

**The bases of clinical oncology: from bench to  
bedside**

**Prof. Annarosa Arcangeli**

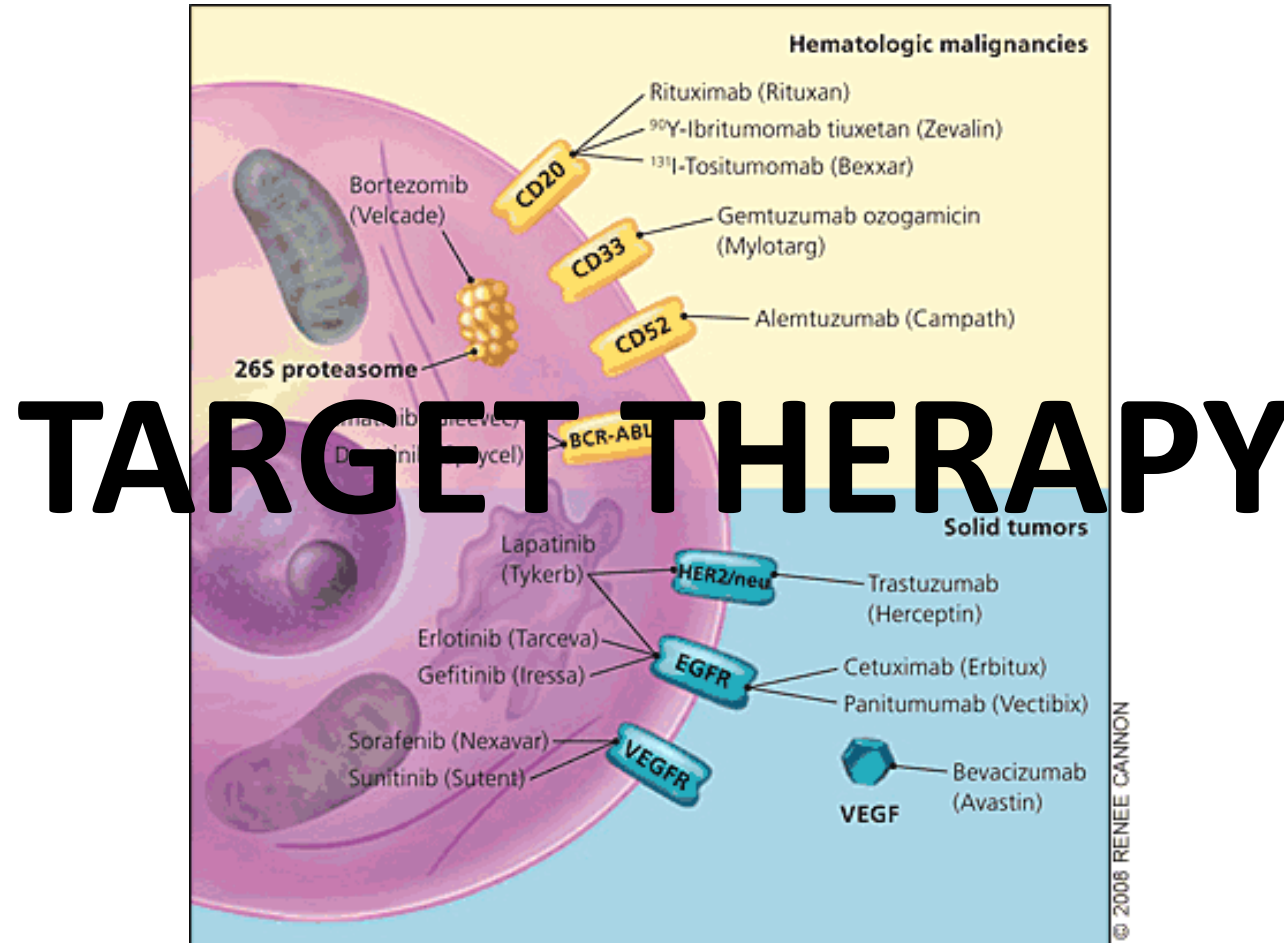
**Department of Experimental and Clinical Medicine**

**Erasmus course Basic & Translational Oncology**

**In collaboration with Université de Paris**

**Firenze, 20-24 January 2020**

# How can we translate basic science to the medical practice in oncology?



*Not All Patients are the Same*

Favorable prognosis  
Favorable response

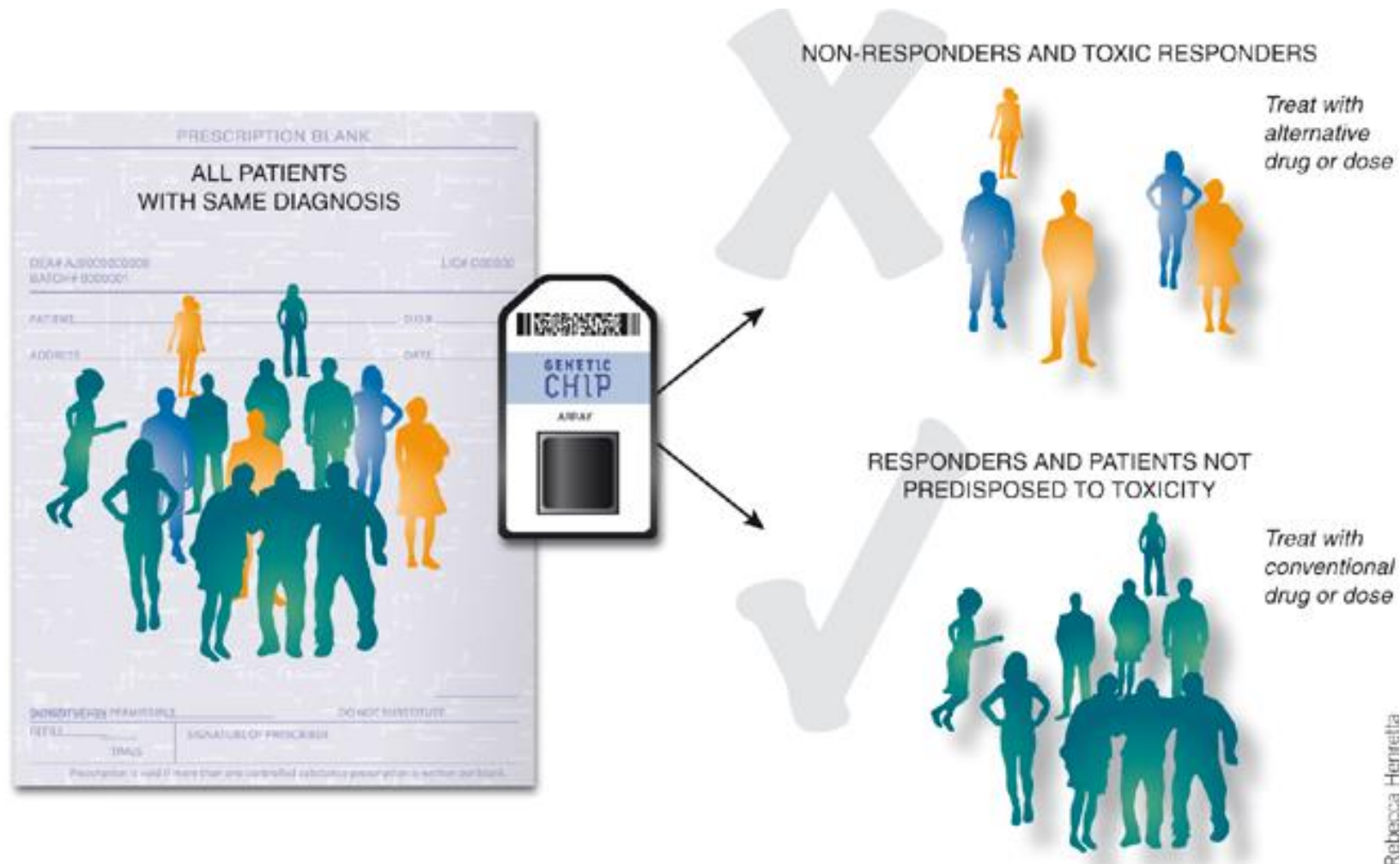
Unfavorable prognosis  
Unfavorable response

Increased toxicity

# Personalized medicine

## Biomarkers

### Predictors of benefits



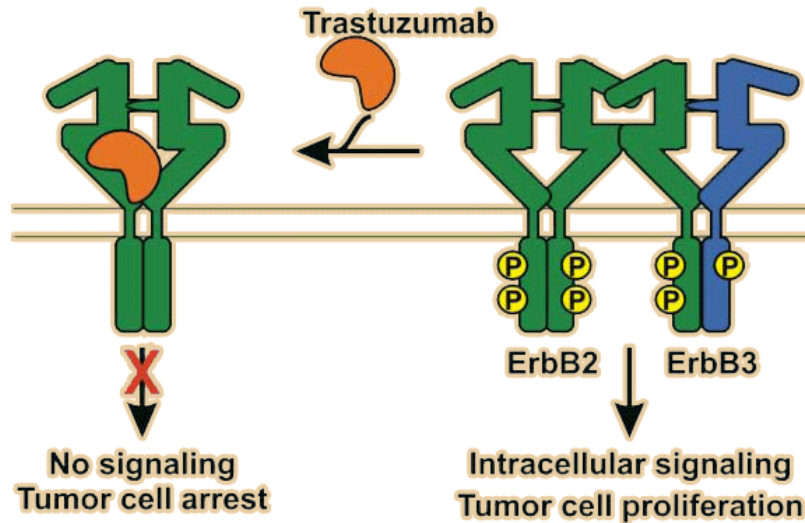
# Some examples

- **HER2**
- **K-Ras**
- **BCR-ABL (9:22 translocation)**

# HER2/neu

- The *HER2/neu* gene encodes one of a family of human epidermal growth-factor receptors.
- This gene is frequently amplified in breast cancer cells, resulting in increased amounts of HER2 cell surface protein.
- HER2-expressing tumors are sensitive to **Transtuzumab (herceptin)**, a monoclonal antibody therapy.
- HER2 protein is detected by immunohistochemistry (IHC).
- *HER2/neu* gene amplification is detected by fluorescence in situ hybridization (FISH).
- **Patients carrying a HER2/neu positive cancer (e.g. Breast Cancer) can be treated with Transtuzumab (Herceptin)**

