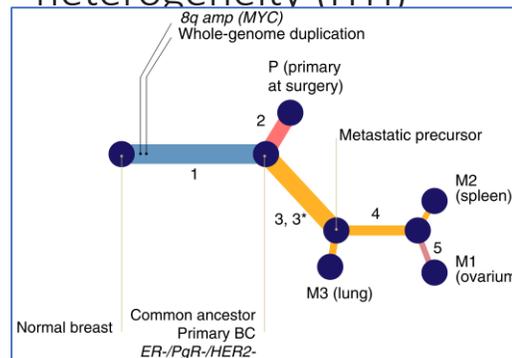
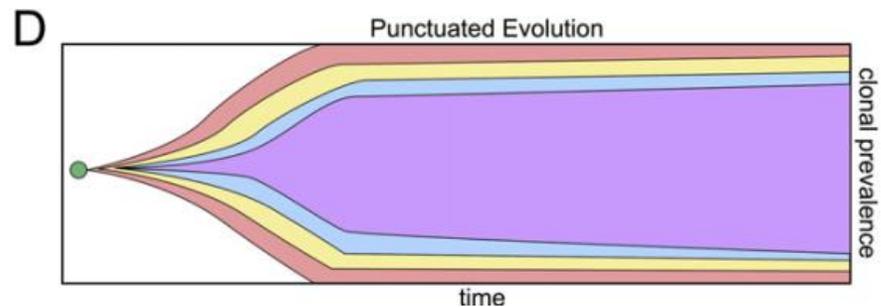
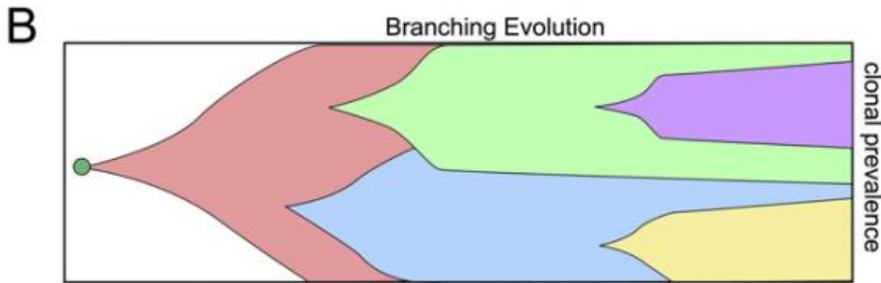
- CNAs and chromosomal structural variants follow a *PE* model (D)

- They occur in *early* punctuated bursts of evolution, and stably expand

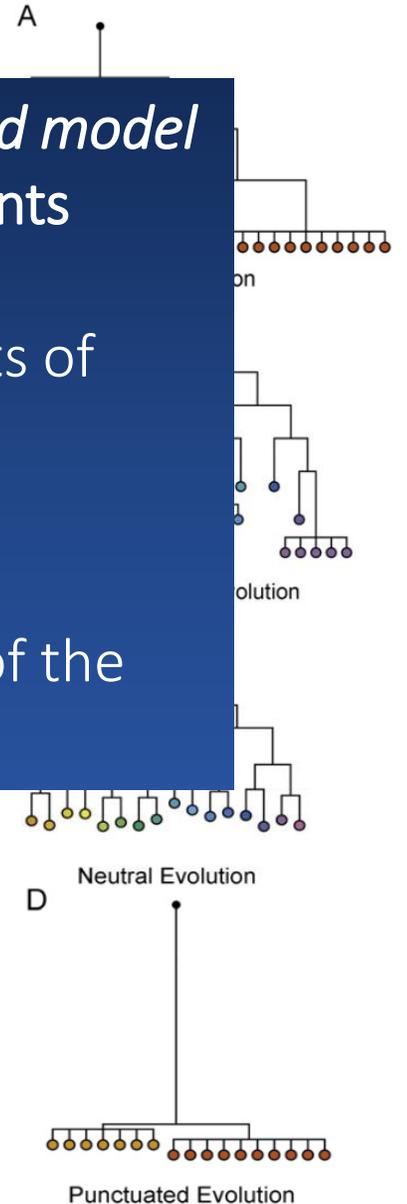
- Point mutations follow a *BE* model (B)

- *Gradual* evolution over the lifetime of the tumor, leading to clonal expansions

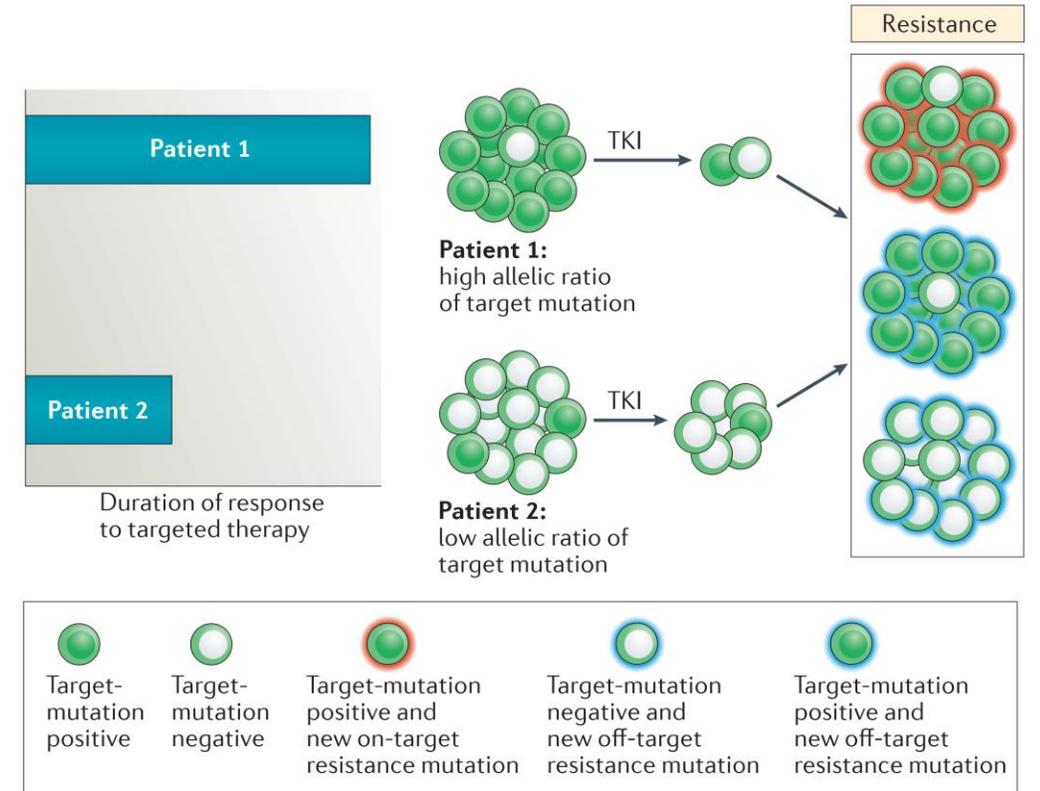
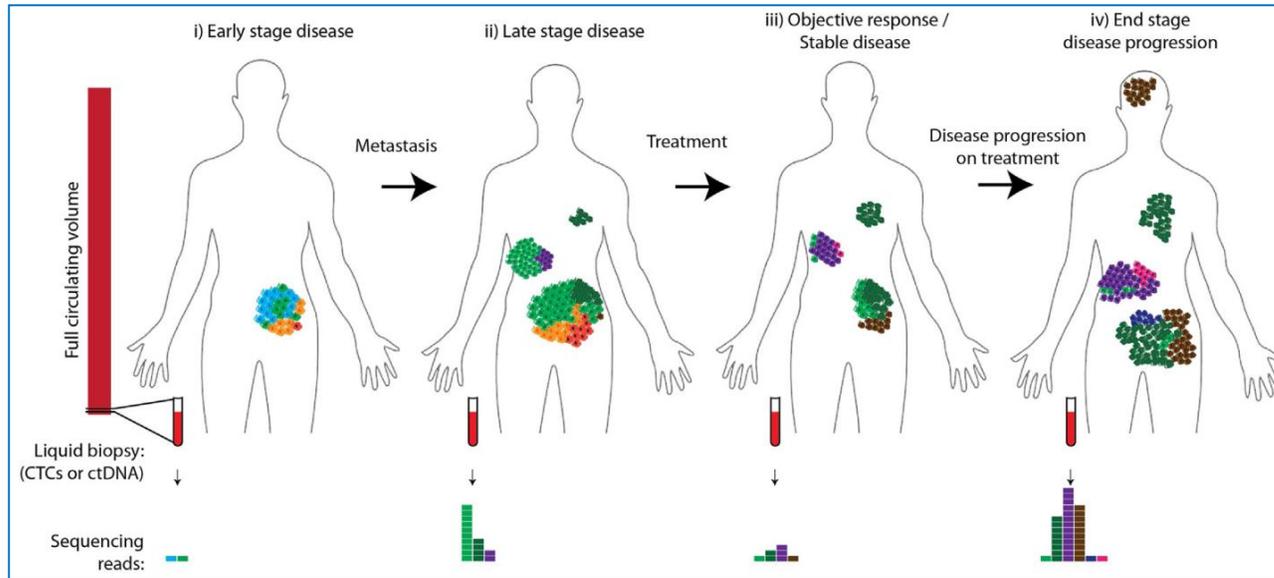
leading to genetic drift and extensive intratumor heterogeneity (ITH)



genomic aberrations
Phylogenetic analysis performed
using somatic mutations and
a copy number aberrations



Tumor heterogeneity *increases* over time and is correlated to *treatment resistance*

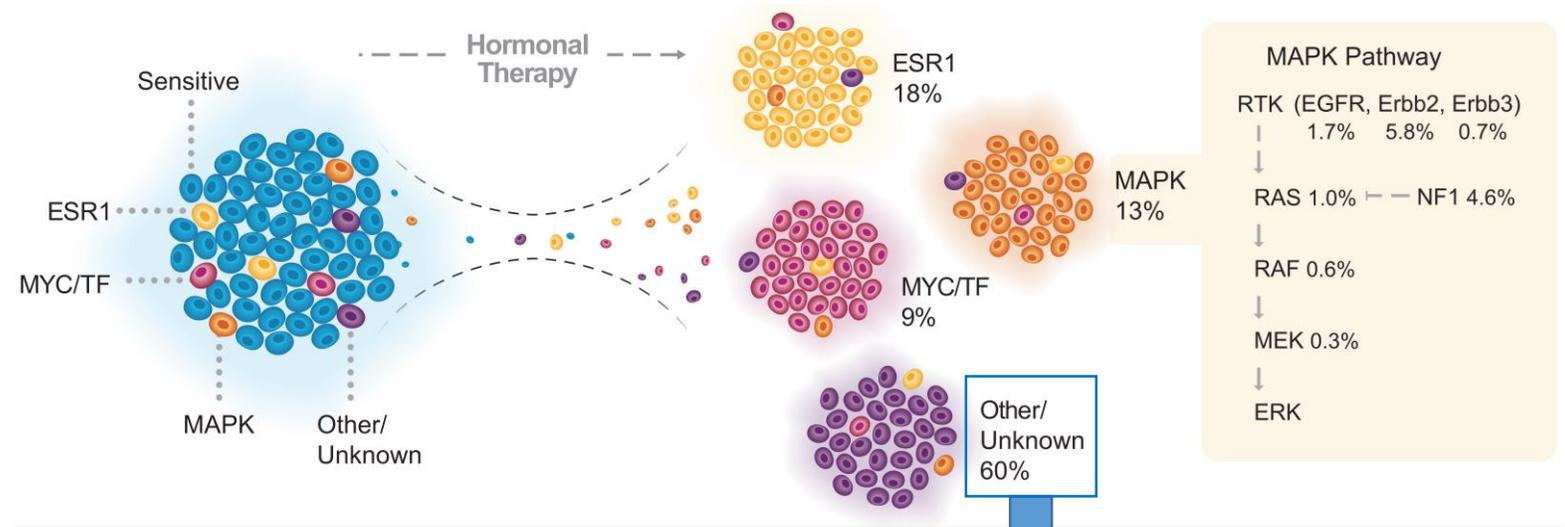


Taxonomy of the mechanisms of *resistance* to *endocrine therapy*

Highlights

- We performed prospective sequencing of 1,501 HR⁺ breast cancers in the clinical setting
- MAPK and TF alterations were present in 22% of 692 HR⁺ post-endocrine therapy tumors
- MAPK and TF alterations were mutually exclusive with *ESR1* mutations
- MAPK and TF alterations were associated with shorter response to endocrine therapies

TF = transcription factor

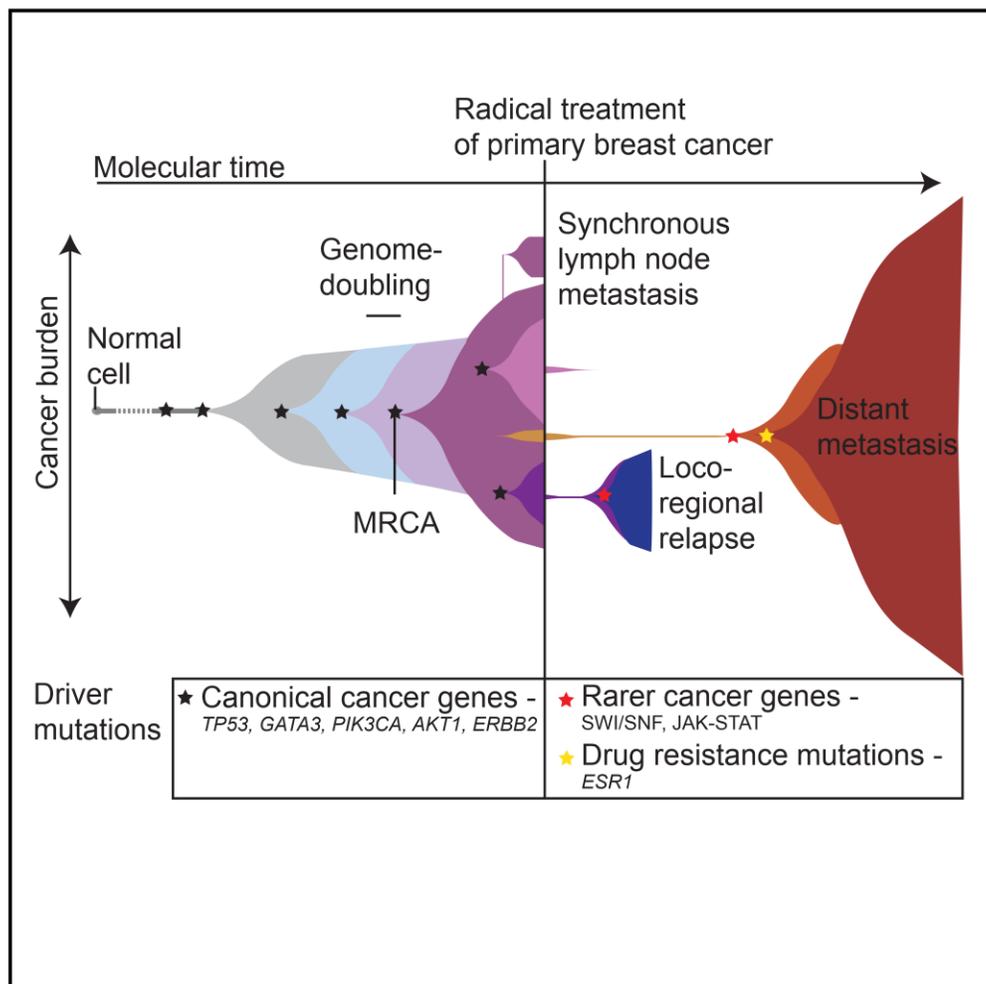


Genomics alterations associated to treatment **resistance** may

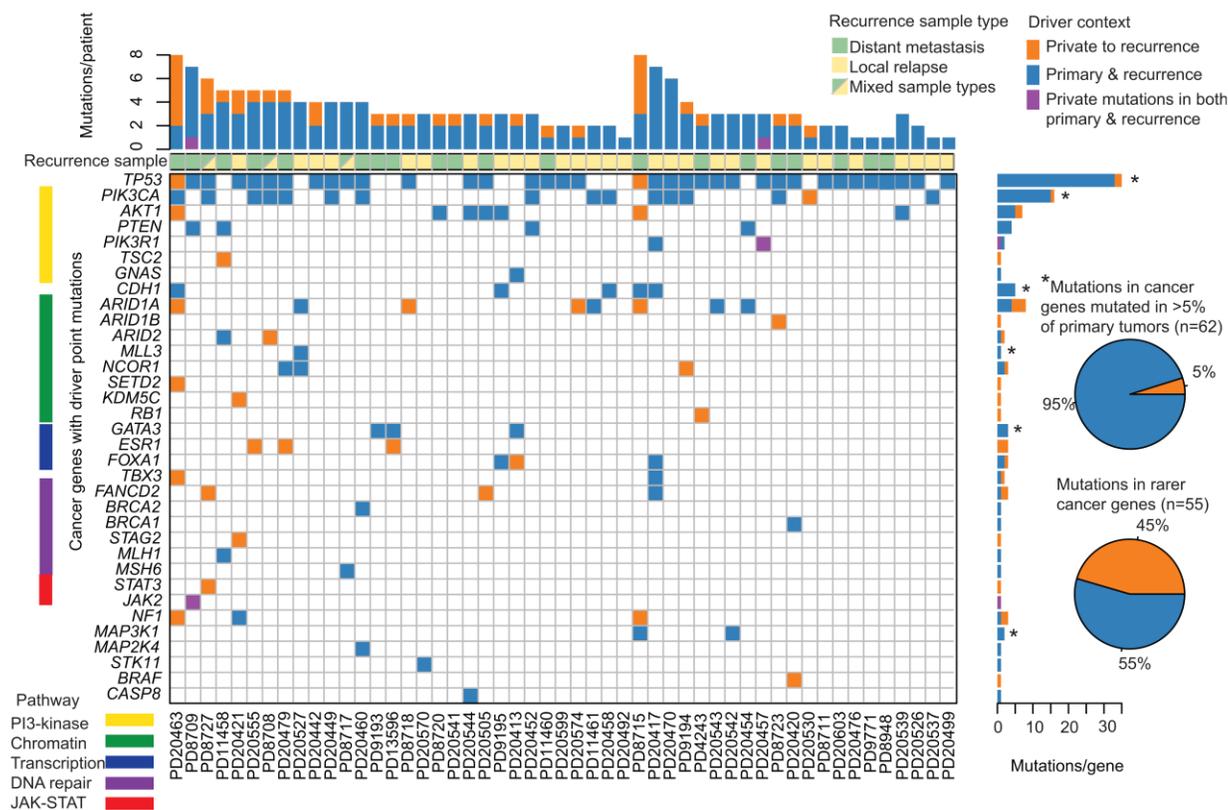
- *pre-exist* in the pre-treatment tumors and expand
- be *acquired* under the **selective pressure** of endocrine therapy

Role of transcriptional reprogramming, epigenetics, tumor microenvironment...?
→ Go beyond “single gene” vision

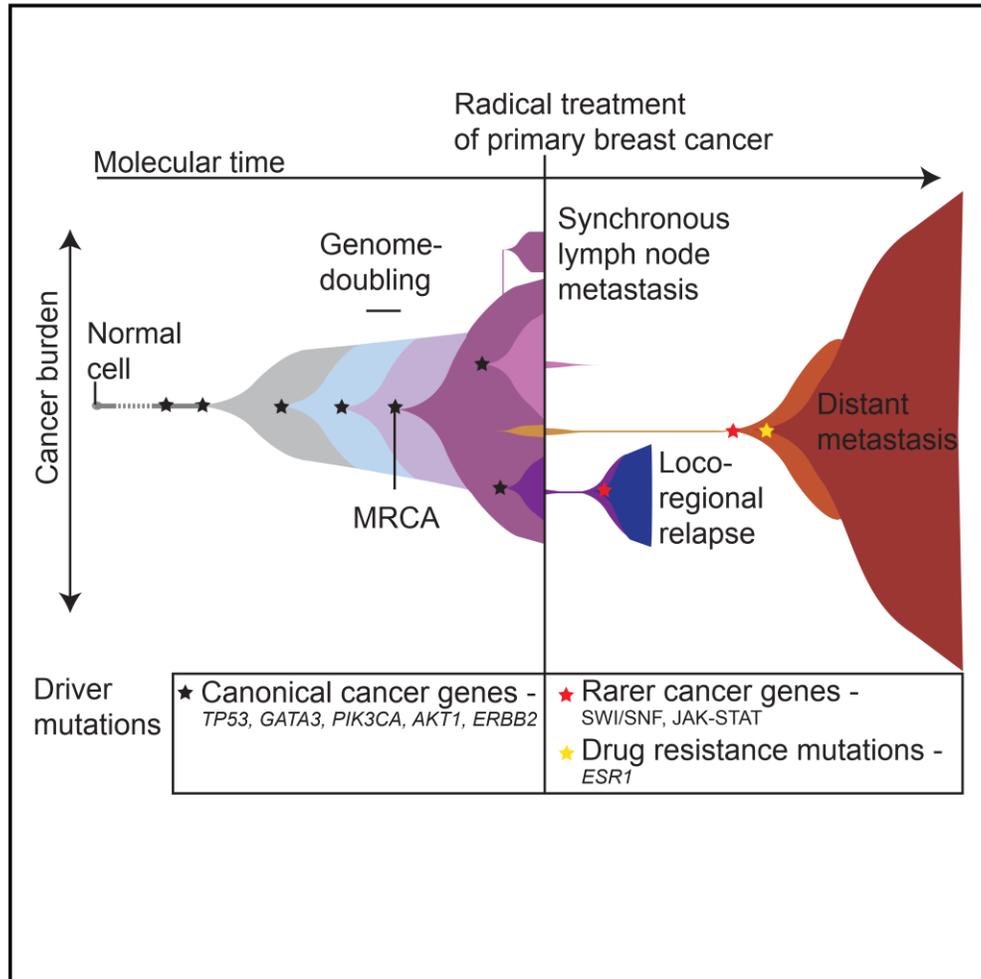
Progression of breast cancer – when?



Metastases mostly *disseminate late* from primary breast tumors, keeping most drivers, but **continue to acquire mutations**

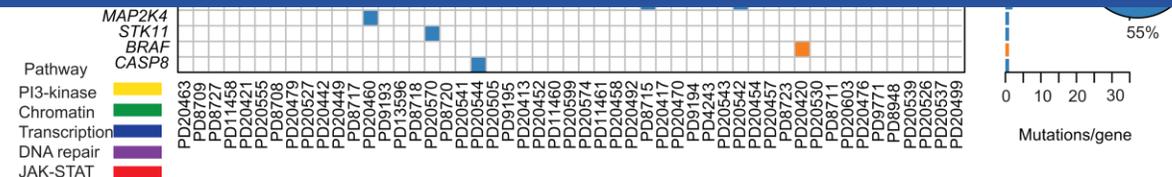


Progression of breast cancer – when?



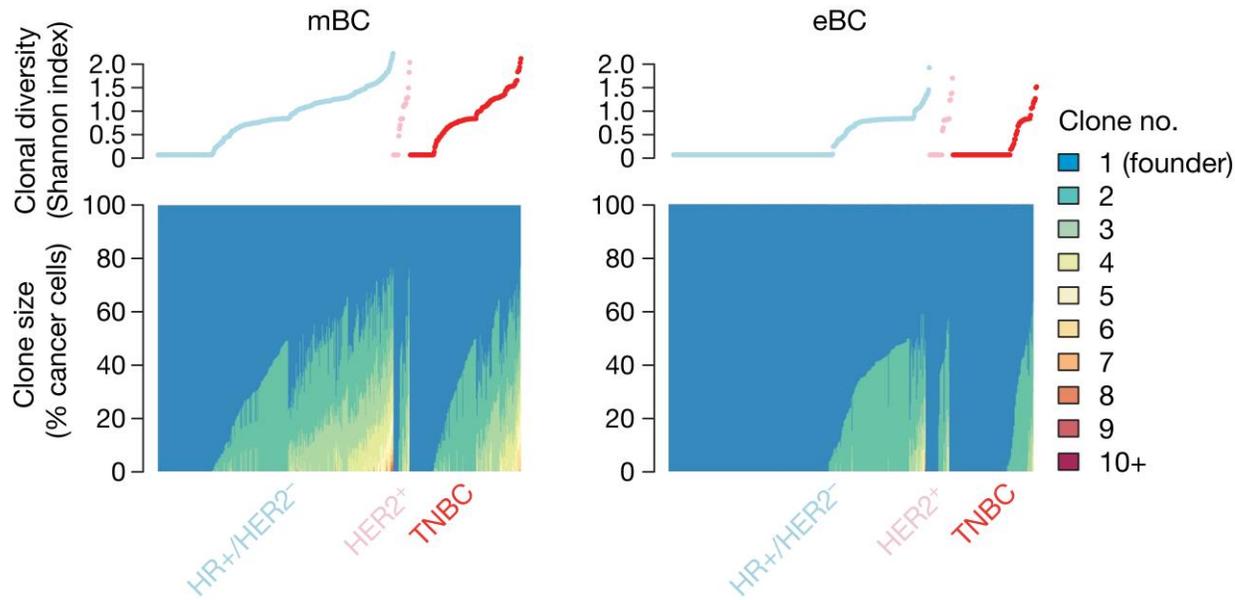
Metastases mostly *disseminate late* from primary breast tumors, keeping most drivers, but **continue to acquire mutations**

- The genome of the **primary** tumor represents a good *proxy* for that of the cells that ultimately seeded the relapse
→ Important for **adjuvant treatments**
- The genome of a **metastatic clone** undergoes extended **changes** by the time it has expanded to be clinically detectable

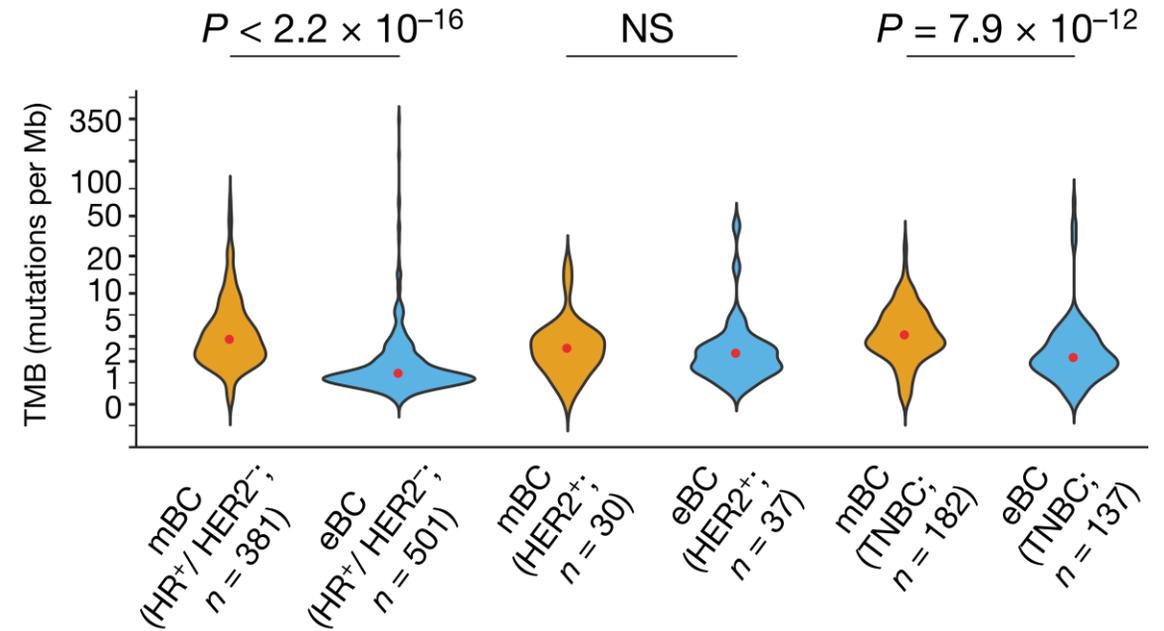


Advanced-stage BCs are *more complex* than early-stage BCs

Increase in the *clonal diversity* in mBC



Increase of *mutational load* in mBC



eBC = early-stage breast cancer
mBC = metastatic breast cancer
TMB = tumor mutational burden

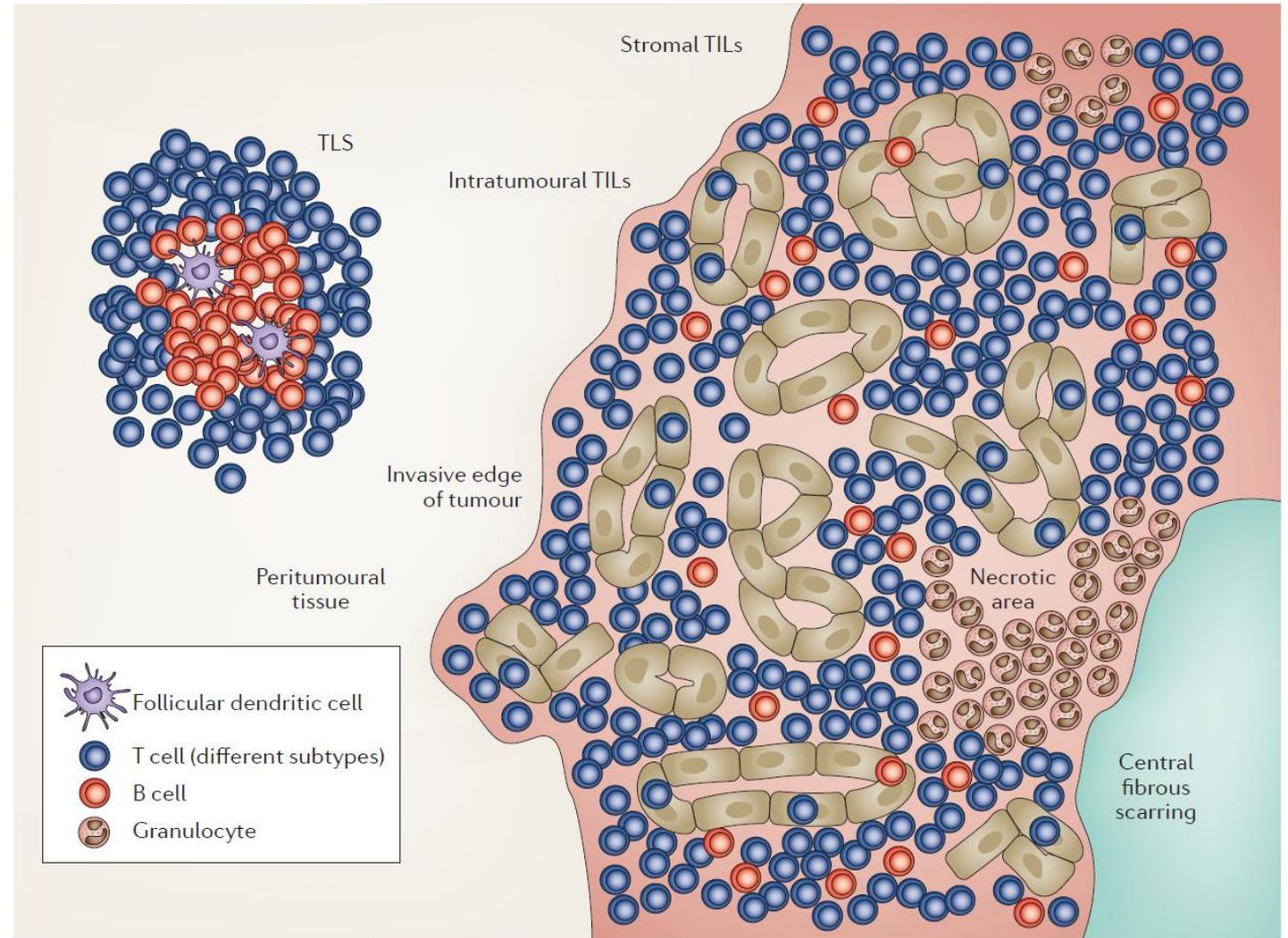
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The *immune* microenvironment

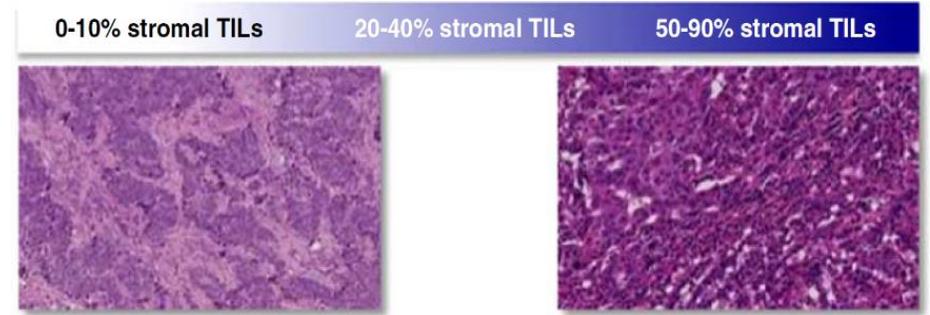
Characterization of the tumor immune microenvironment can be performed at different levels

- **Quantification** of tumor-infiltrating lymphocytes (TILs, e.g. H&E staining)
- Characterization of TIL **subpopulations** (e.g. IHC, IF, flow cytometry)
- Description of the TIL **geographic distribution**



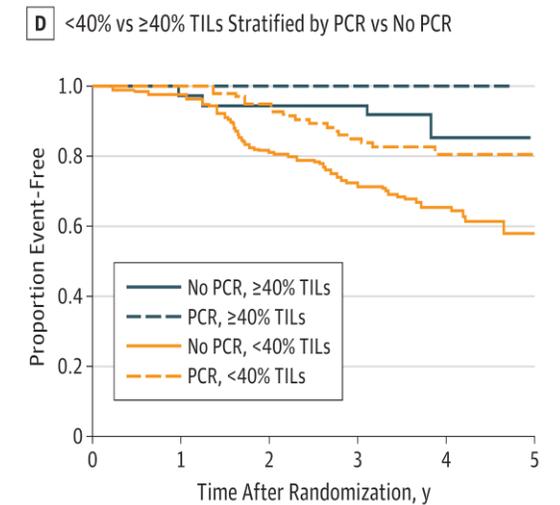
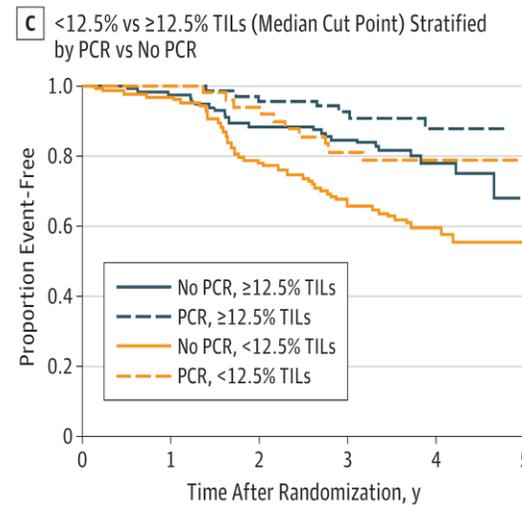
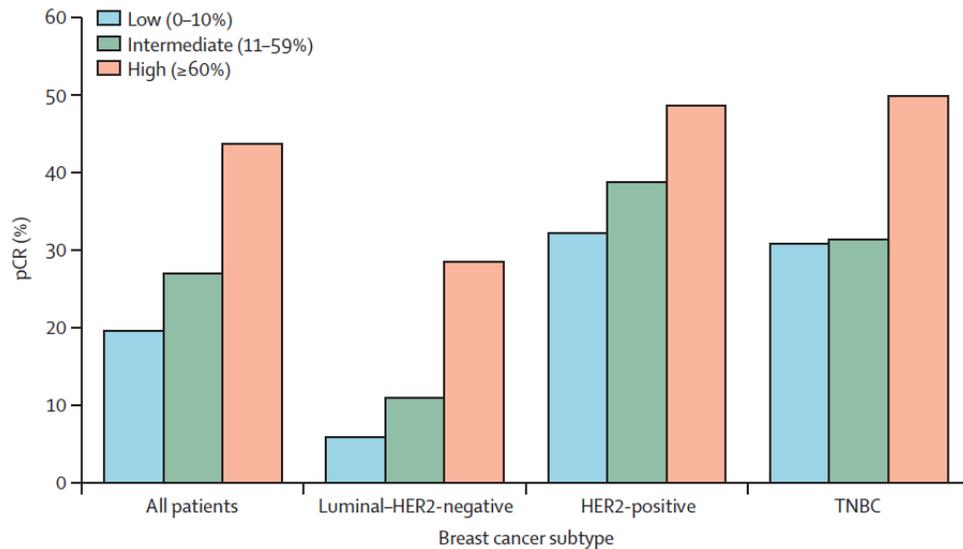
TIL levels as a biomarker