# Herceptin



*Badache and Hynes, Cancer Cell 2004*

Trastuzumab is a humanized monoclonal antibody anti-ErbB-2

Efficacy on primary tumors with ErbB-2 amplification:

- Inhibits angiogenesis
- induces cytotoxicity
- increase response to chemotherapy
- Inhibits the activation of ErbB-2

Metastatic tumors develop resistance to Herceptin within 12 months



Increase of PI3K activation

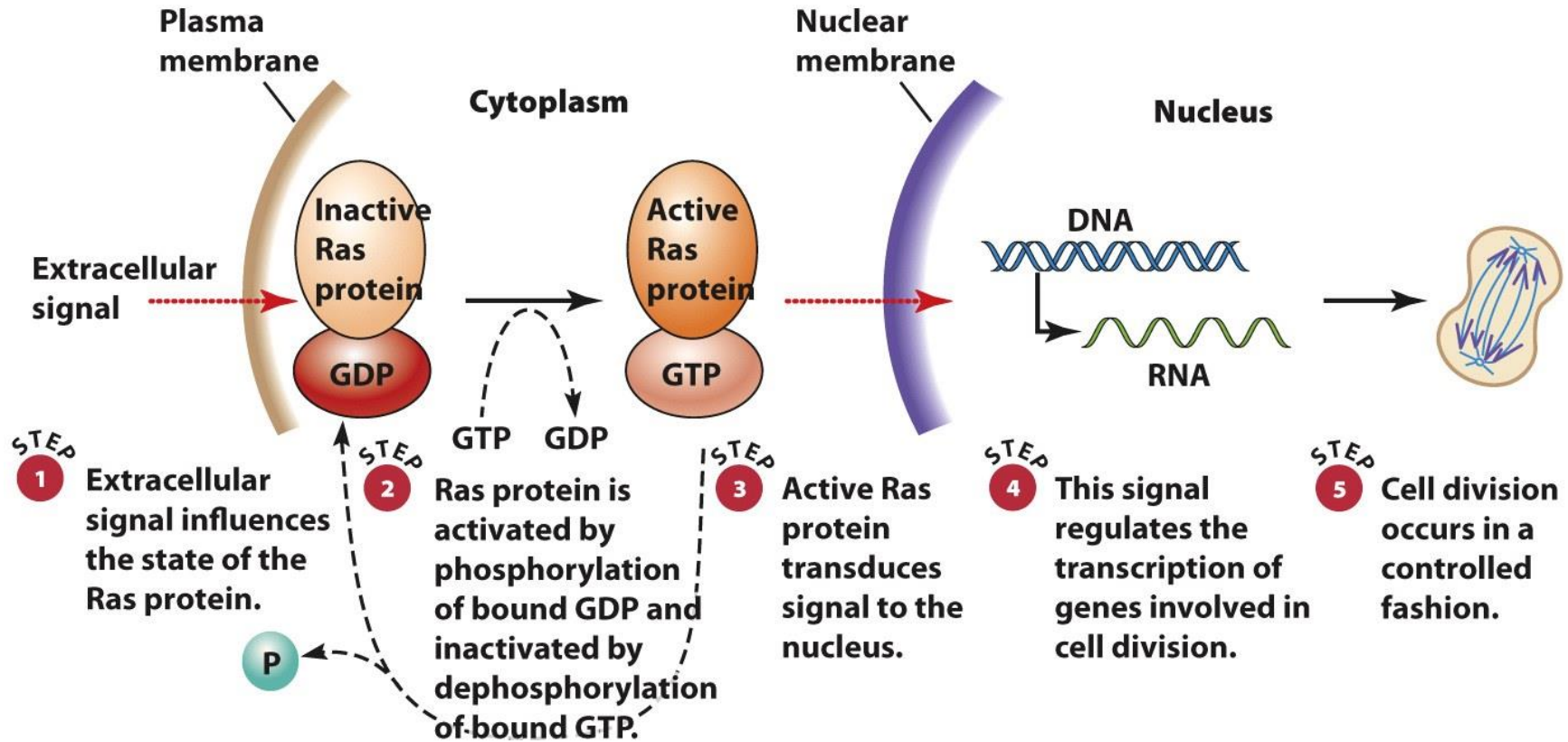
# *K-ras*

- The Kirsten rat sarcoma viral oncogene (*K-ras*) encodes a key component of cell signaling.
- Mutations in *K-ras* are the most common oncogene mutations in cancer.
- *K-ras* mutations are associated with tumor malignancy and may affect response to some therapies.
- *K-ras* gene mutations are detected by SSCP or direct sequencing.