Current clinical data establish the clinical validity of higher TIL levels as a *predictive* and *prognostic biomarker*



Pooled analysis of 3771 BC patients treated with **neoadjuvant** therapy

Higher TIL levels → better **Event-Free Survival** independently of pCR in NeoALTTO



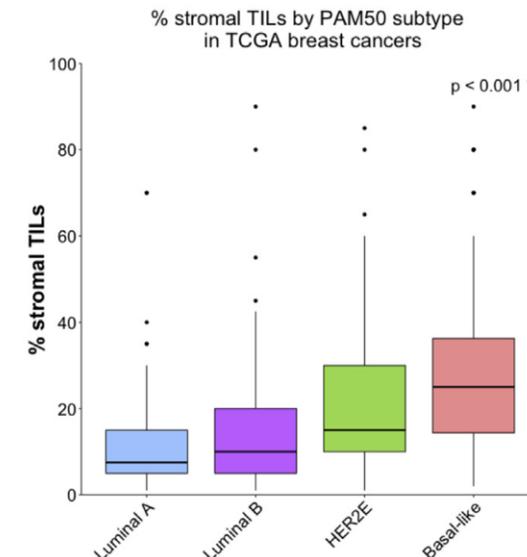
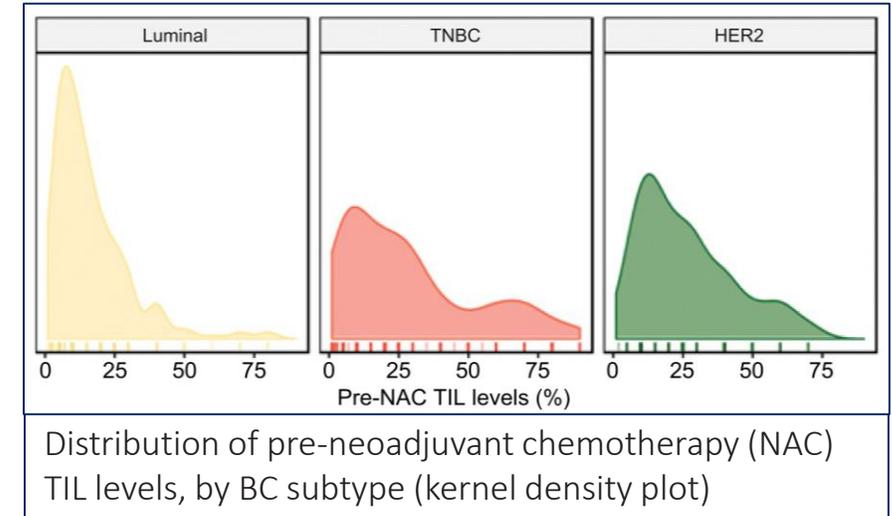
TIL levels in breast cancer subtypes

Median *levels of stromal TILs* (scored following international guidelines¹, usually higher than intratumoral TILs):

- Luminal BC → 7-10%
- HER2-positive BC → 15-20%
- TNBC → 15-20%

HER2-positive and triple-negative BCs are considered more *immunogenic* than luminal BC

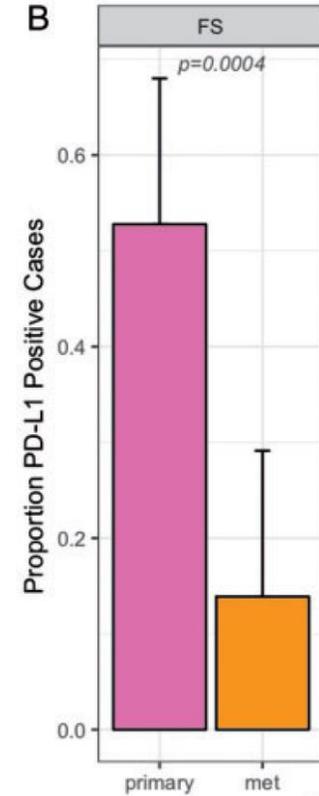
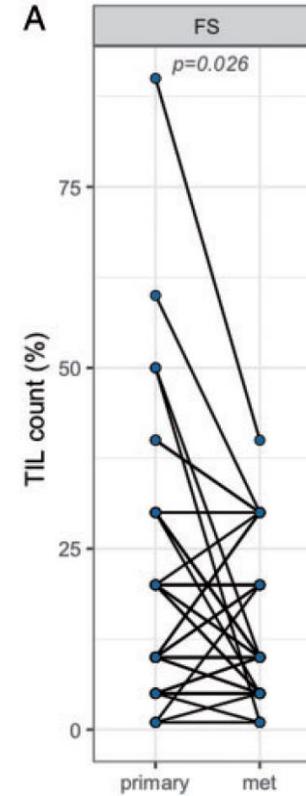
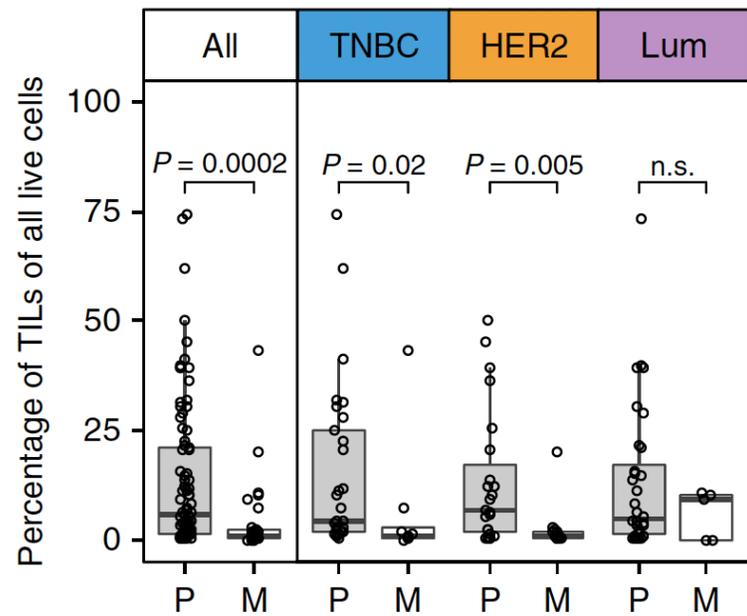
- Mutational load
- Neoantigen load
- Antigen presentation
- Immunosuppressive environment
- ...



¹Salgado R, et al. Ann Oncol 2015
Luen S, et al. Breast 2016
Solinas C, et al. Breast 2017
Hamy AS, et al. Clin Cancer Res 2019

TIL levels in primary vs metastatic breast cancer

Levels of TILs (and PD-L1) are *lower* in *metastatic* lesions compared to the *primary* tumor
 → *immune escape*



TIL and PD-L1 protein expression in paired primary and metastatic cancers assessed on full sections (FSs)

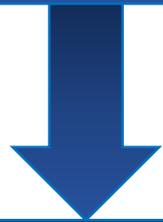
(A) TIL count (%)

(B) PD-L1 positivity rates, defined as $\geq 1\%$ of stromal or tumor cells showing IHC staining

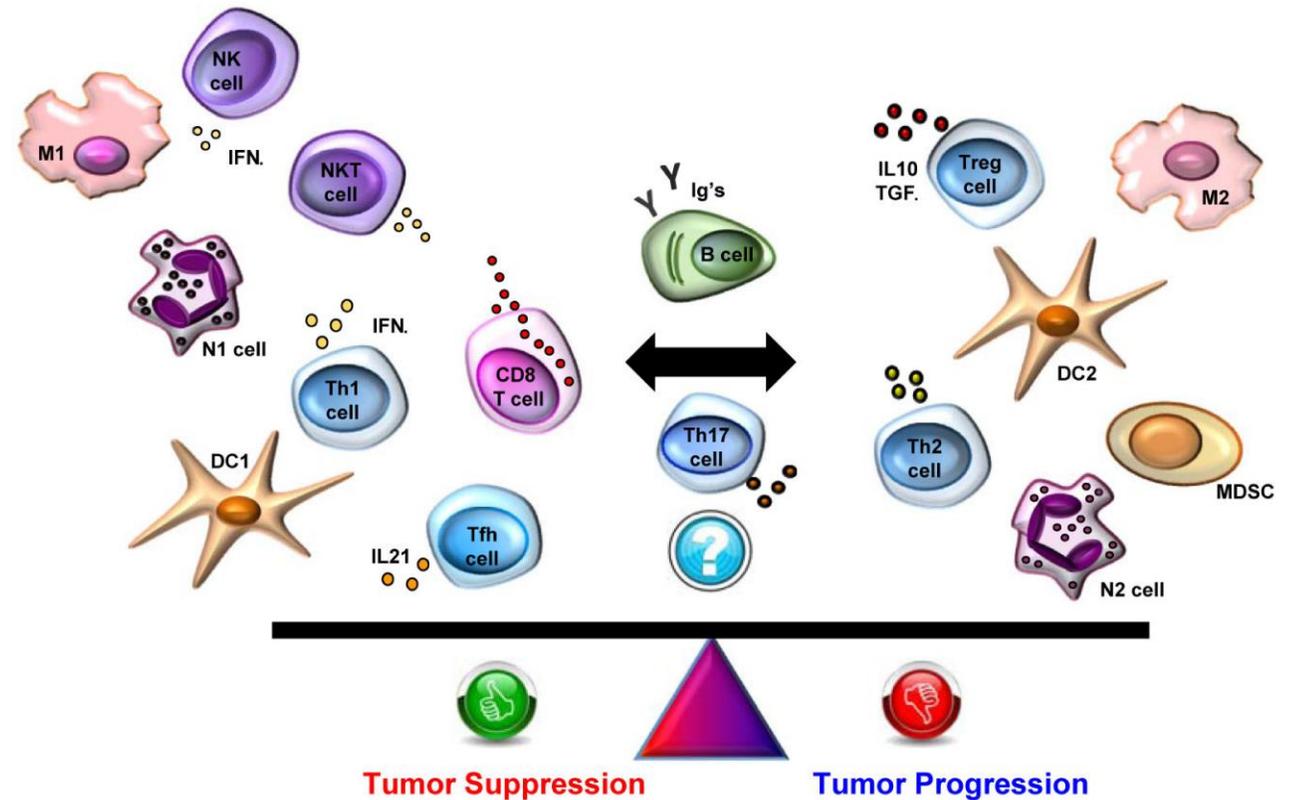
Characterization of *TIL subpopulations*

The cellular constituents of the host immune response to tumors can:

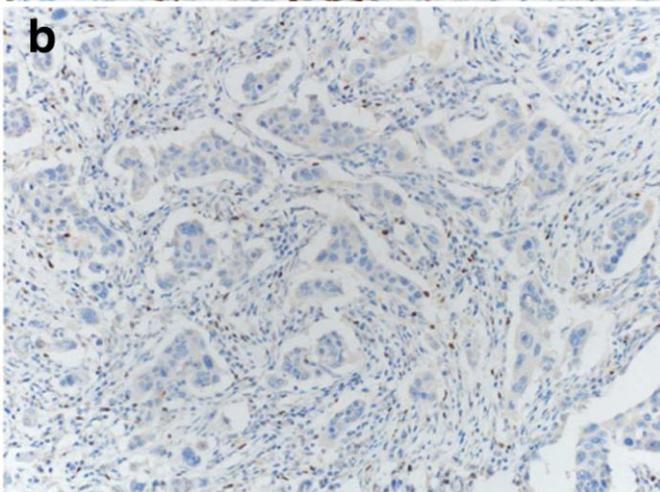
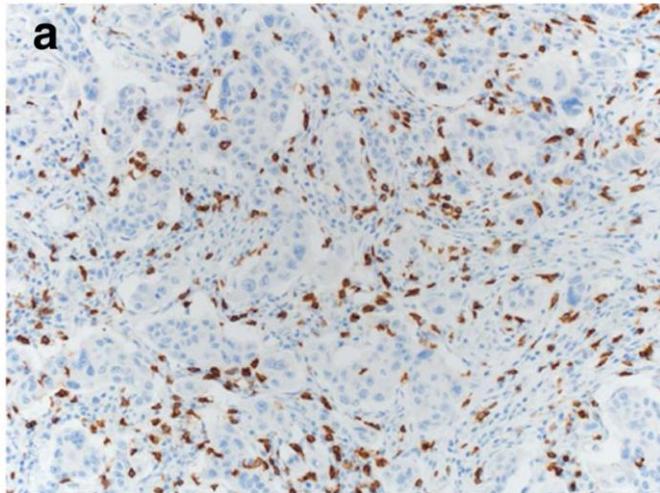
- *control* tumor growth
- contribute to an immunosuppressive environment that *promotes* tumor progression



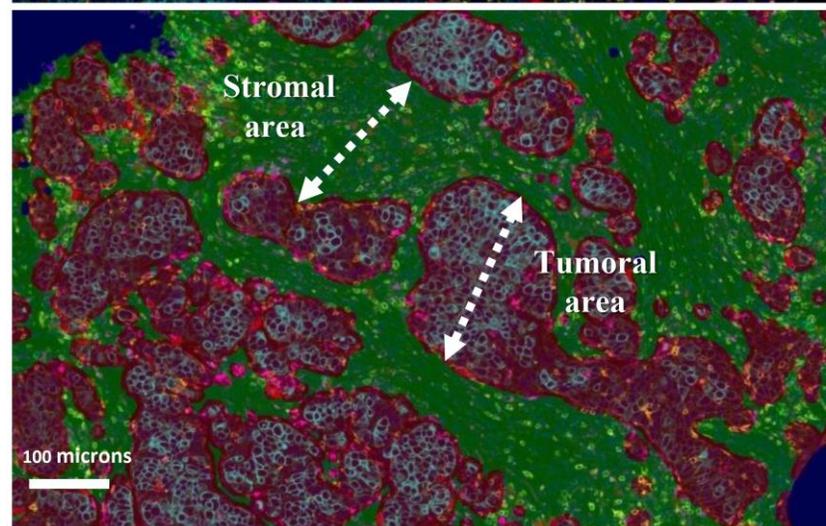
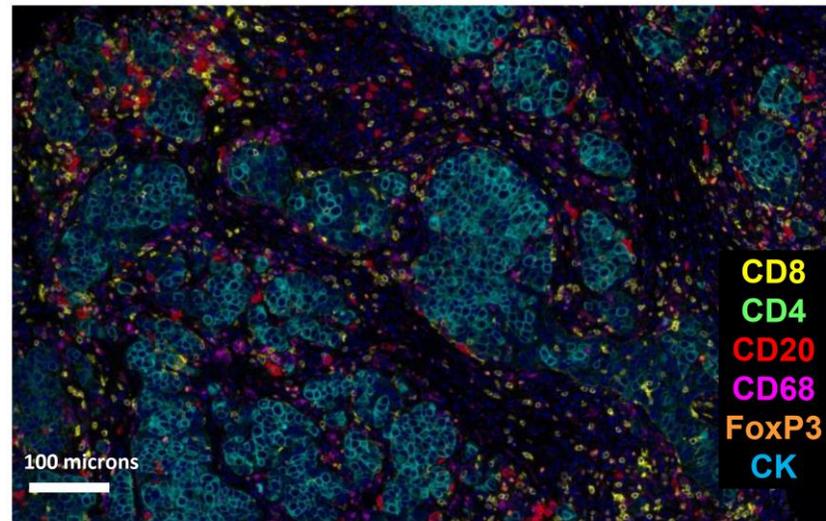
TIL levels alone may not be enough when searching for robust biomarkers



Characterization of *TIL subpopulations* – *Methods*



Tumor CD8+ (a) and FOXP3+ (b) expression as assessed with immunohistochemistry



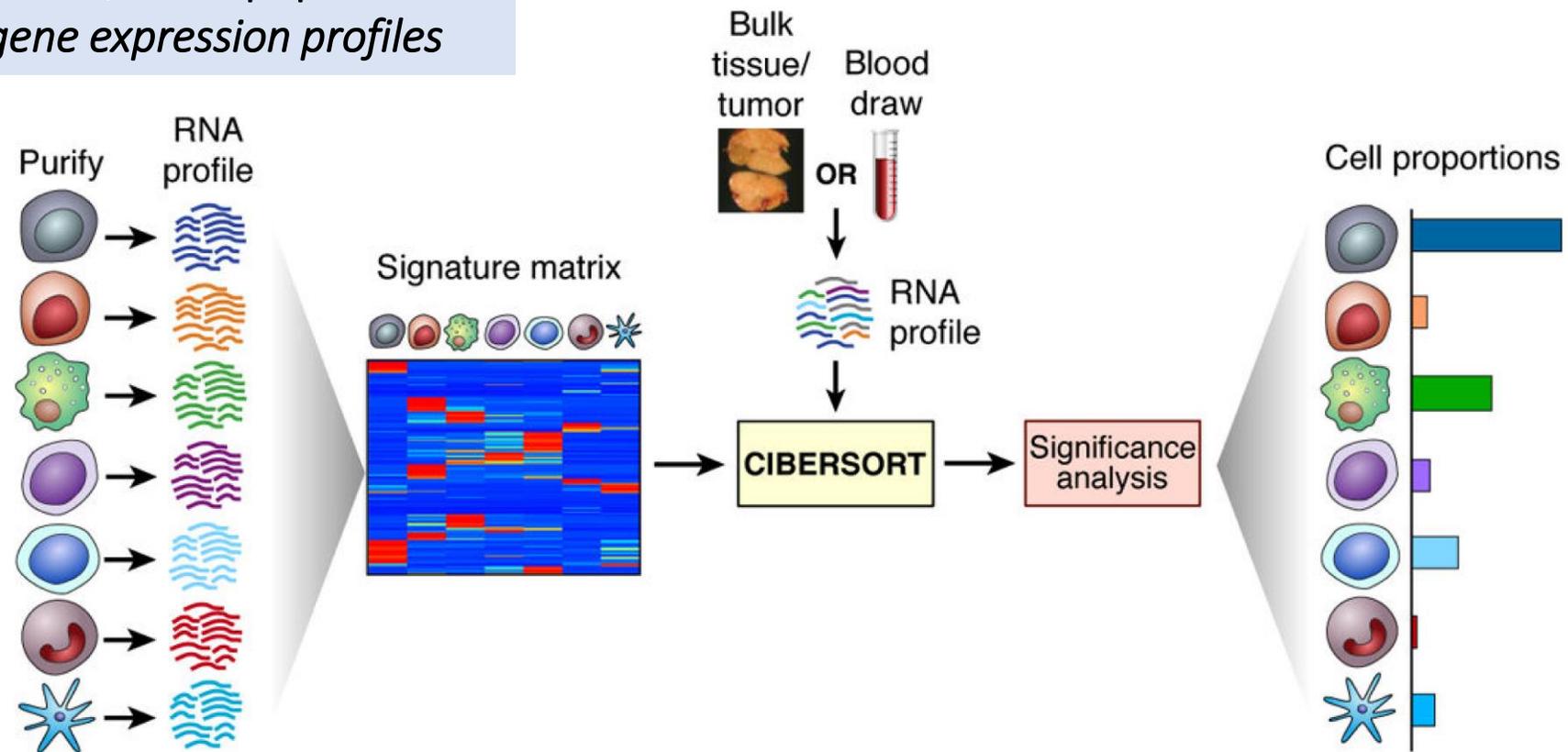
Multiplexed immunofluorescence of tumor infiltrates

- CD8+ → cytotoxic T cells
- CD4+ → helper T cells
- CD20+ → B cells
- CD68+ → Macrophages
- FoxP3+ → regulatory T cells
- CK → Cytokeratin (epithelial cells)

Cluster characterized by high CD4, CD8, CD20 stromal-TILs and CD20 intratumoral-TILs associated with higher pCR rates after lapatinib + trastuzumab

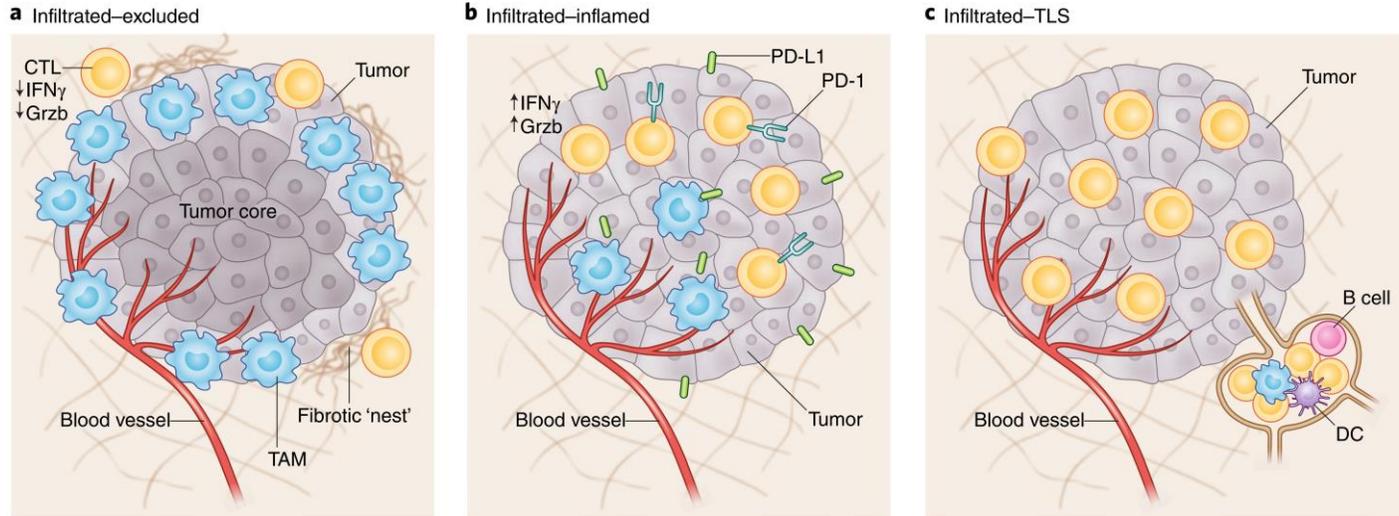
Characterization of *TIL subpopulations* – *Bioinformatics tools*

Computational methods can be used to estimate TIL levels and/or subpopulations from *bulk tissue gene expression profiles*



Overview of CIBERSORT

Immune phenotyping – *geographic distribution*



TLS = tertiary lymphoid structures
 CTL = cytotoxic lymphocyte
 TAM = tumor-associated macrophages
 DC = dendritic cell

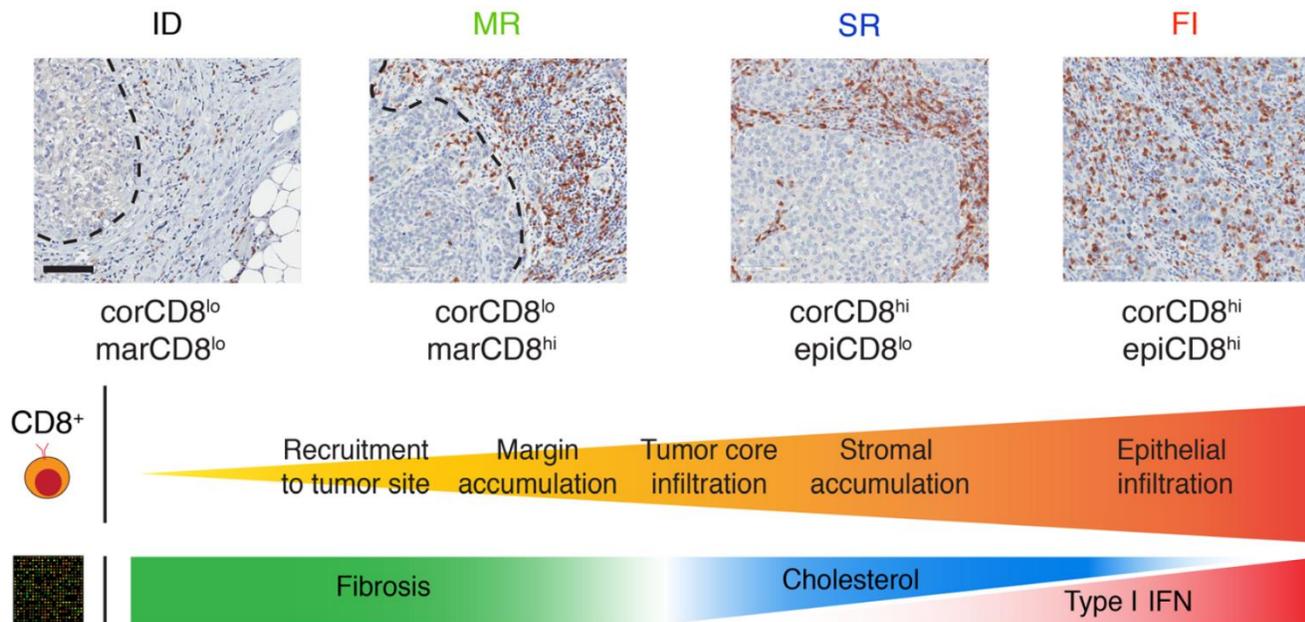
Tumor Immune-Microenvironment (*TIME*) classification in TNBC

ID: Immune Desert

MR: Margin-Restricted

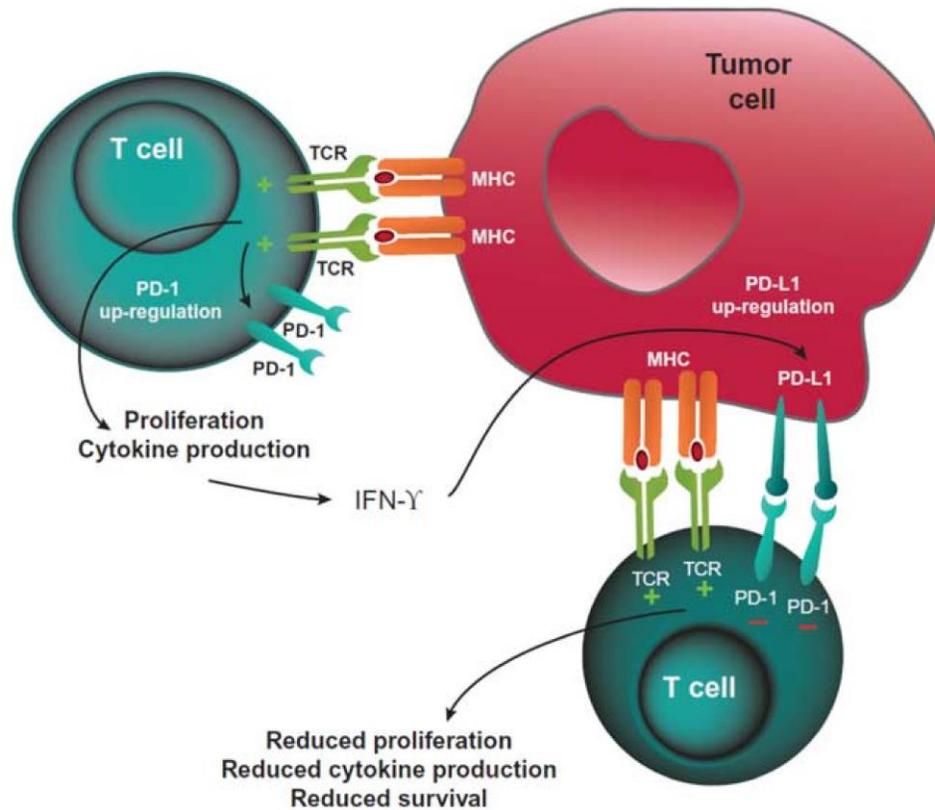
SR: Stroma-Restricted

FI: Fully Inflamed

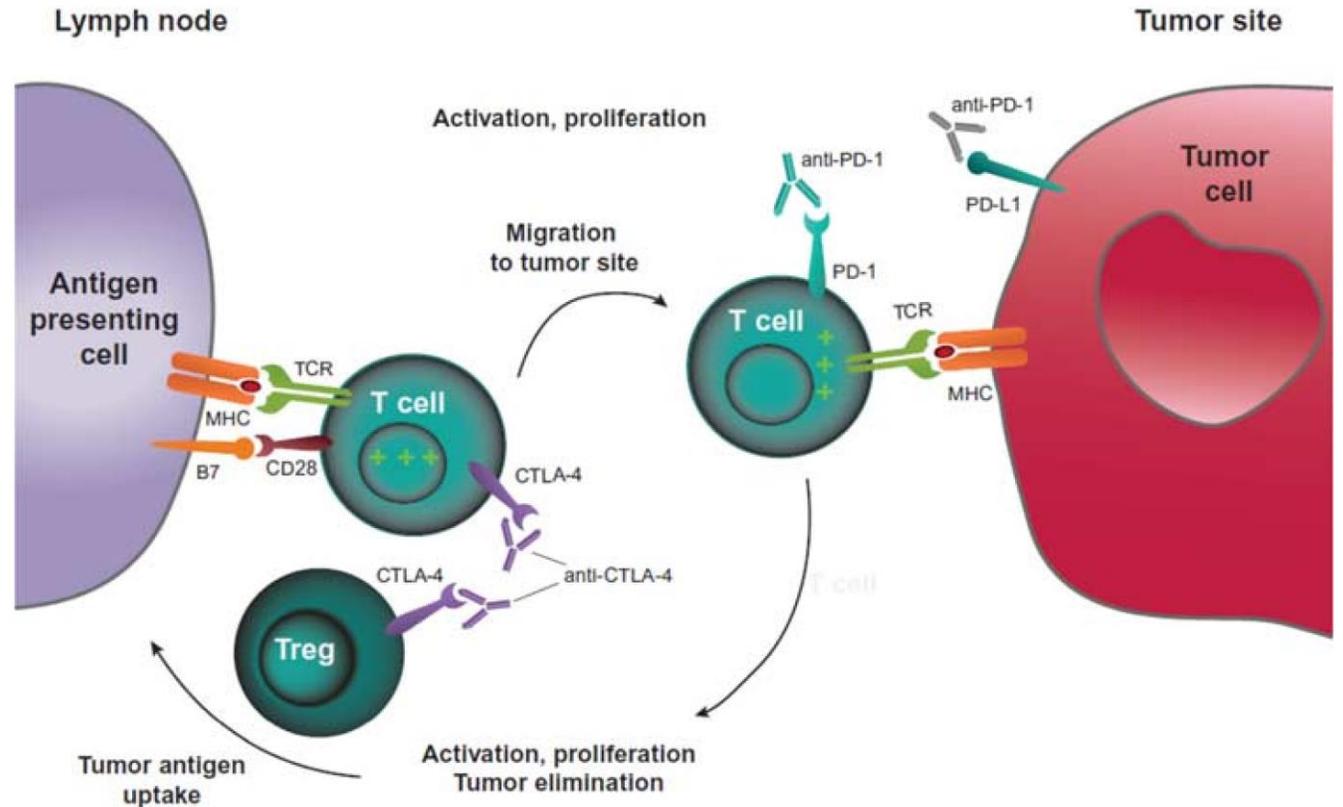


TILs and *PD1/PD-L1* axis

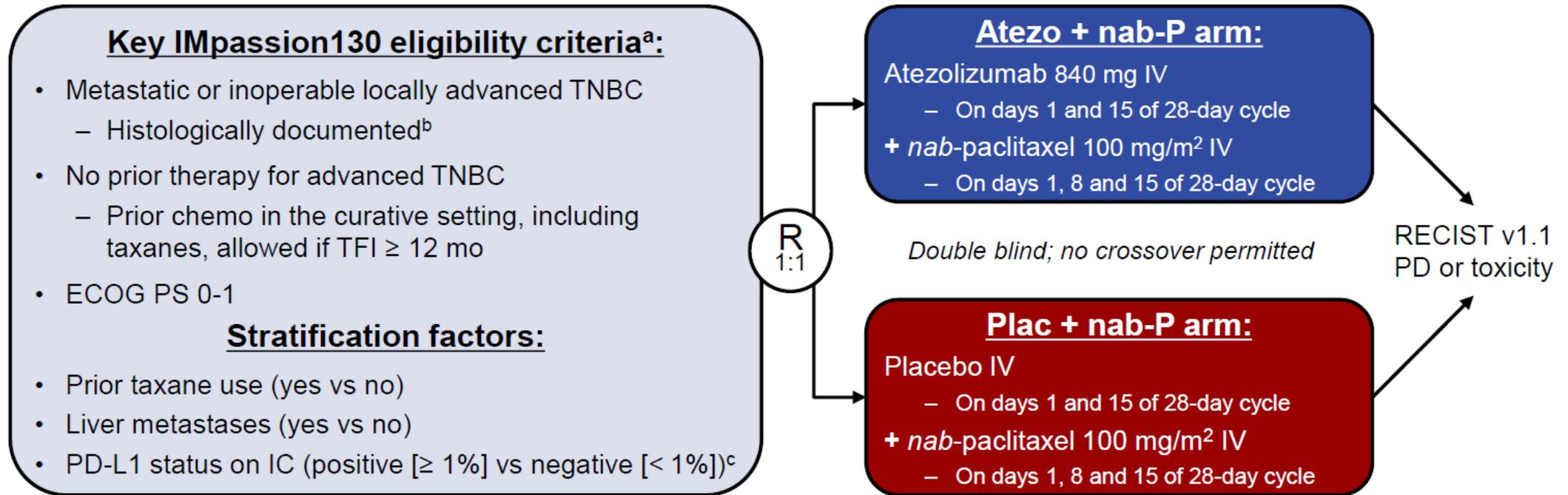
PD-1-mediated inhibition of T cells



CTLA-4 and PD-1 pathway blockade



IMpassion130 – study design

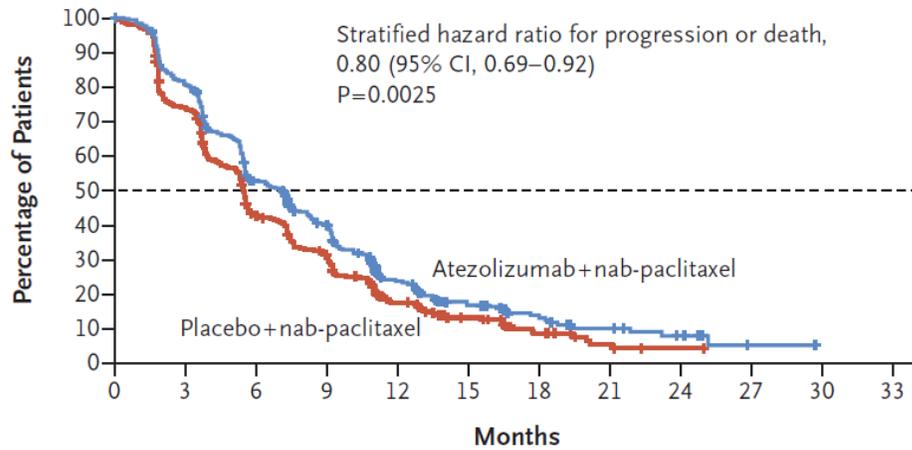


- Co-primary endpoints were PFS and OS in the ITT and PD-L1+ populations^d
 - Key secondary efficacy endpoints (ORR and DOR) and safety were also evaluated

IMpassion130 – results – PFS

A Progression-free Survival in the Intention-to-Treat Population

	No. of Events/ No. of Patients	Median Progression-free Survival (95% CI) <i>mo</i>	1-Yr Rate of Progression-free Survival (95% CI) %
Atezolizumab+Nab-Paclitaxel	358/451	7.2 (5.6–7.5)	23.7 (19.6–27.9)
Placebo+Nab-Paclitaxel	378/451	5.5 (5.3–5.6)	17.7 (14.0–21.4)

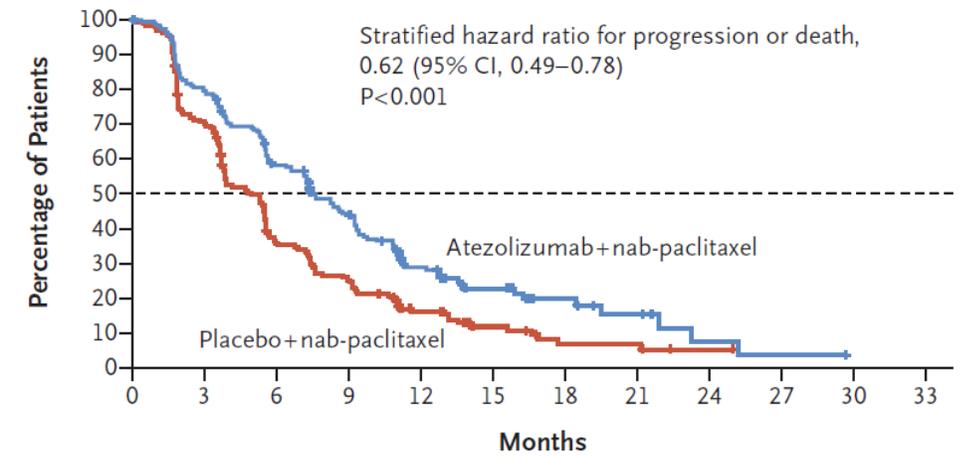


No. at Risk

Atezolizumab+ nab-paclitaxel	451	360	226	164	77	34	20	11	6	1	NE	NE
Placebo+ nab-paclitaxel	451	327	183	130	57	29	13	5	1	NE	NE	NE