# Normal Ras Protein Signaling

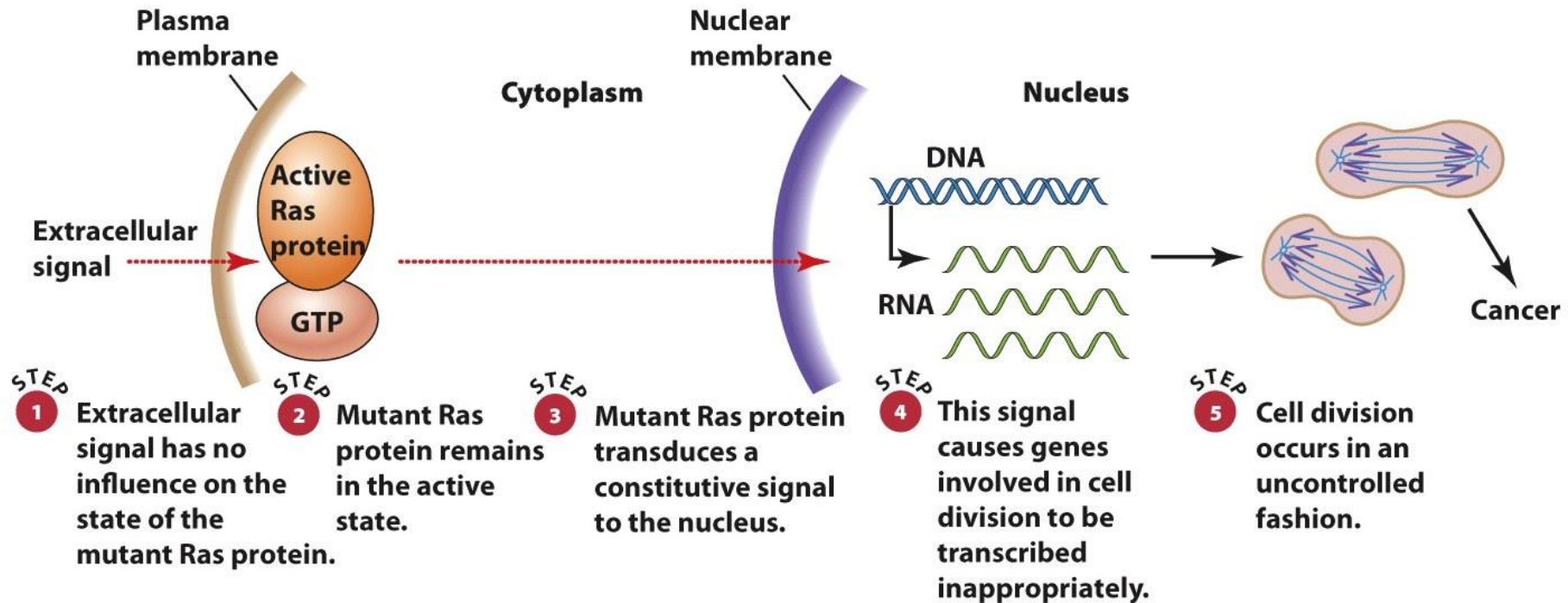
## Normal Ras protein is regulated



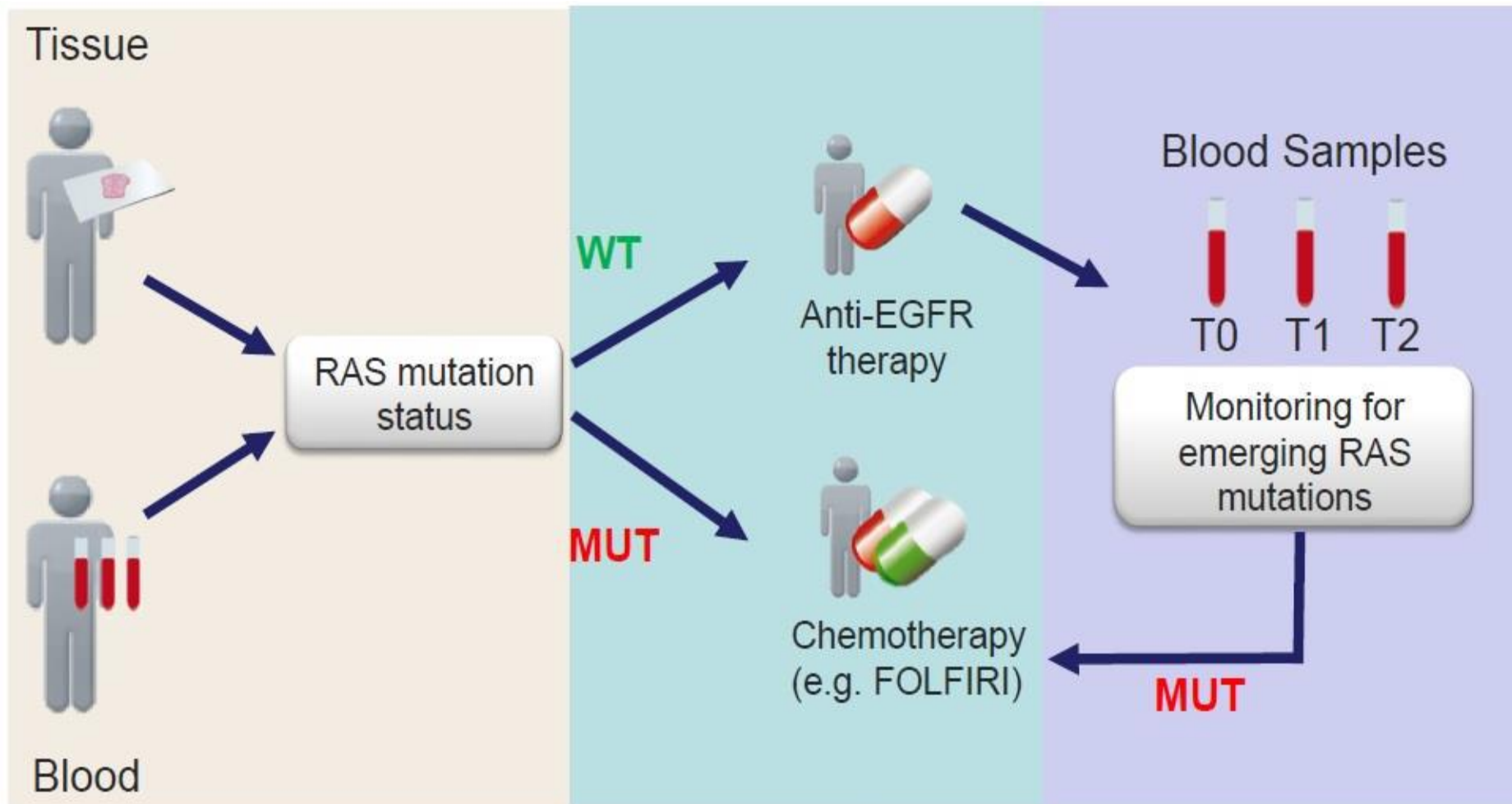
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# Mutant Ras Protein (V12 or G12V) is Unregulated

## Mutant Ras protein is unregulated



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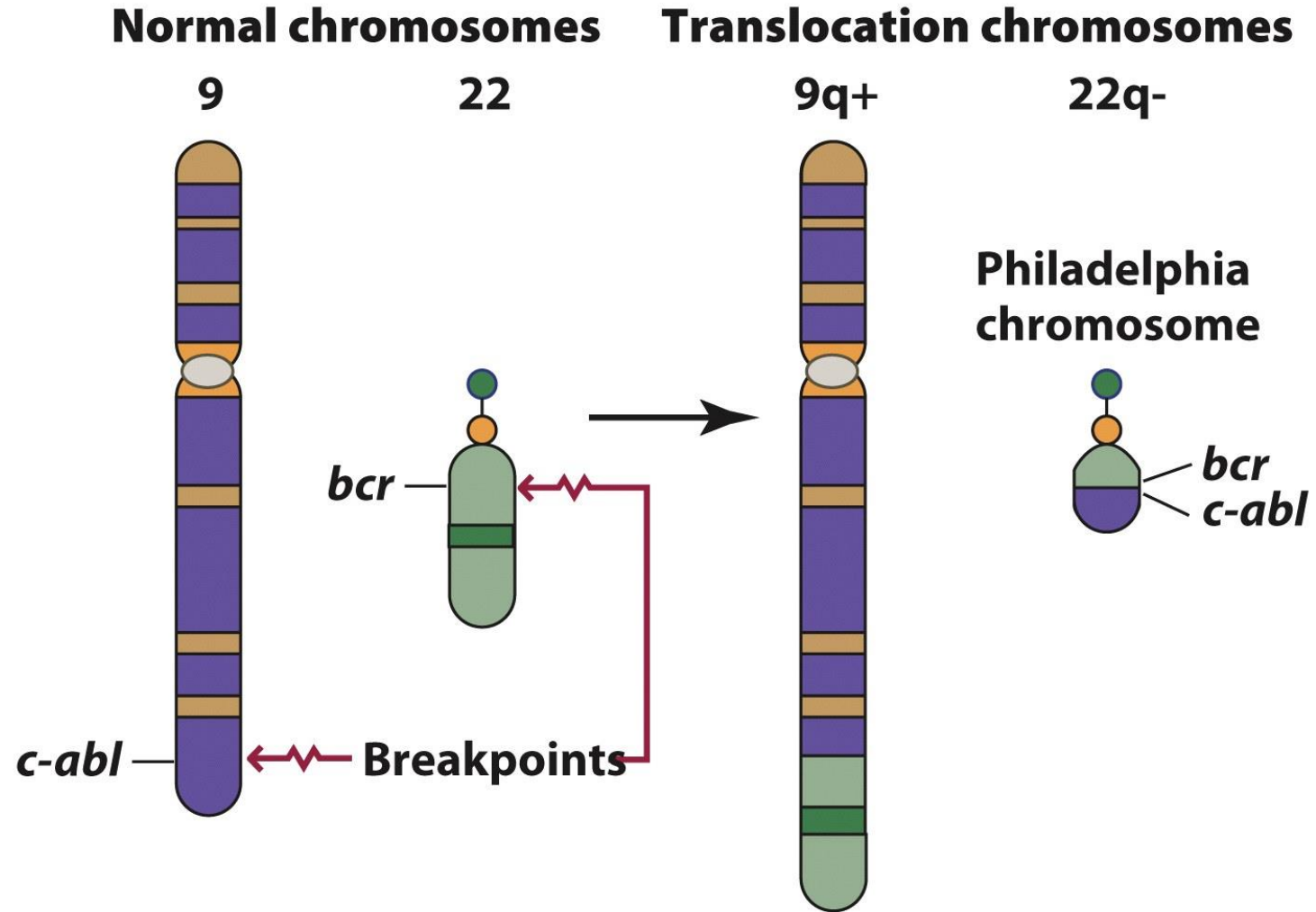


*Example: Blood-based RAS testing for colorectal cancer*

## Philadelphia Chromosome/t(9; 22)/ Bcr/abl

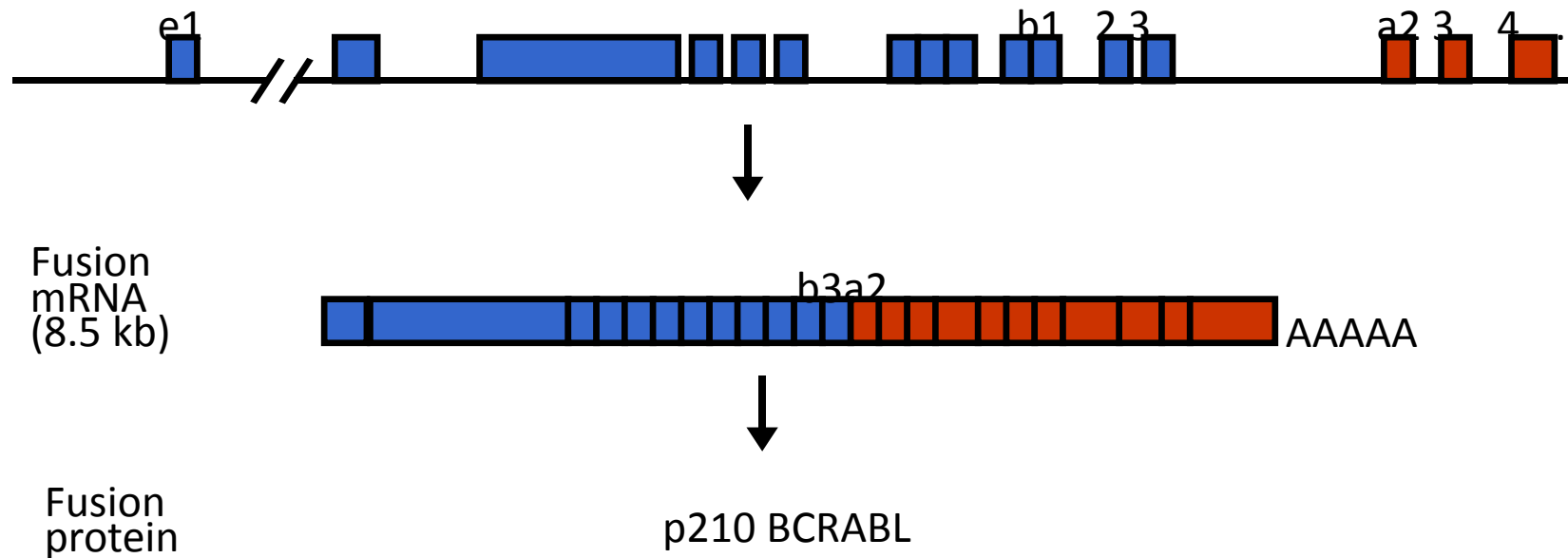
- t(9;22) is a reciprocal translocation between the long arms of chromosomes 9; 22 is found in **chronic myelogenous leukemia** and **acute lymphoblastic leukemia**.
- This translocation forms a chimeric gene between the breakpoint cluster region (**BCR**) gene on chromosome 22 and the Abelson leukemia virus (**ABL**) gene on chromosome 9.
- The translocated chromosome is the **Philadelphia chromosome**.