B Progression-free Survival in the PD-L1-Positive Subgroup

	No. of Events/ No. of Patients	Median Progression-free Survival (95% CI) <i>mo</i>	1-Yr Rate of Progression-free Survival (95% CI) %
Atezolizumab+Nab-Paclitaxel	138/185	7.5 (6.7–9.2)	29.1 (22.2–36.1)
Placebo+Nab-Paclitaxel	157/184	5.0 (3.8–5.6)	16.4 (10.8–22.0)



No. at Risk

Atezolizumab+ nab-paclitaxel	185	146	104	75	38	19	10	6	2	1	NE	NE
Placebo+ nab-paclitaxel	184	127	62	44	22	11	5	5	1	NE	NE	NE

Table 1 | **Factors that predict response to immune checkpoint inhibitor therapy**

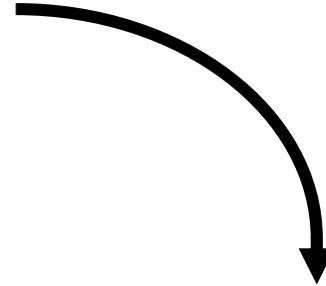
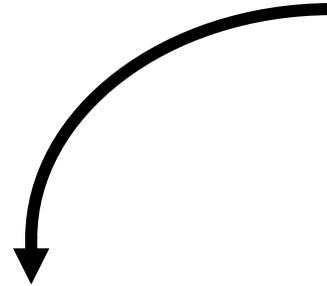
Factor	Association with favourable clinical outcome	Validated in phase III clinical trial?	Predictive versus prognostic ^a	Cancer type	Tissue type for biomarker assessment ^b	Possible assay type for biomarker assessment
Tumour mutation burden	Positive	Yes	Predictive	Multiple cancer types	Blood or tumour tissue	NGS WES or targeted gene panel sequencing
PDL1 expression	Positive	Yes	Predictive	Multiple cancer types	Tumour tissue	Immunohistochemistry
Copy number variation	Negative	TBD	Prognostic, predictive or both	Multiple cancer types	Tumour tissue	NGS WES or targeted gene panel sequencing
HLA class I diversity	Positive	TBD	Predictive	Melanoma and NSCLC	Blood	NGS WES or PCR-based typing
LOH at HLA class I alleles	Negative	TBD	Predictive	Melanoma	Tumour tissue	TBD
T cell repertoire clonality change	Positive	TBD	Predictive	Melanoma	Tumour tissue or blood	TBD
T cell-inflamed microenvironment	Positive	TBD	Prognostic, predictive or both	Multiple cancer types	Tumour tissue	NGS RNA-seq or immunostaining
<i>SERPINB3</i> or <i>SERPINB4</i> mutations	Positive	TBD	Predictive	Melanoma	Tumour tissue	NGS WES
Gut microbial diversity	Positive	TBD	Predictive	Melanoma	Oral or gut	PCR or NGS
Specific gut microbial species	Positive or negative	TBD	Predictive	Melanoma	Oral or gut	PCR or NGS
TGFβ expression	Negative	TBD	Predictive	Colon cancer and urothelial cancer	Tumour tissue	NGS RNA-seq or expression panel
Mutations in the β-catenin pathway	Negative	TBD	Predictive	Melanoma	Tumour tissue or blood	NGS WES, targeted gene panel sequencing or RNA-seq
<i>JAK2</i> mutations (rare) ^c	Negative	TBD	Predictive	Melanoma	Tumour tissue or blood	NGS WES or targeted gene panel sequencing
<i>B2M</i> mutations (rare) ^c	Negative	TBD	Predictive	Melanoma	Tumour tissue or blood	NGS WES or targeted gene panel sequencing
<i>STK11</i> mutations (common)	Negative	TBD	Predictive	NSCLC	Tumour tissue or blood	NGS WES or targeted gene panel sequencing

HLA, human leukocyte antigen; LOH, loss of heterozygosity; NSCLC, non-small-cell lung cancer; NGS, next-generation sequencing; PDL1, programmed cell death 1 ligand 1; RNA-seq, RNA sequencing; TBD, to be determined; TGFβ, transforming growth factor-β; WES, whole-exome sequencing. ^aPredictive refers to a given biomarker that has an effect dependent on the immune checkpoint inhibitor therapy, and prognostic refers to a biomarker that has a specific effect independent of the therapy. ^bBlood detection of mutations refers to cell-free DNA analysis. ^c*JAK2* and *B2M* mutations are controversial. Responses have been seen in patients with these mutations. Intratumoural heterogeneity likely needs to be assessed along with these mutations.

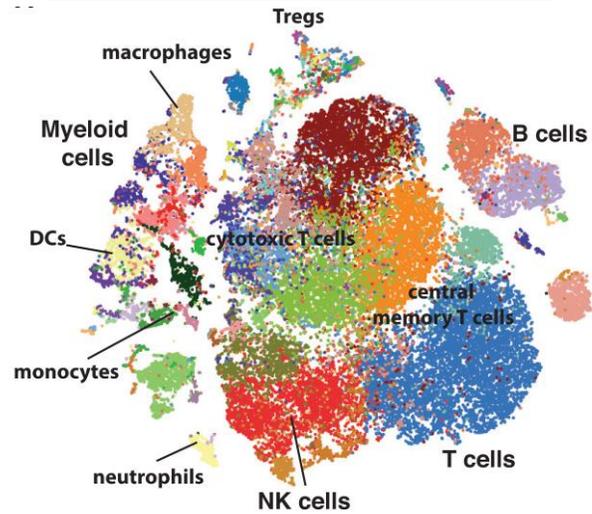
Outline

- Introduction to the clinical classification
- Molecular characterization and intertumor heterogeneity
 - Examples of clinical applications
- The challenge of intratumor heterogeneity
- The role of the tumor immune microenvironment
- ***Novel translational research tools and future perspectives***

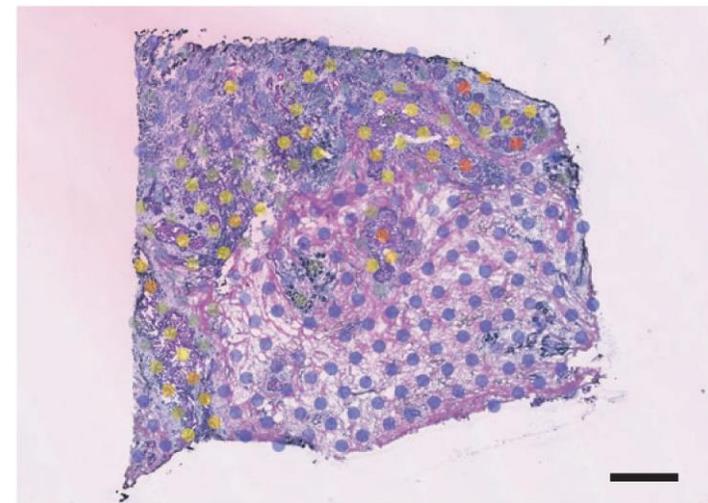
Technological advances



Single-cell analysis



Spatial transcriptomics

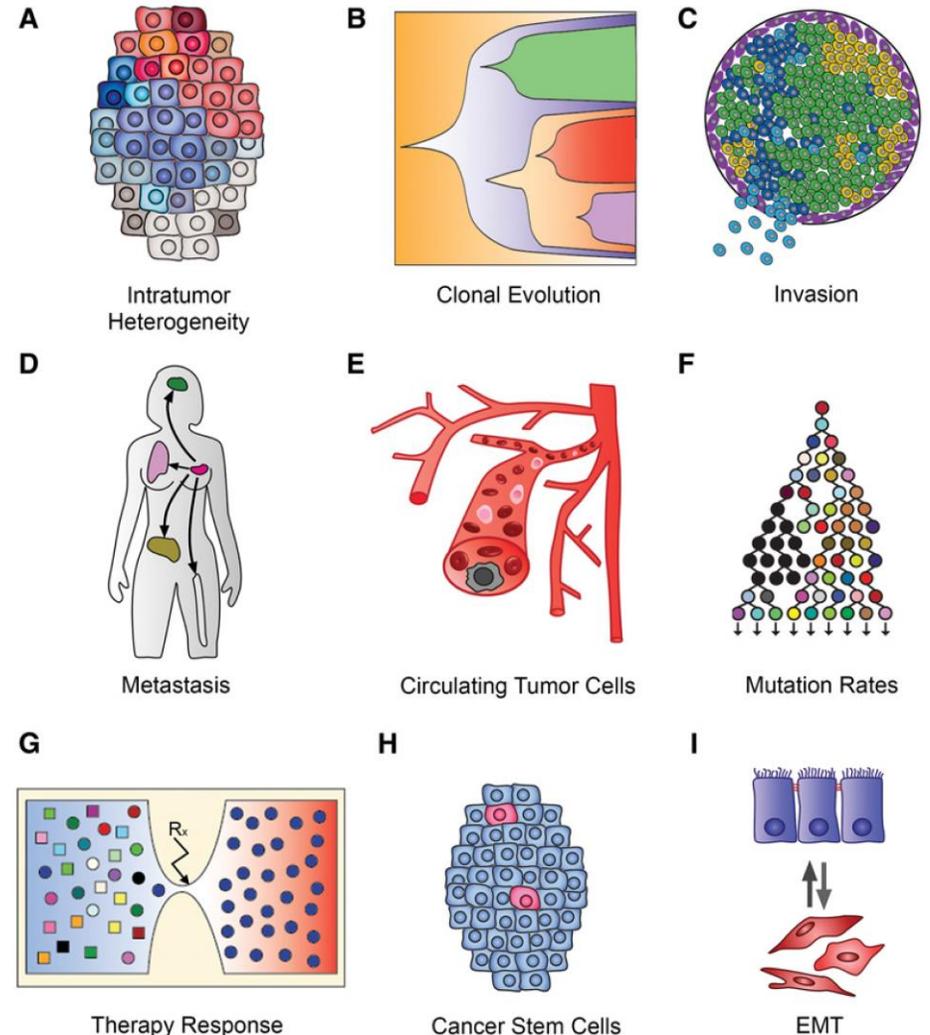


Single-cell analysis

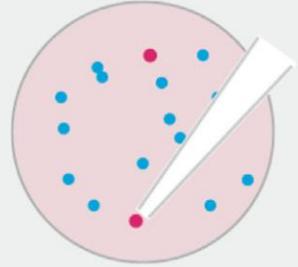
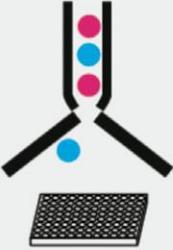
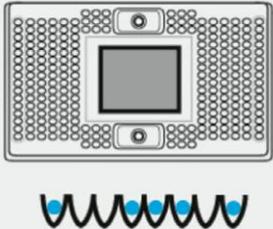
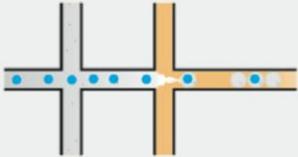
Bulk tissue → mixture of **different types of cells** (tumor cell subpopulations, immune cells, stroma, ...)
→ **Transcriptomics/genomics studies** use RNA/DNA sequencing of **homogenized samples**
→ Averaged transcriptome and mixture of mutational/CNAs data from **different cell types**

- Examples of single-cell “omics” techniques:
 - **RNA-sequencing**
 - **DNA-sequencing** (e.g. for CNV)
 - **ATAC-sequencing** (for single-cell **epigenomics**)
 - **Immune profiling** (e.g. cell surface proteins, antigen specificity)

Applications of single-cell sequencing in cancer research



Single cell *isolation methods* for RNA-seq

	Micro-manipulation / Automated Pipetting	FACS	Microwell encapsulation	Droplet encapsulation
				
Cell Stress	Low	Moderate	Moderate	Moderate
Selection	Yes	Yes	No* / Yes ⁺⁺	No*
Doublet	Low	Low	Low-High	Moderate
Throughput	Low	Moderate	Moderate	High
Capture efficiency	Low	Moderate	Moderate	Low-Moderate
Academic / Commerical scRNA workflow	- CellenONE (Cellenion) [‡] - Smart-Seq2 (42)	- MARS-Seq (39) - Smart-Seq2 (42)	- C1 (Fluidigm) - ddSeq (Biorad / Illumina) - iCell8 (Clontech) ⁺⁺ - Rhapsody (BD)	- InDrop (1CellBio) - DropSeq (Dolomite-bio) - 10X (Chromium)
Example of use	Fragile rare cells	Rare cells based on phenotype or marking	Large cell numbers	Large cell numbers

[‡]Automated pipetting system

*Preselection or enrichment can be performed prior

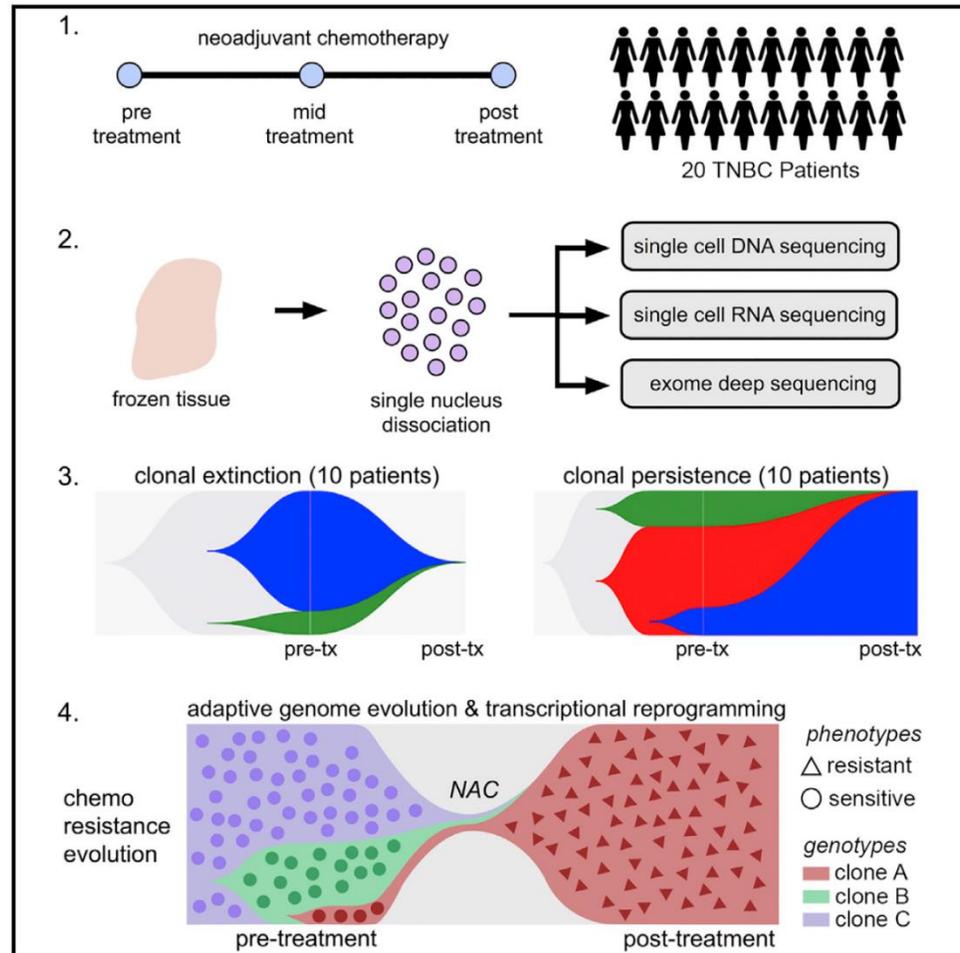
⁺⁺Only reagents added to wells containing singlets, determined by system

FWP: Fluidigm white paper

PB: Product brochure / manual

Chemoresistance evolution in TNBC delineated by single-cell sequencing

Graphical Abstract



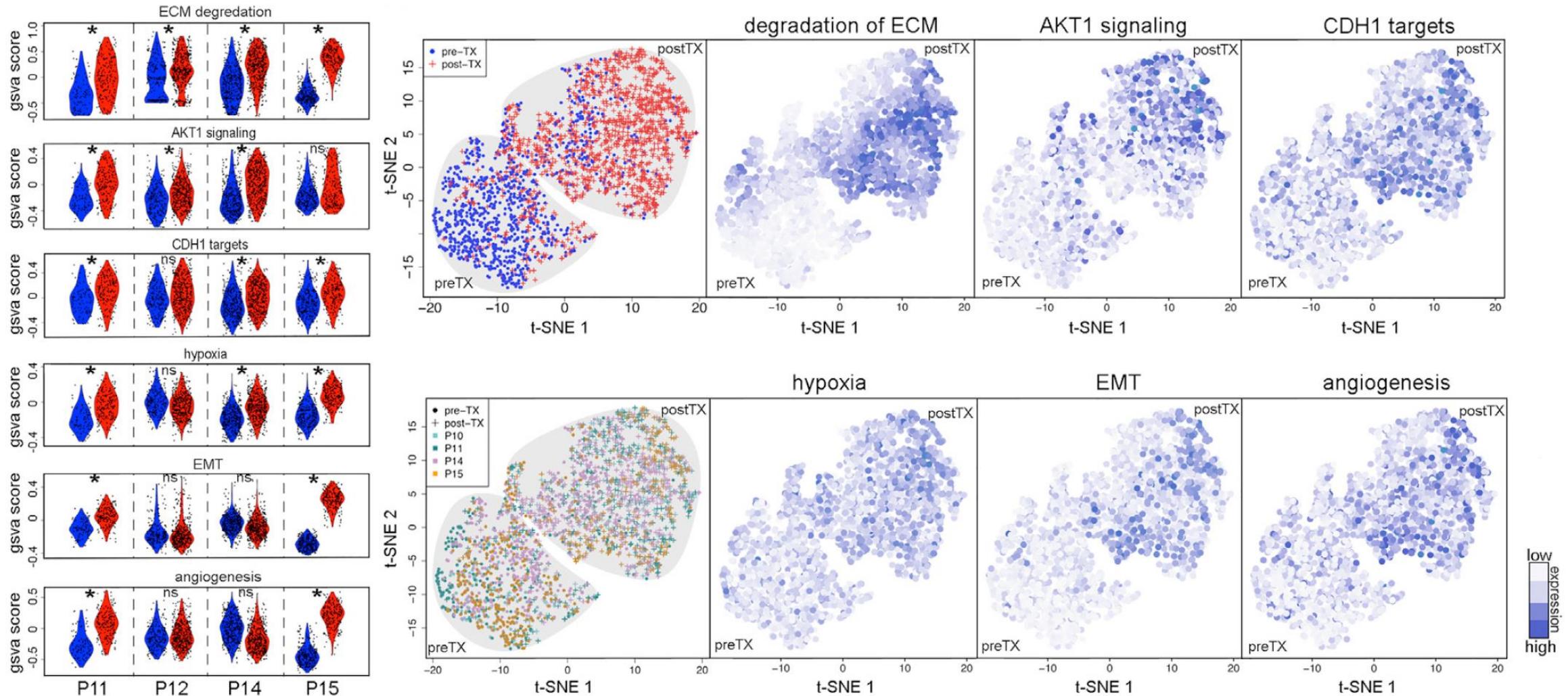
30-50% of TNBCs are **resistant** to NeoADJ chemotherapy
→ lack of genomic biomarkers