# The Philadelphia Chromosome

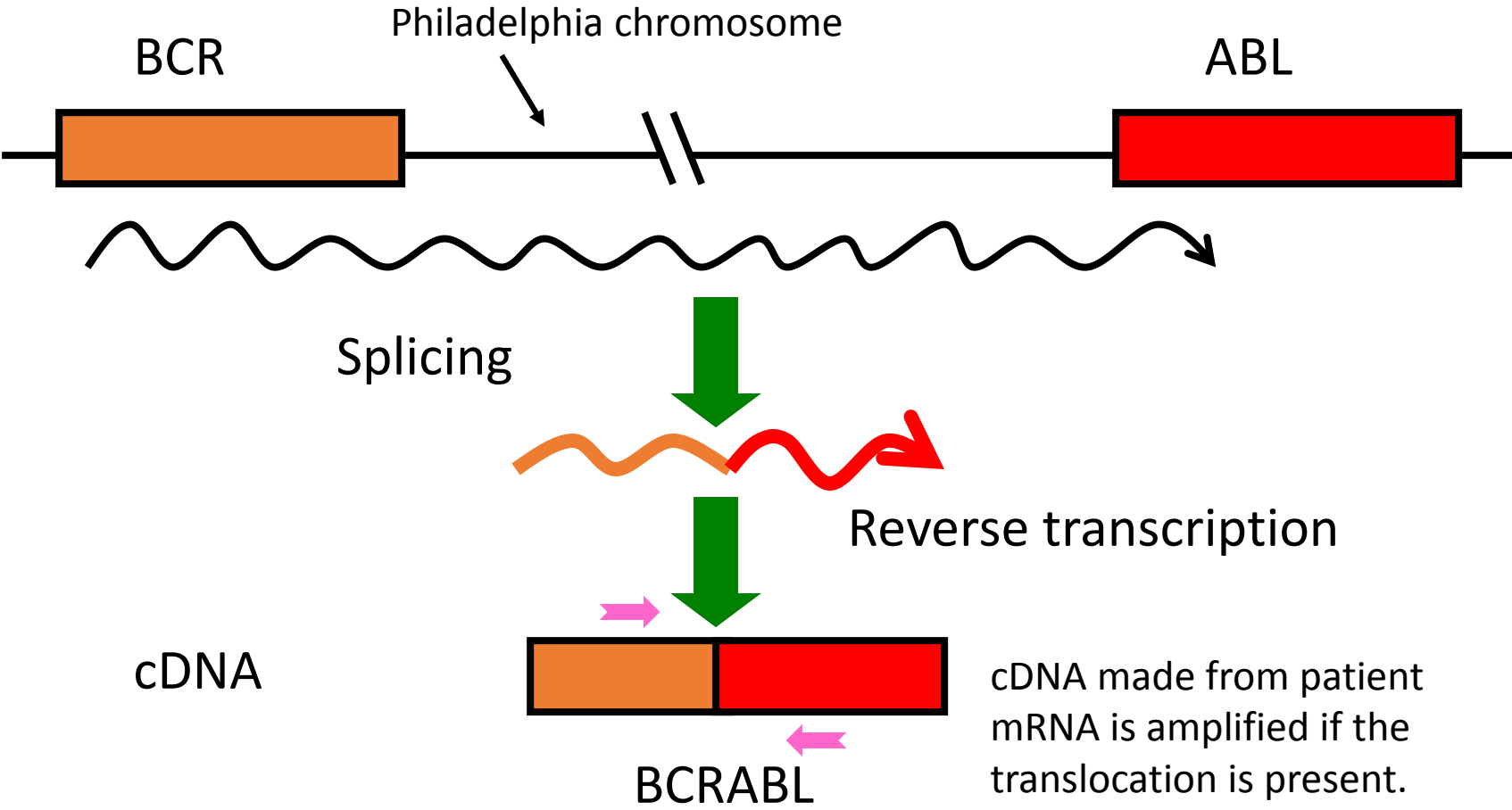


# Translocations Used in Diagnosis and Monitoring of Hematological Tumors: t(9; 22)

The chimeric gene, **BCRABL**, produces an abnormal protein that drives the tumor cell phenotype.



# Detection of t(9; 22) by RT-PCR

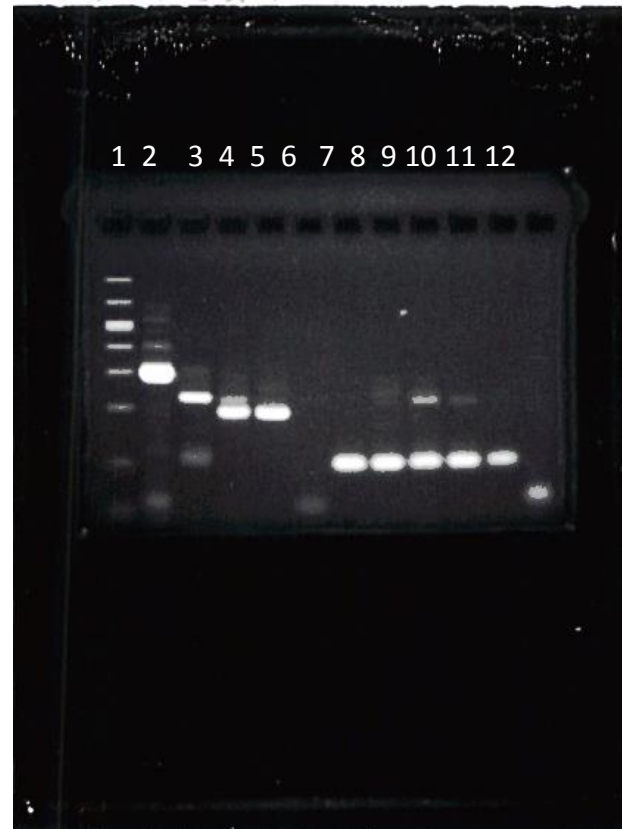


# Detection of t(9; 22) by RT-PCR

1 = molecular weight  
standard  
2-5 = positive for  
translocation  
6 = negative  
7-11 = amplification controls  
12 = blank

Translocation  
products  
(BCRABL)

The band size is  
determined by  
different  
chromosome 22  
breakpoints.