Highlights

- Single-cell sequencing of breast cancer patients treated with chemotherapy
- Patients showed clonal persistence or extinction in response to therapy
- Resistance occurred through adaptive selection of pre-existing genomic aberrations
- Chemotherapy induced transcriptional reprogramming of resistant signatures

Gene signatures associated with *chemoresistance* in TNBC

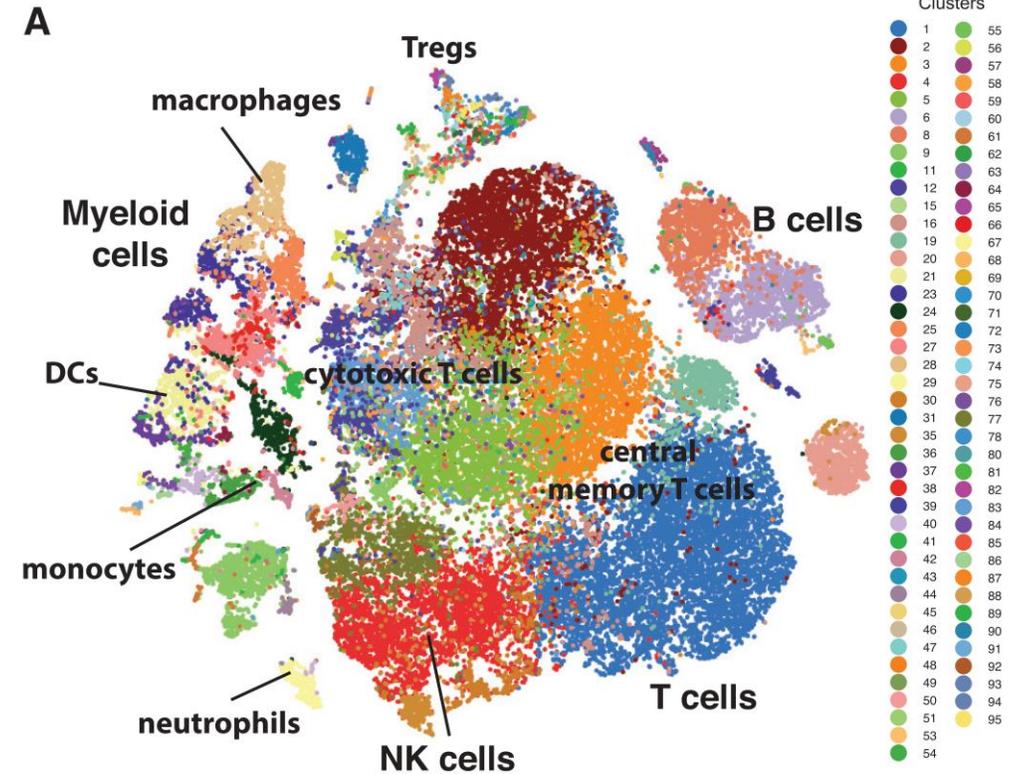
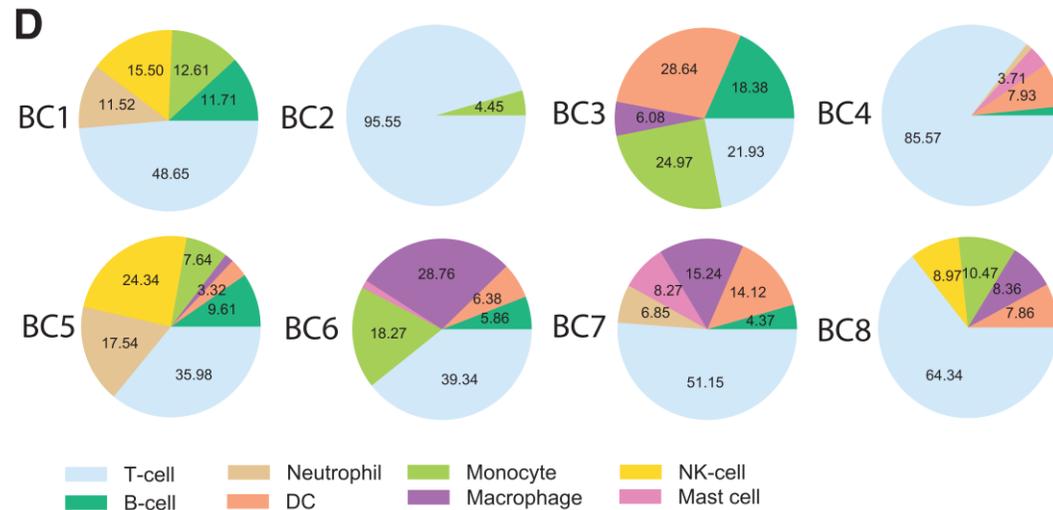
→ transcriptional reprogramming and therapeutic opportunities to overcome resistance



Combined single-cell data from four clonal persistence patients

Characterization of the tumor *immune microenvironment*

- Single-cell RNA-sequencing of breast tumor immune microenvironment to build *immune atlas* in breast carcinoma
- This atlas revealed **vast diversity** in immune cells of both the adaptive and innate immune systems

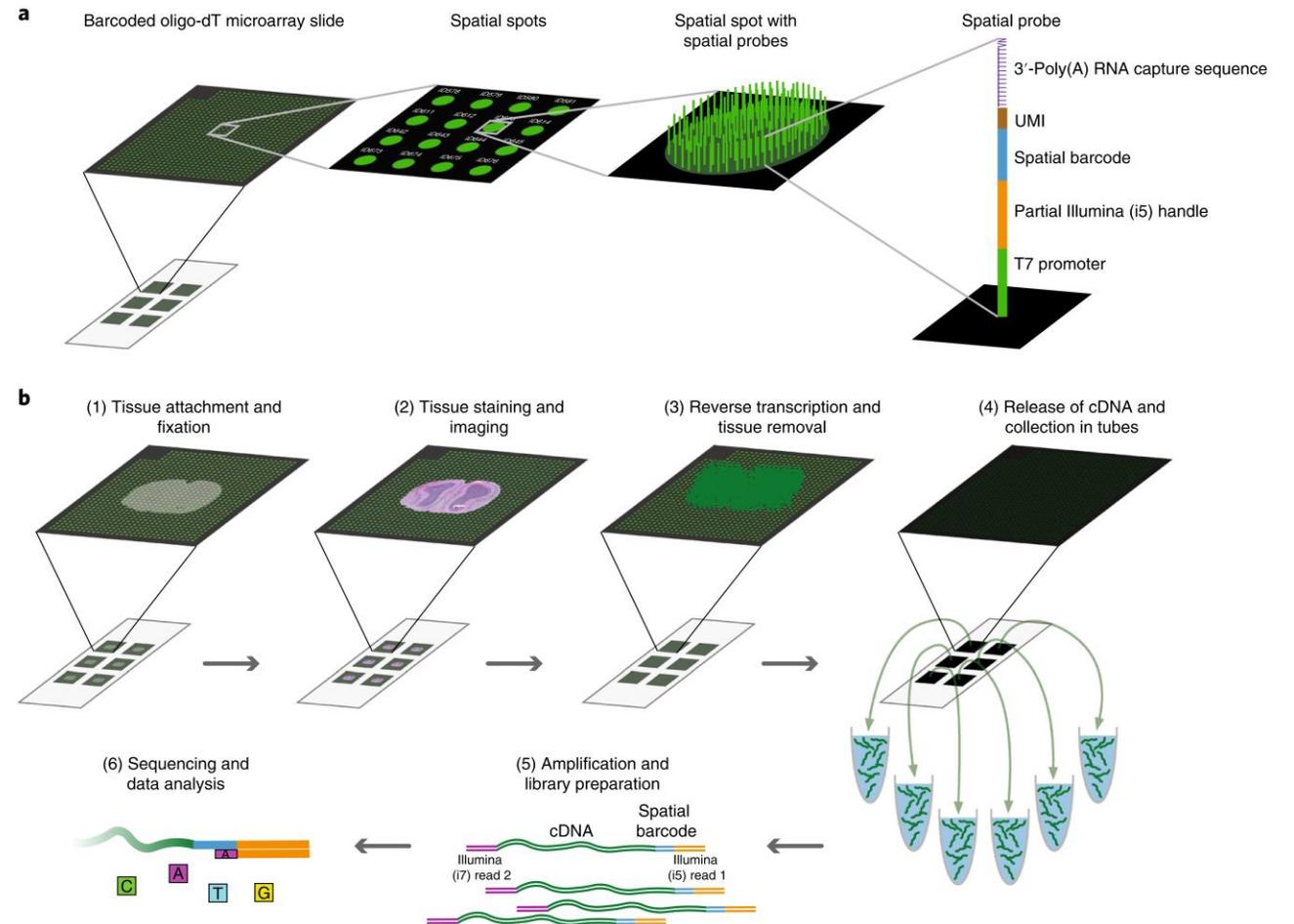
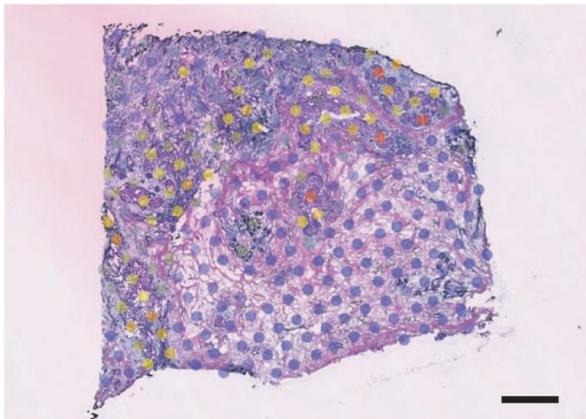


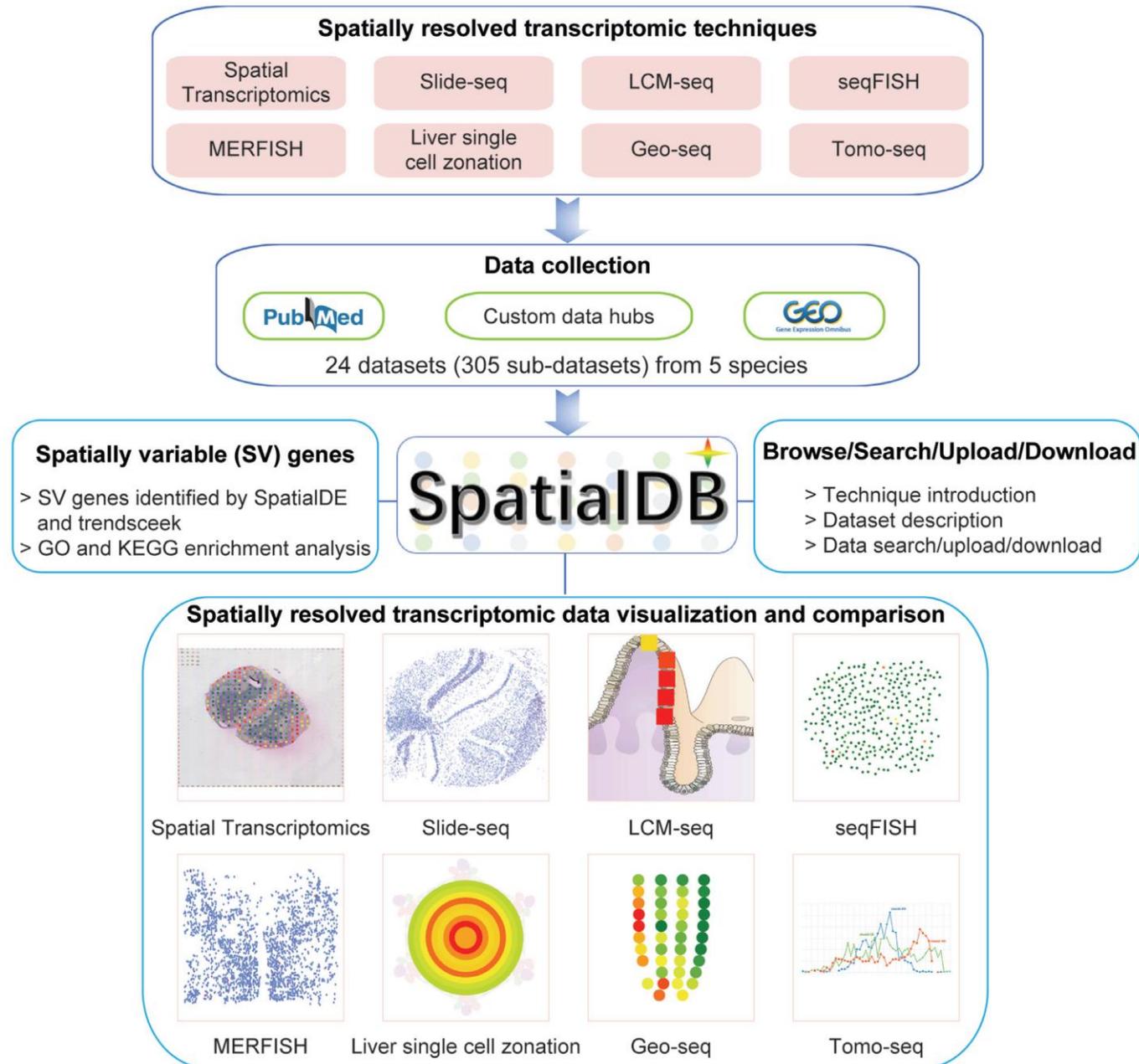
Breast immune cell atlas constructed from combining all patient samples (BC1-8) and tissues projected with t-SNE. Each dot represents a cell, colored by cluster.

t-SNE = t-distributed stochastic neighbor embedding

Spatially resolved transcriptomics

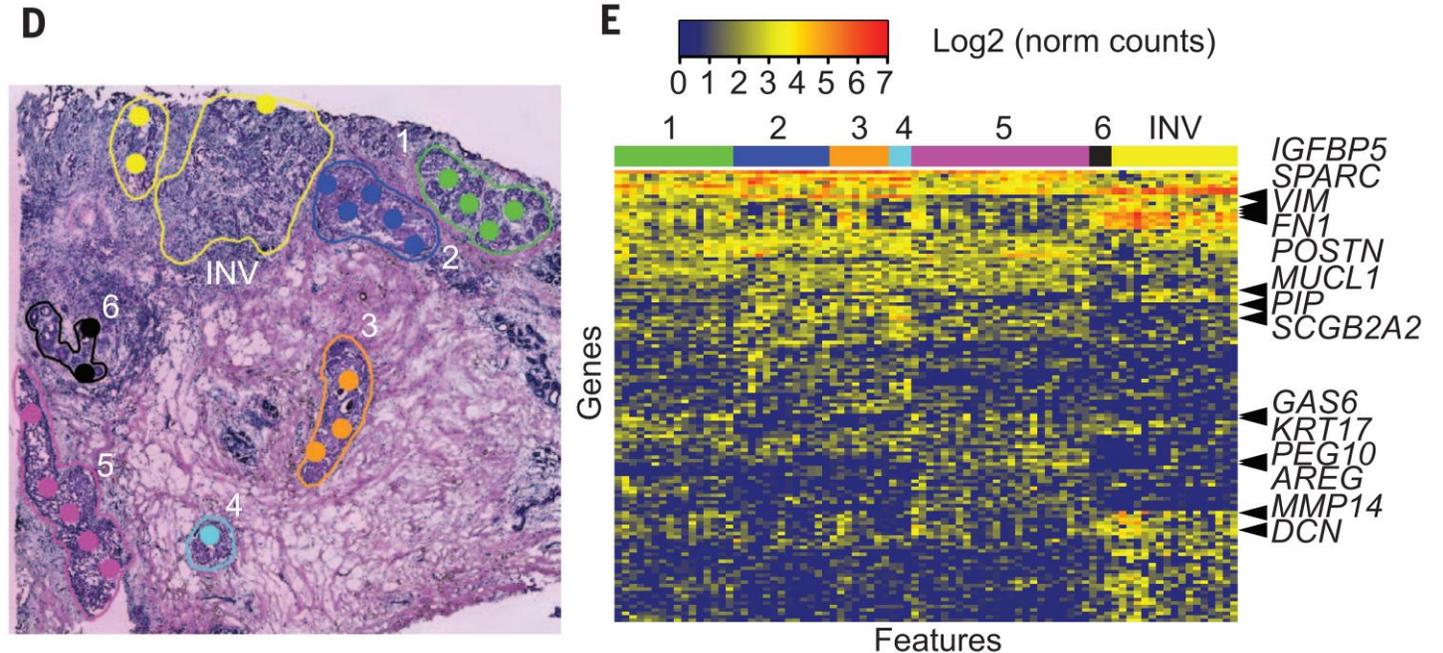
- **Single-cells** are collected from suspensions of dissociated tissue
→ *loss of spatial information*
- Spatial transcriptomics allows to **retain** the **positional context** of *gene expression levels* and to combine those data with *morphological information*
- Resolution down to **1-10 cells**





Spatial transcriptomics and *tumor heterogeneity*

- Different tumor areas (e.g. ductal carcinoma *in situ*) can present high degree of **heterogeneity** in gene expression
- Unexpected level of heterogeneity within a biopsy, which would **not** be possible to detect with regular **bulk** transcriptome analysis

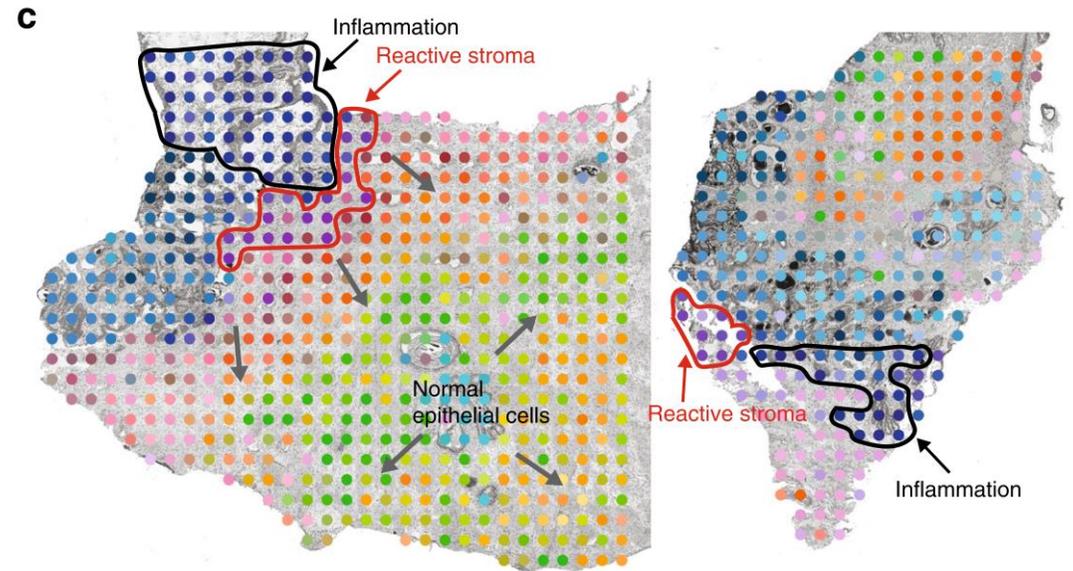


(D) Histological section of a breast cancer biopsy containing invasive ductal cancer (INV) and six separate areas of ductal cancer in situ (1 to 6), with analyzed spatial transcriptomics features.