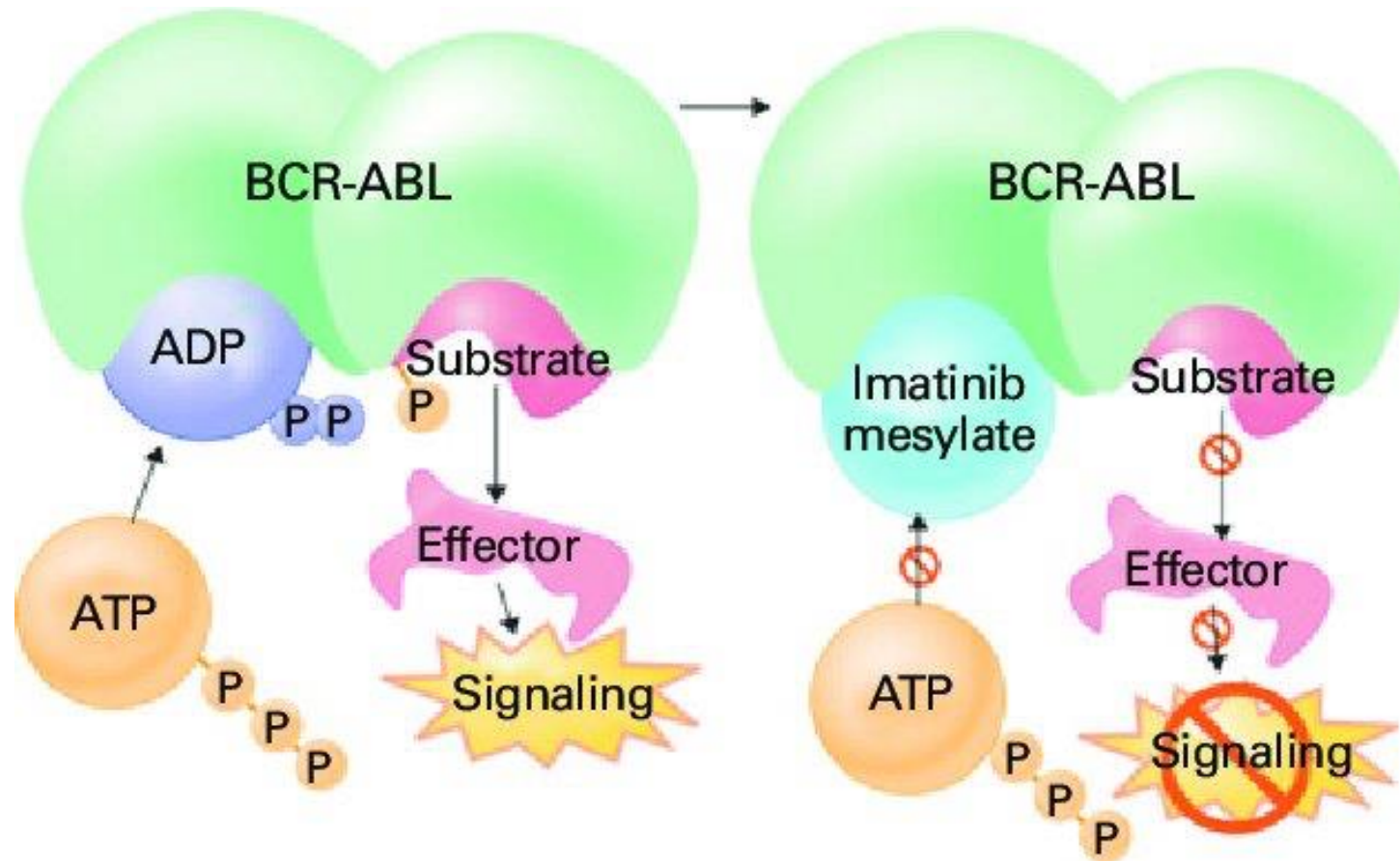


Agarose gel

Translocation  
products  
(ABL)