(E) Gene expression heat map over the different areas in four adjacent sections (D)

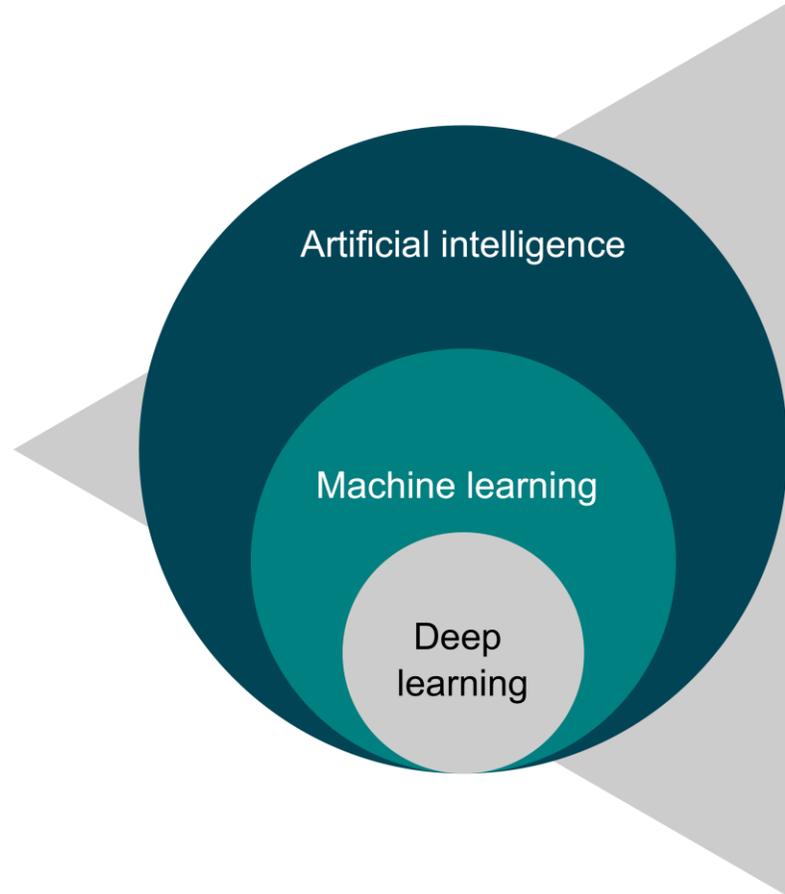
Spatial transcriptomics – *potential applications*

- Measure **gene activity** and **map biological processes** (e.g. EMT, CSC, immune response)
- Correlation between **gene expression** and **morphological intratumor heterogeneity**
- Characterization of heterogeneous tumor cell **subpopulations**
- Characterization of **cell-cell interactions** (e.g. tumor cells and microenvironment, including immune and stroma cells)



Stromal heterogeneity and reactive stroma in the microenvironment of inflammation in prostate cancer

Artificial Intelligence



Artificial intelligence

Artificial Intelligence is the science of making machines do things requiring human intelligence. It is human intelligence in machine format where computer programs develop data-based decisions and perform tasks normally performed by humans.

Artificial intelligence is any computer program that does something smart.

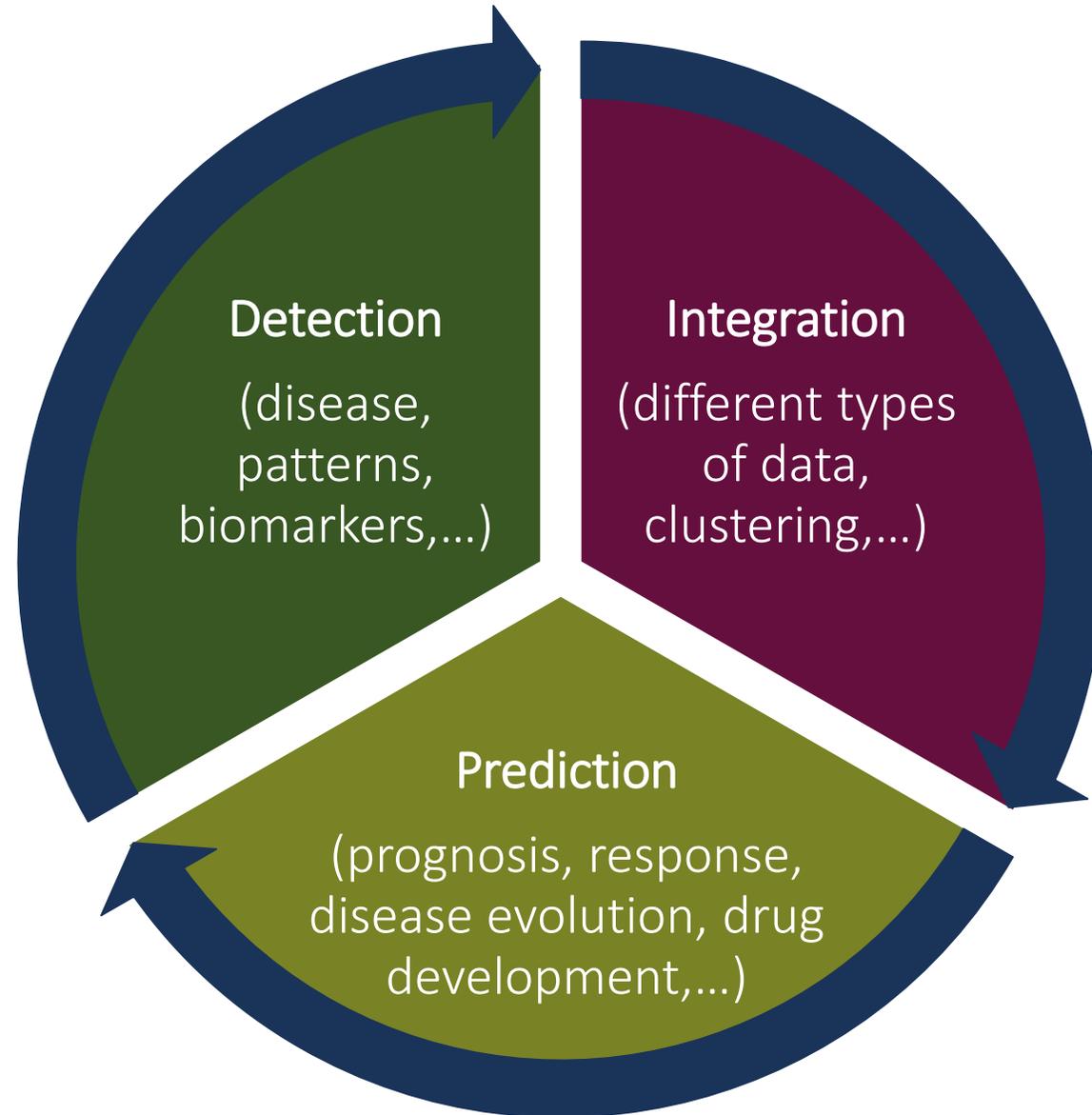
Machine learning

Machine learning takes artificial intelligence a step further in the way that algorithms are programmed to learn and improve without the need for human data input and reprogramming.

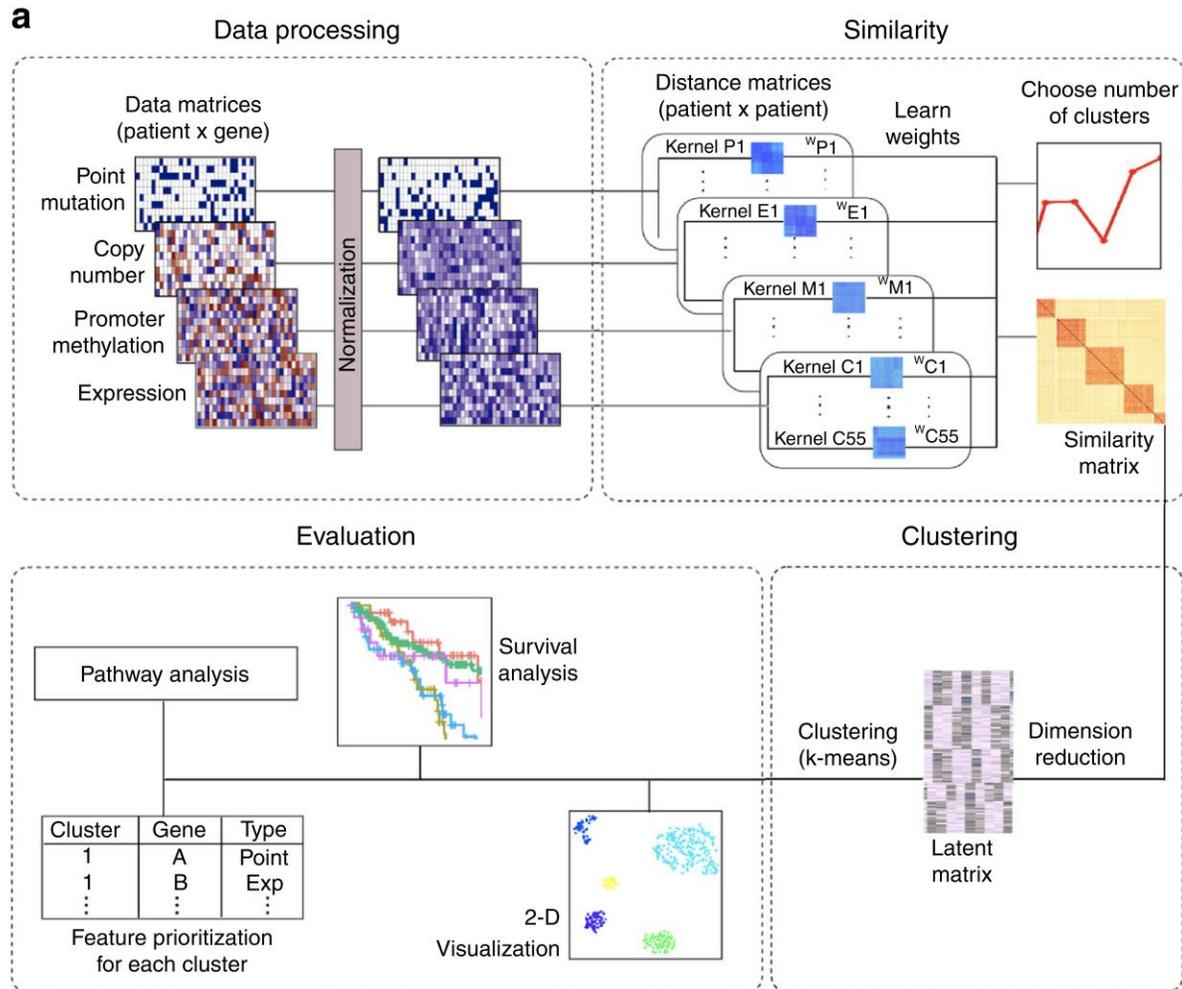
Deep learning

Deep learning is the next generation of machine learning that introduces multiple layers of learning from massive datasets. Deep learning decisions and data classifications are refined at each layer to produce accurate insights.

Artificial intelligence in oncology

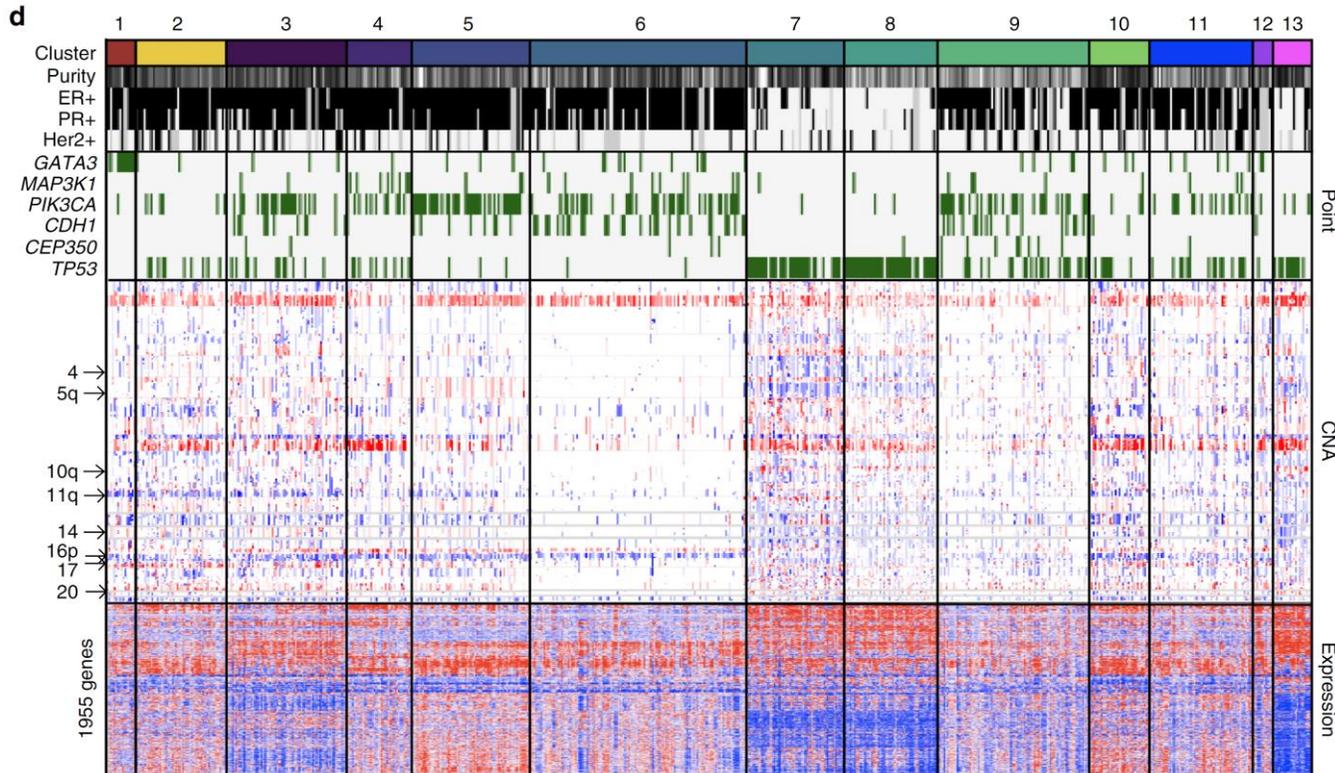


Integration of *multi-omic data* to predict prognosis

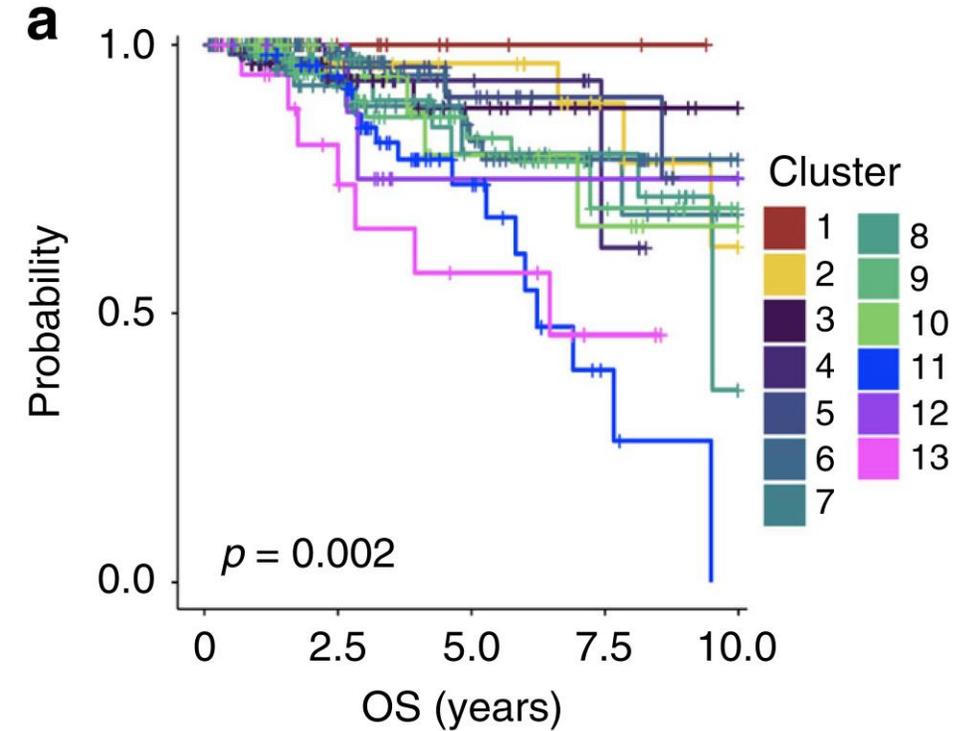


- Cancer Integration via Multikernel Learning (CIMLR) method applied to *multi-omic data* from 36 cancer types from TCGA to reveal molecular subtypes
- Discovered *subtypes* exhibit significant differences in patient **survival** for 27 cancer types
- This method *outperformed* current state-of-the-art tools in **speed, accuracy,** and **prediction** of patient survival