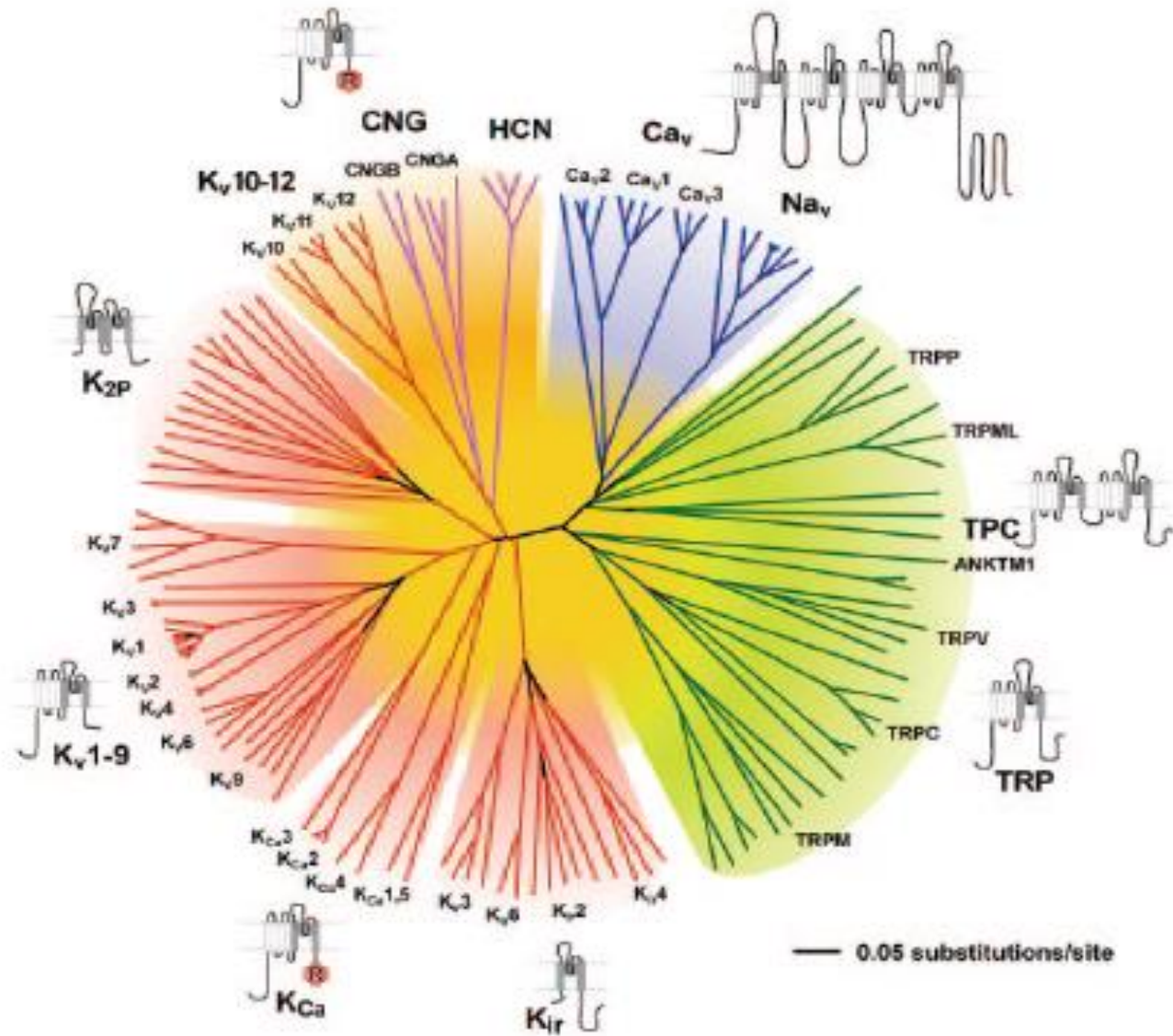
# IMATINIB

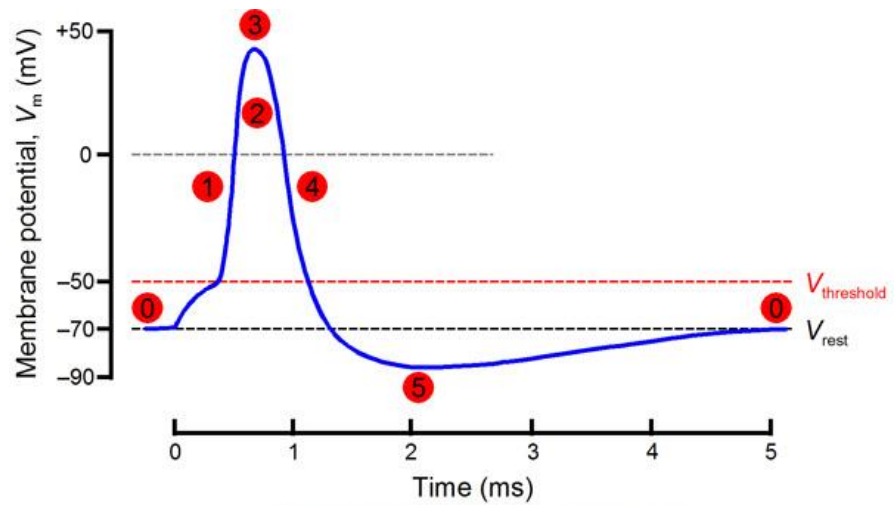


**How can we translate basic science in  
ion channels to the medical practice in  
oncology?**

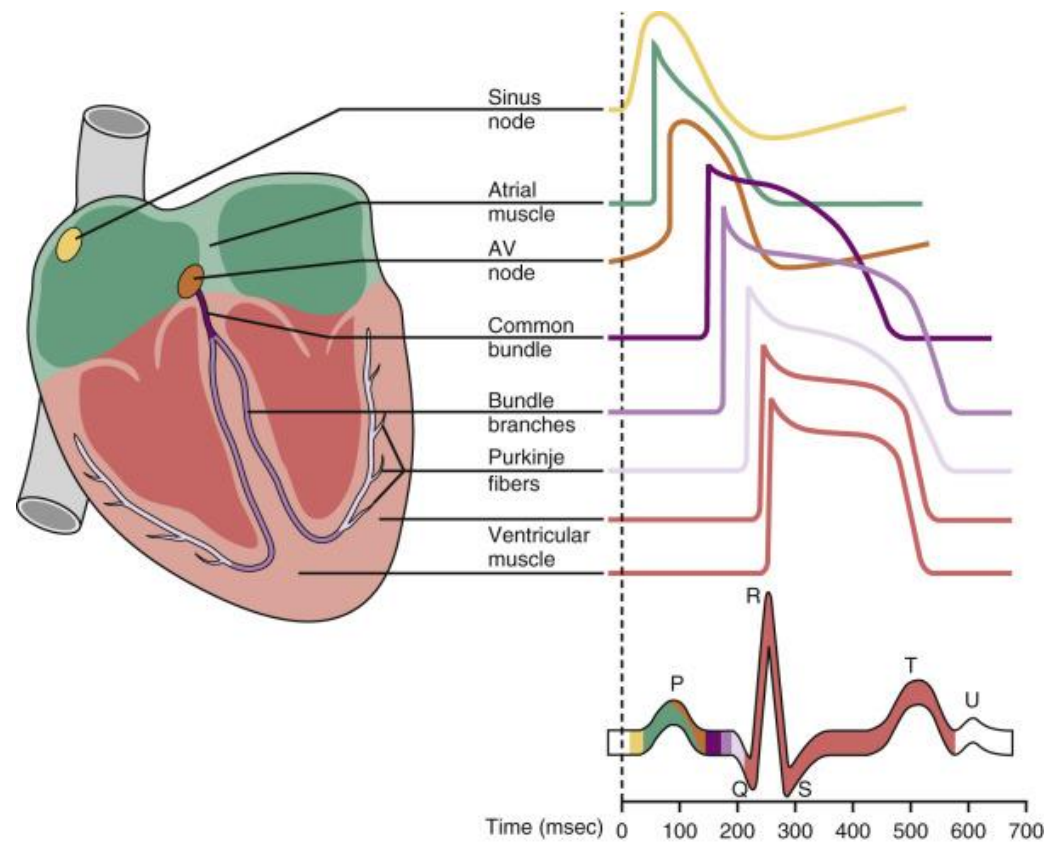
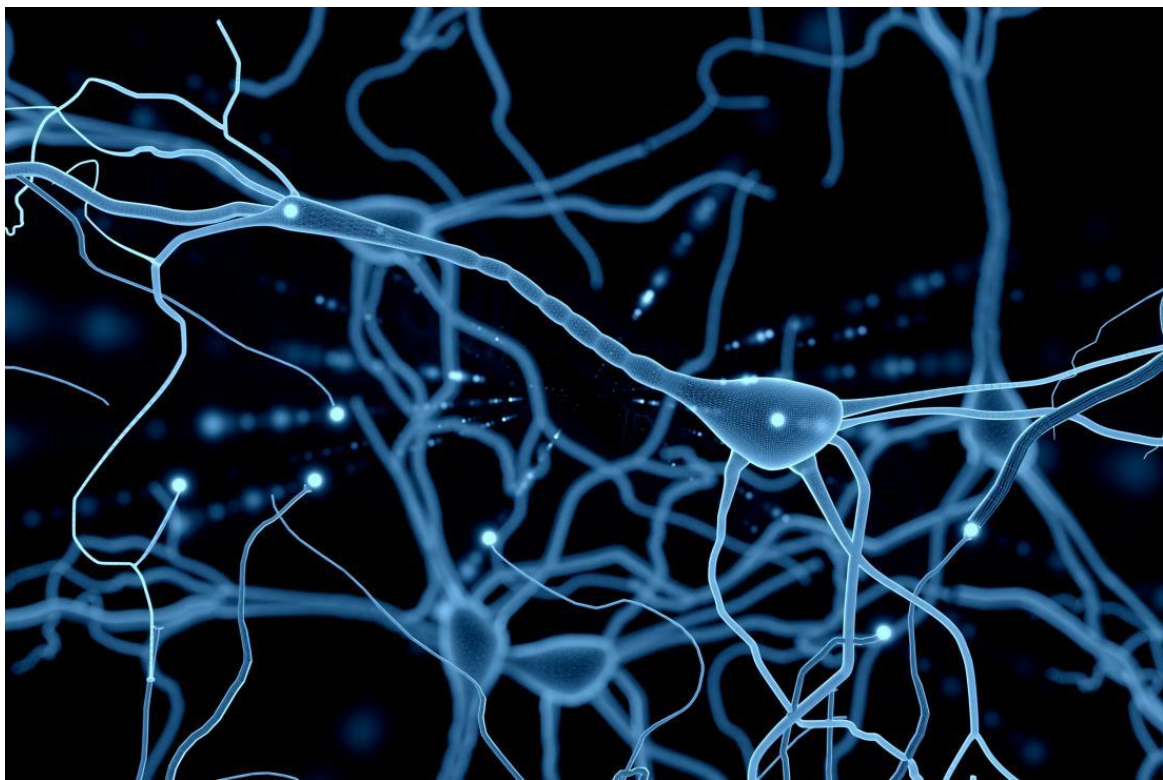
# **ION CHANNELS IN CANCER**

# ION CHANNELS

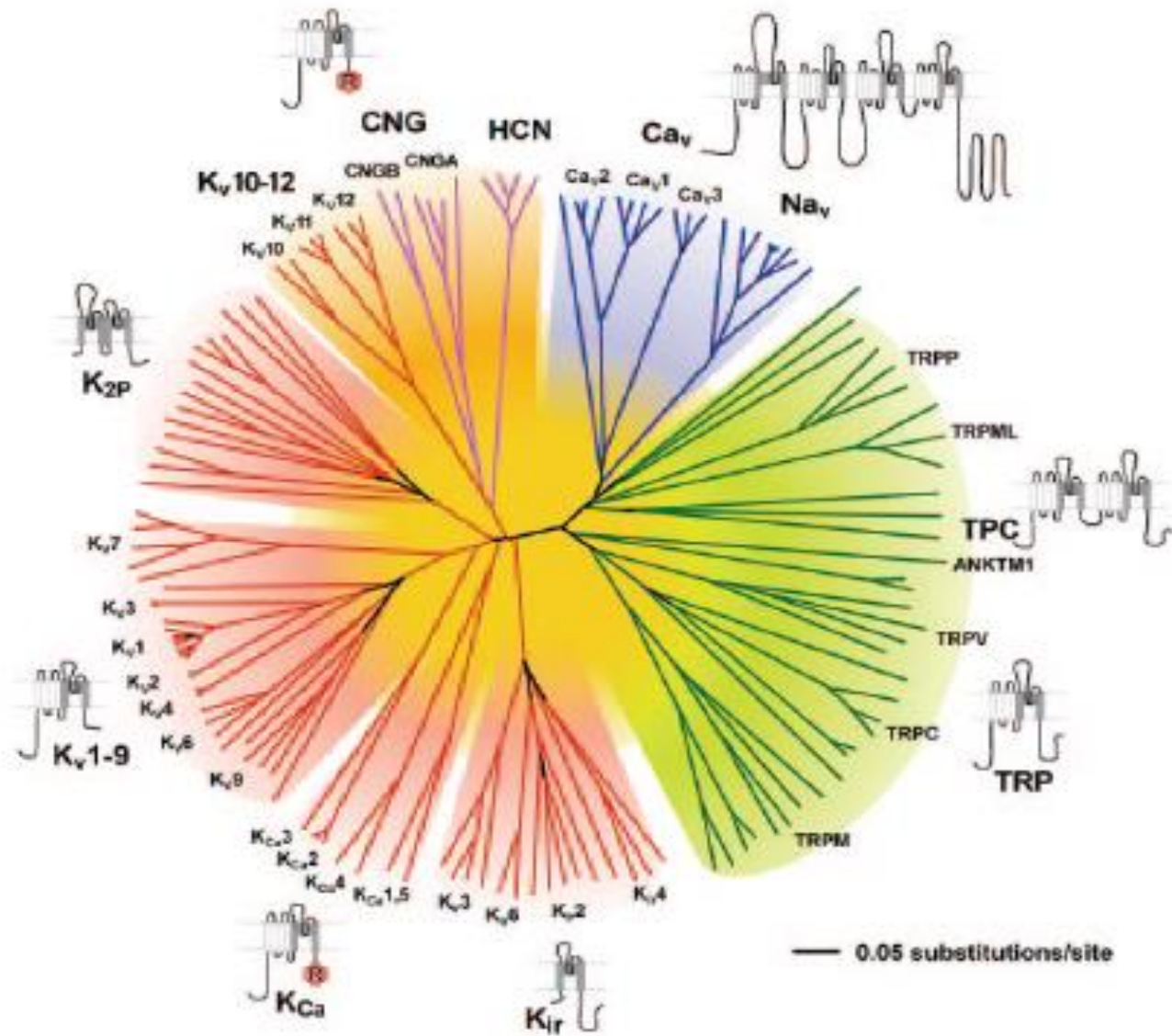




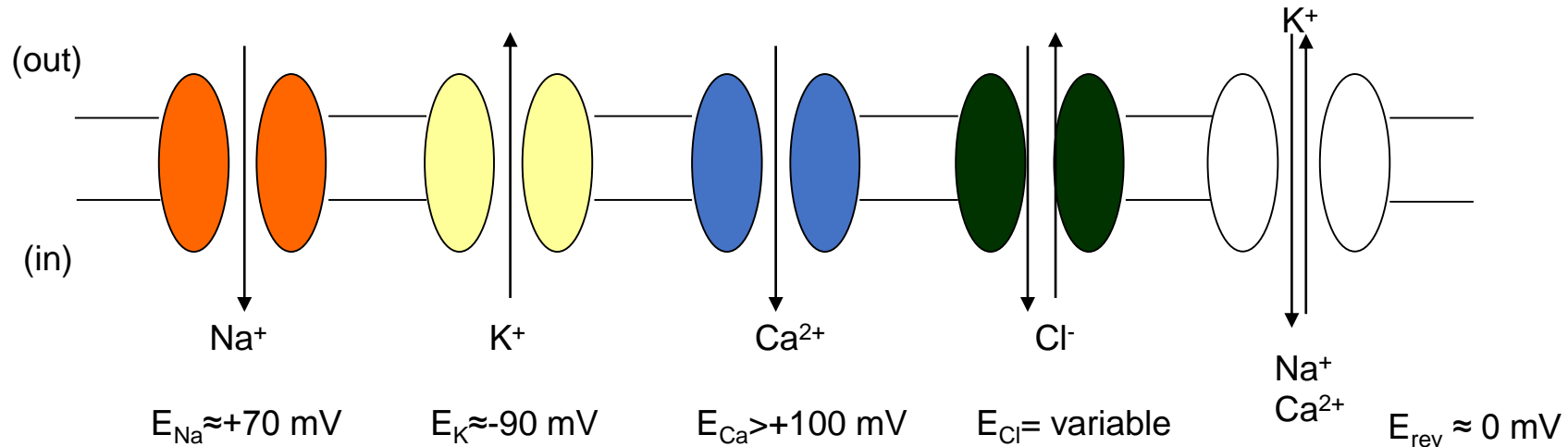
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# ION CHANNELS



## Ion channels and membrane potential ( $V_m$ )



An ion channel highly selective for a given ion type  $i$  tends to bring  $V_m$  close to the equilibrium (Nernst) potential ( $E_i$ ) for that ion, at which the net flux of  $i$  is null.