CIMLR in breast cancer



In breast cancer, CIMLR separates 663 tumors into **13 clusters** with different overall and disease-specific survival

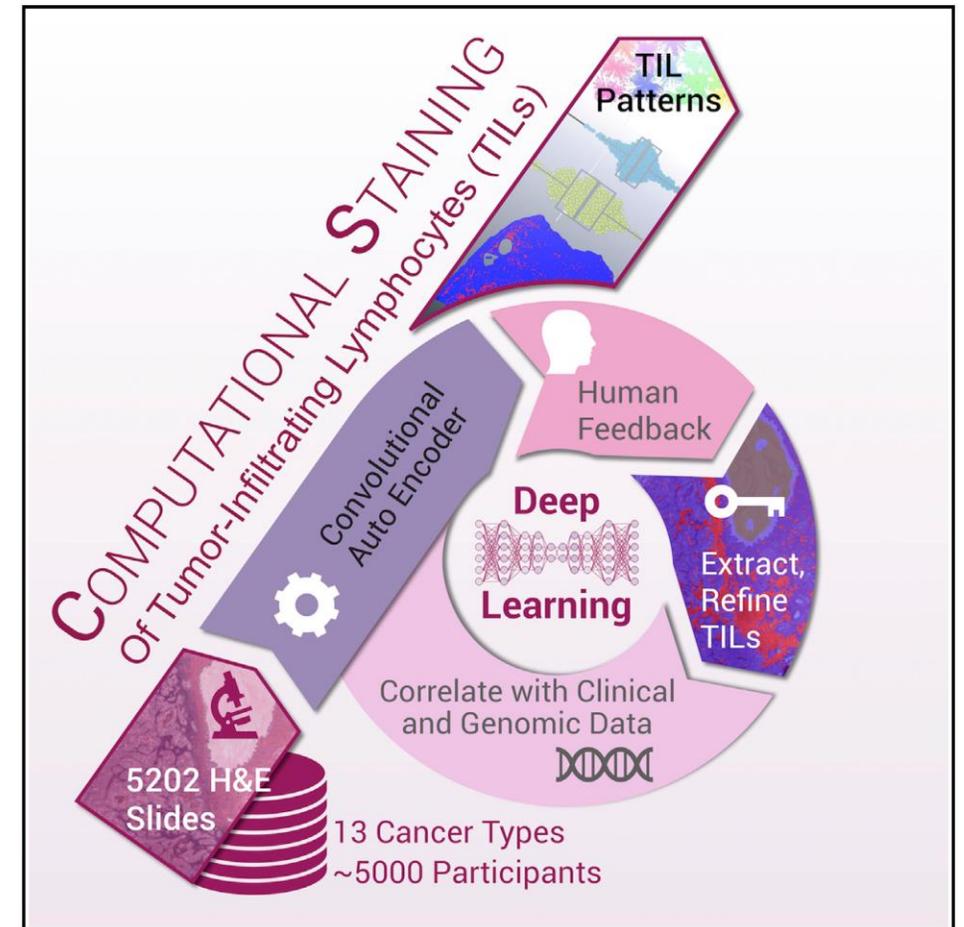


Kaplan–Meier curves showing **overall survival** for the 13 clusters of **breast cancer**

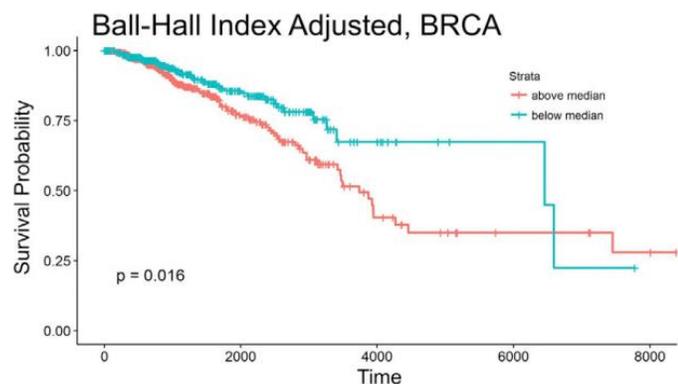
Digital pathology and machine learning to score Tumor Infiltrating Lymphocytes

Highlights

- Deep learning based computational stain for staining tumor-infiltrating lymphocytes (TILs)
- TIL patterns generated from 4,759 TCGA subjects (5,202 H&E slides), 13 cancer types
- Computationally stained TILs correlate with pathologist eye and molecular estimates
- TIL patterns linked to tumor and immune molecular features, cancer type, and outcome



Automated assessment of local structures in the TIL infiltrate and association with molecular and clinical readouts



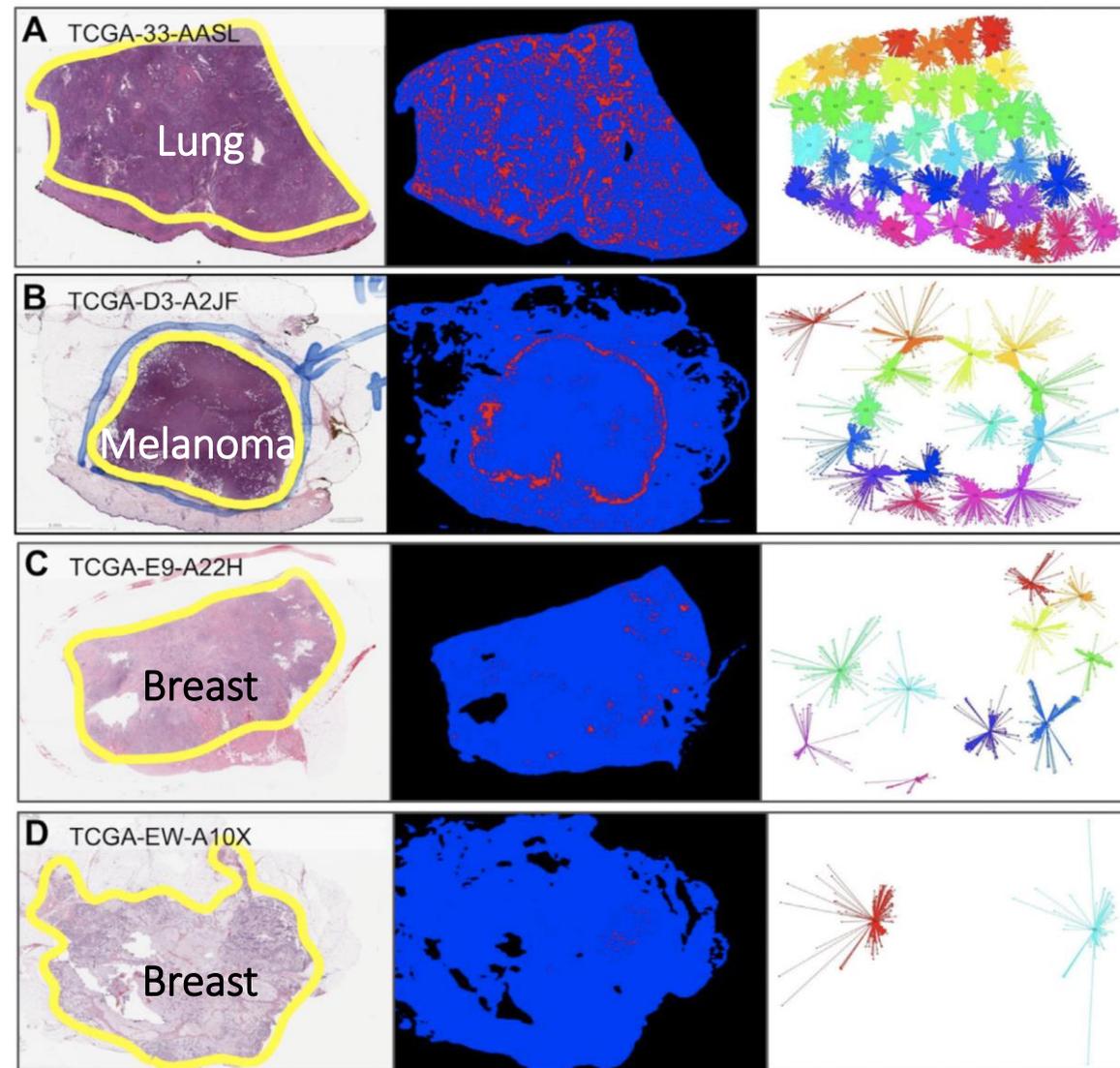
Association of TIL Local Spatial Structure with survival in breast cancer (BRCA)

(A–D) Four cases representing different degrees of lymphocyte infiltration.

Left: H&E diagnostic image at low magnification with tumor regions circled in yellow.

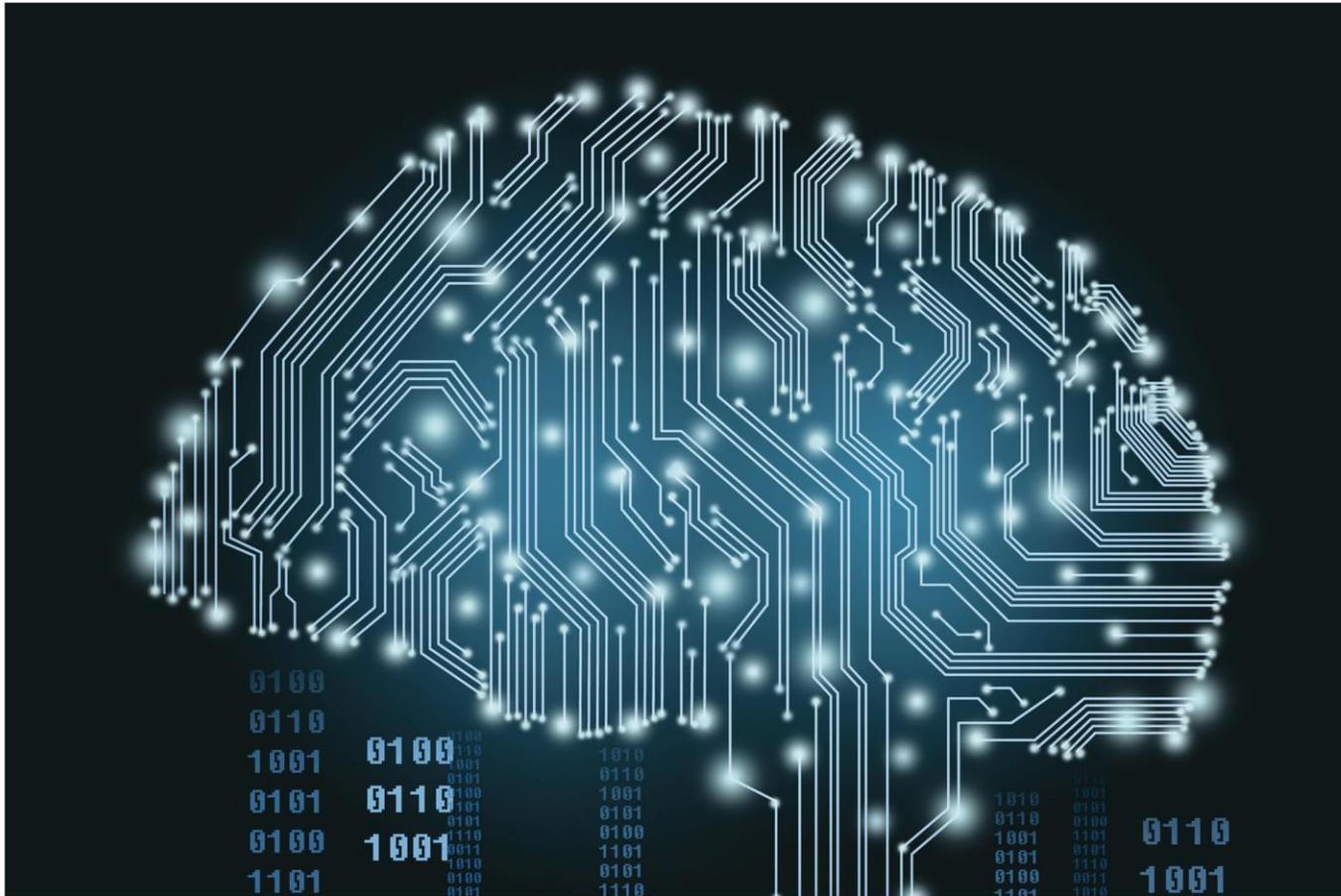
Middle: TIL map; red represents a positive TIL patch, blue represents a tissue region with no TIL patch, while black represents no tissue.

Right: diagrams of clusters of TIL patches derived from the affinity propagation clustering of the TIL patches.



E

Participant Barcode	Study	Number of TIL Patches	TIL fraction	Number of TIL Clusters	Cluster Size Mean	Within-Cluster Dispersion Mean	Cluster Extent Mean	Ball Hall Index	Banfield Raftery Index	C Index	Determinant Ratio Index	Global Pattern
TCGA-33-AASL	LUSC	26245	20.6	40	656.1	293456	41.0	447	159518	0.015	2065.4	Brisk Diffuse
TCGA-D3-A2JF	SKCM	6832	4.9	18	379.6	238600	82.1	771	43456	0.022	790.0	Brisk Band-like
TCGA-E9-A22H	BRCA	1000	1.5	10	100.0	54876	51.9	560	6174	0.025	343.0	Non-Brisk Multifocal
TCGA-EW-A10X	BRCA	285	0.1	2	142.5	430332	223.0	3093	2283	0.000	29.6	Non-Brisk Focal



Credit: Dong Wenjie / Moment / Getty

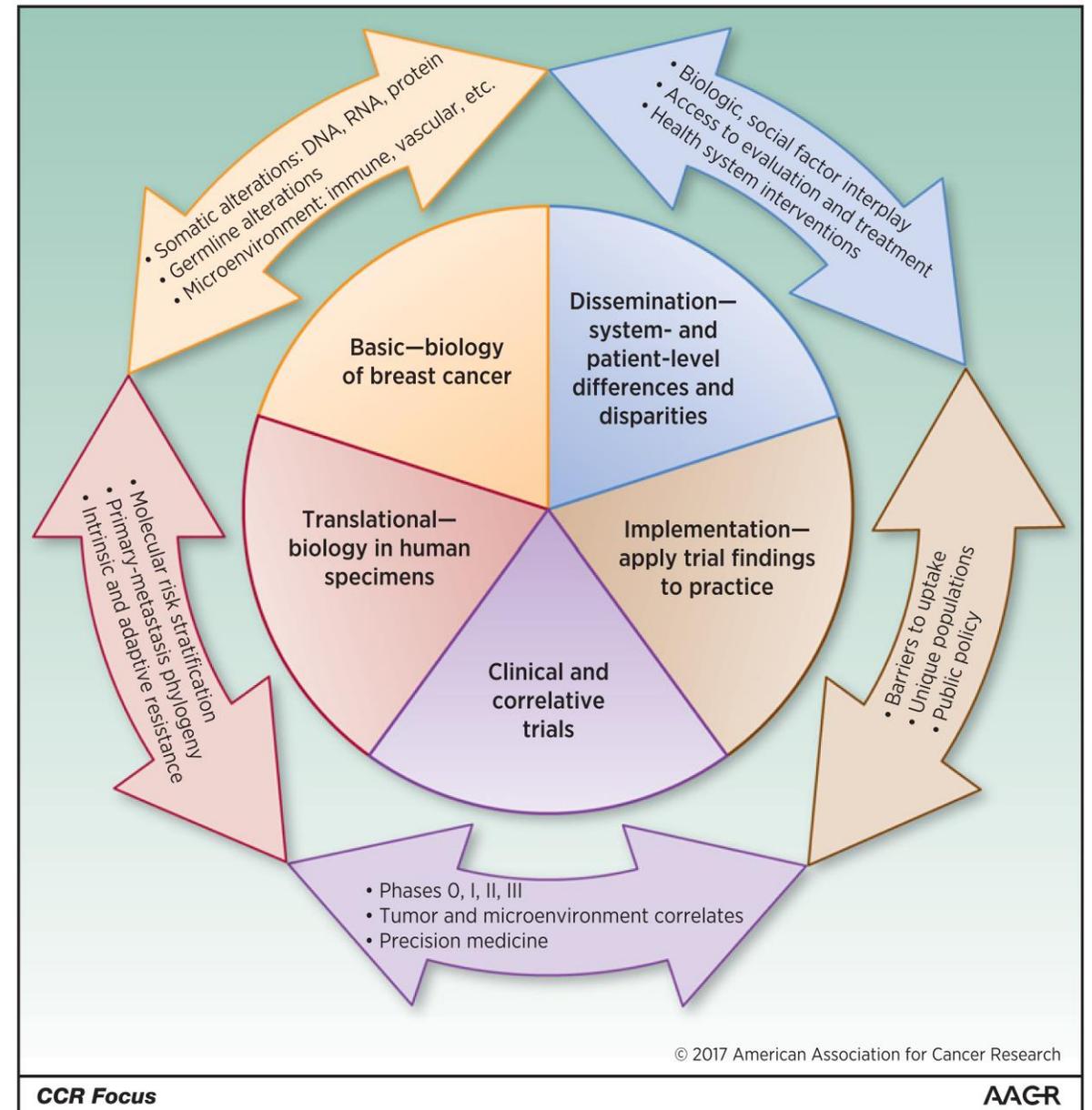
AI added to the curriculum for doctors-to-be

Medical schools and graduate research programs embrace artificial intelligence.

With the technological revolution of AI, may come an educational one: *medical researchers* will have to understand the basics of **artificial intelligence**, and, conversely, *computer scientists* will have to be trained to understand medicine

Conclusions

- Breast cancer is a **heterogeneous** disease
- **Inter- and intra- tumor heterogeneity**
→ Therapeutic and clinical implications
- **Translational research** allows a better understanding of breast cancer **biology** and of the mechanisms of **treatment resistance/sensitivity**
- **Biomarker** identification
→ Integration of multiple “omics” data
- These findings can be applied to **refine patient prognosis** (risk of relapse/progression) and to allow **treatment personalization** at a patient-level





Thank you

Merci

Grazie

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