$E_{\text{K}}$  is around  $-90 \text{ mV}$ , in physiological  $[\text{K}]_o/[\text{K}]_i$ . When  $V_m = E_{\text{K}}$ , the  $\text{K}^+$  outflow driven by the concentration gradient is balanced by the influx driven by the negative  $V_m$ .

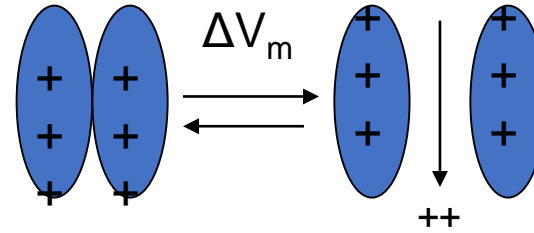
At  $V_m$ 's negative of  $E_{\text{K}}$ ,  $\text{K}^+$  flows into the cell.

At  $V_m$ 's positive of  $E_{\text{K}}$ ,  $\text{K}^+$  flows out of the cell

# Gating of ion channels

Closed  $\rightleftharpoons$  Open

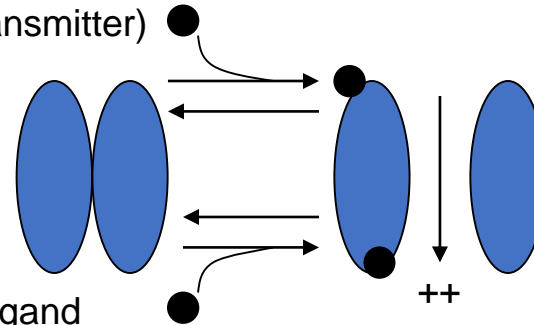
Voltage-gated channels



Ligand-gated channels

Extracellular ligand  
(e.g. neurotransmitter)

Intracellular ligand  
(e.g. cyclic nucleotide)



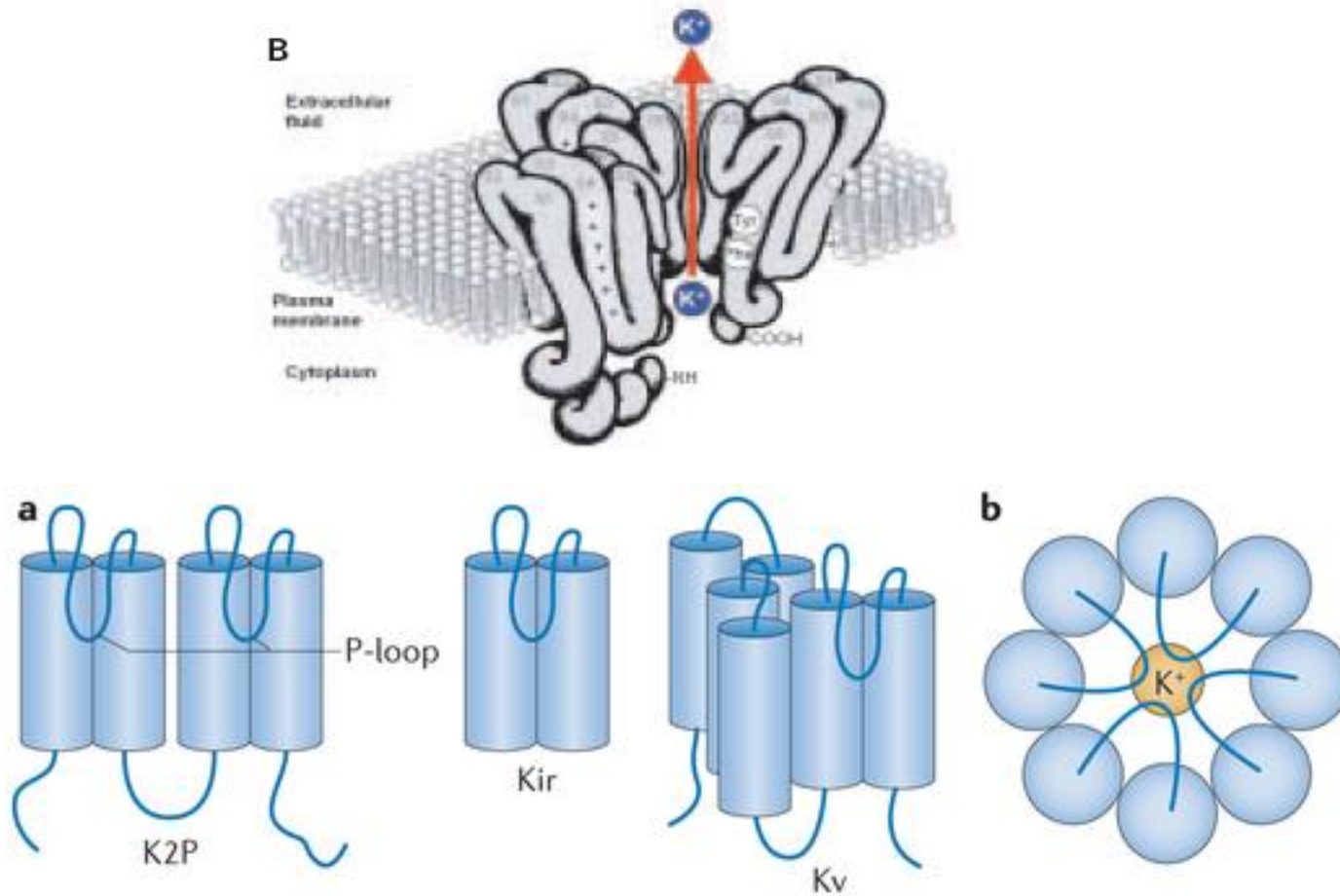
Other gating mechanisms:

mechanically-gated channels  
(cytoskeleton)

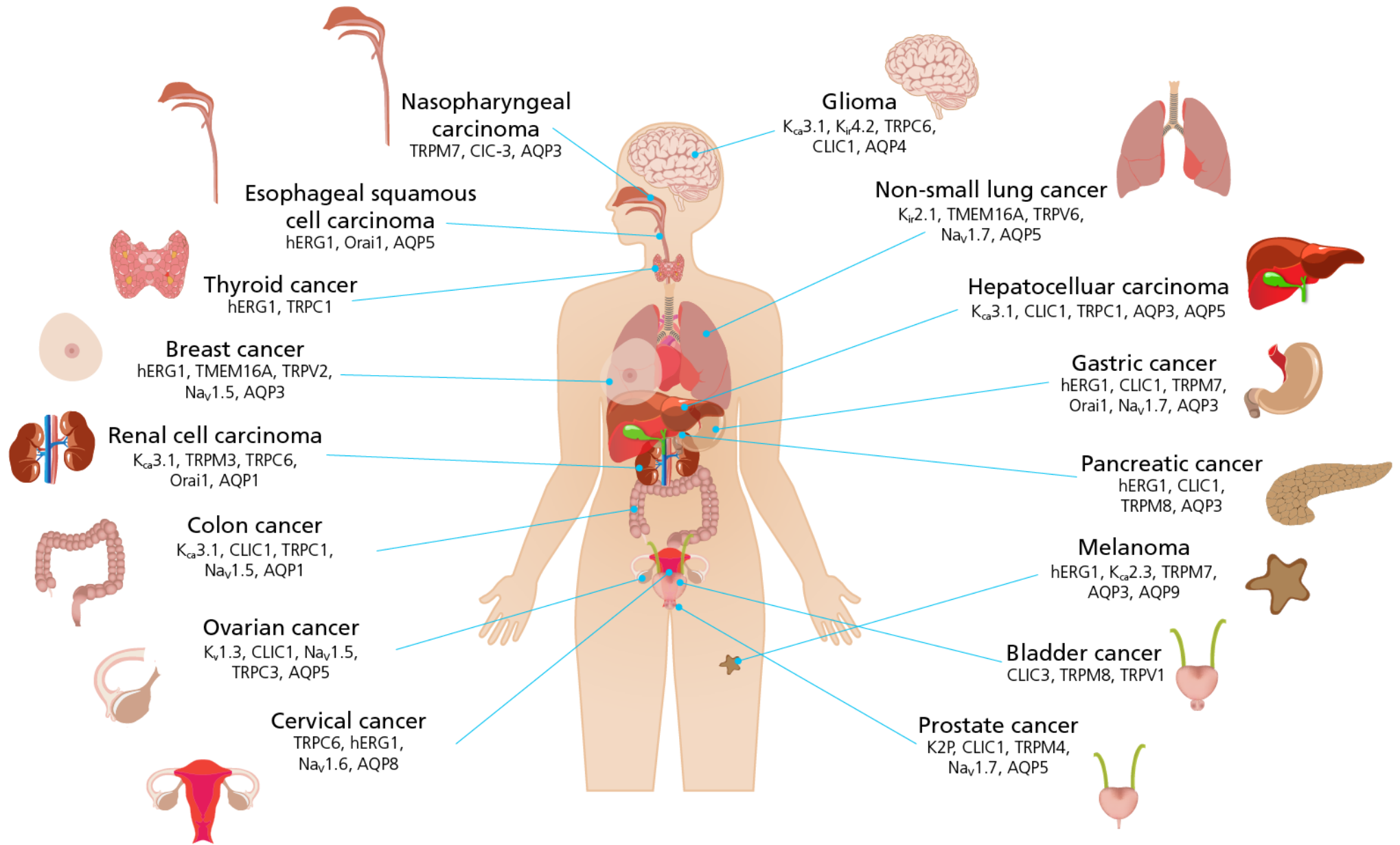
conformational coupling



# K<sup>+</sup> channels: structure



# **ION CHANNELS IN CANCER**



Review

## Ion channel expression as promising cancer biomarker<sup>☆</sup>

Elena Lastraioli, Jessica Iorio, Annarosa Arcangeli<sup>\*</sup>

Department of Experimental and Clinical Medicine, Section of Internal Medicine University of Florence, Florence Italy

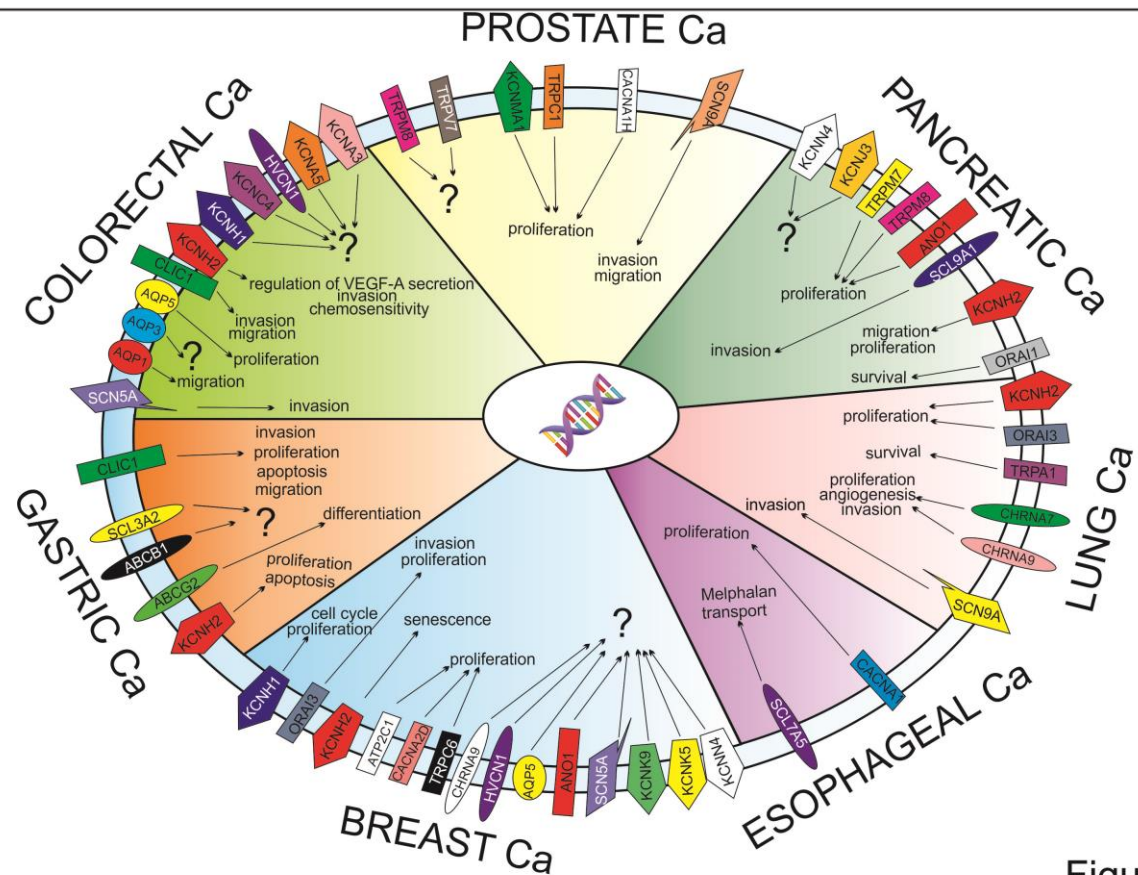
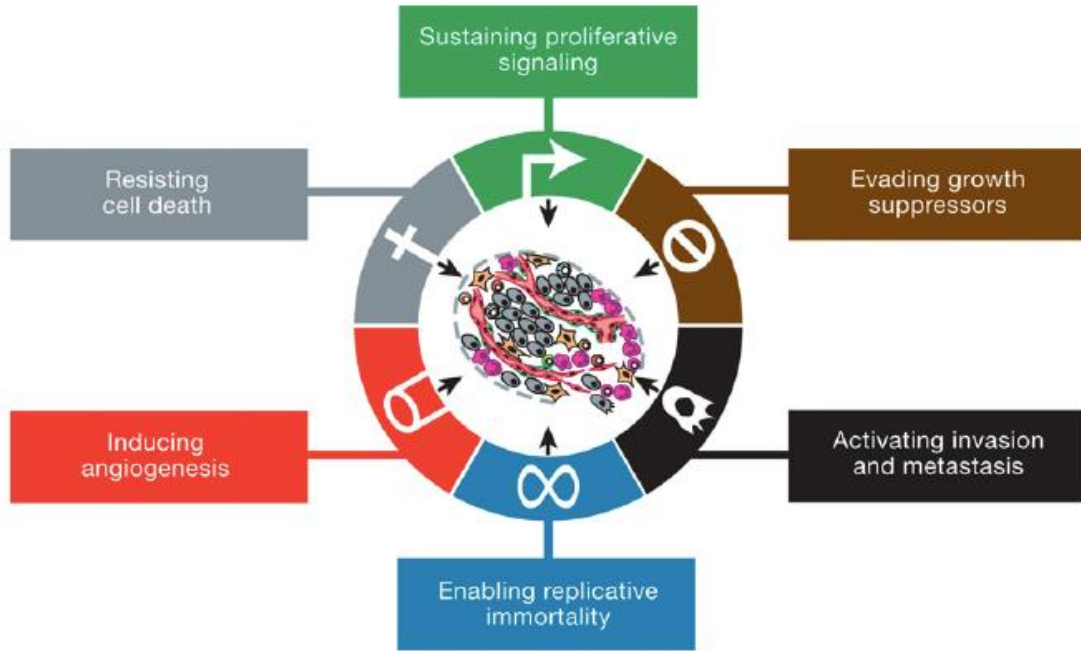


Figure 1

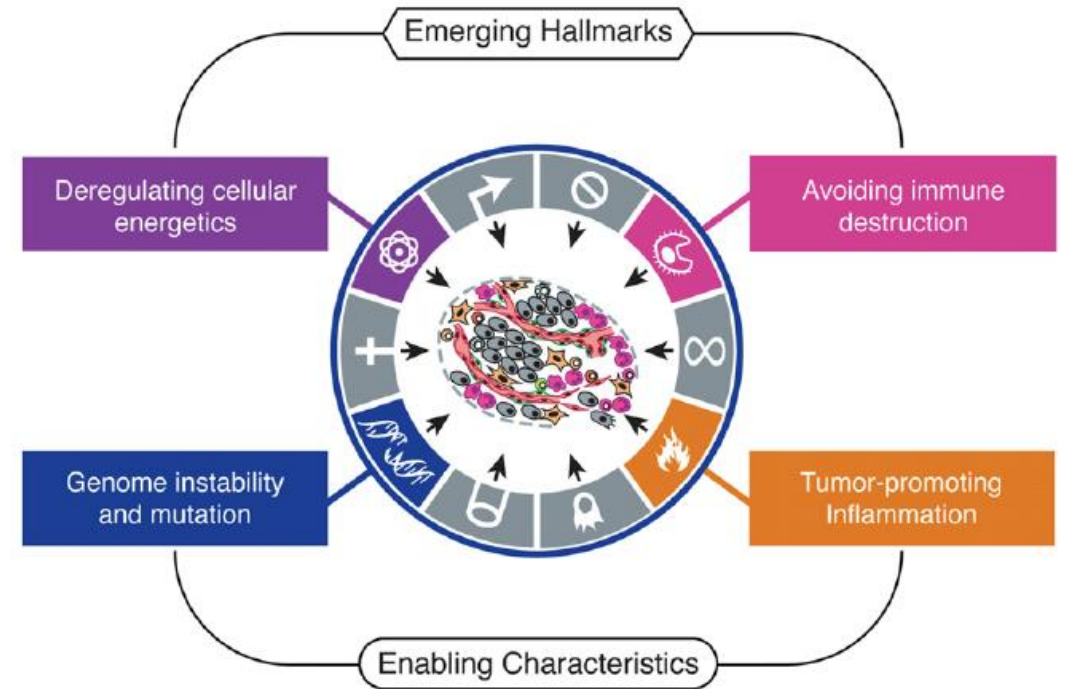
# Hallmarks of Cancer

Hanahan and Weimberg, Cell, 2000

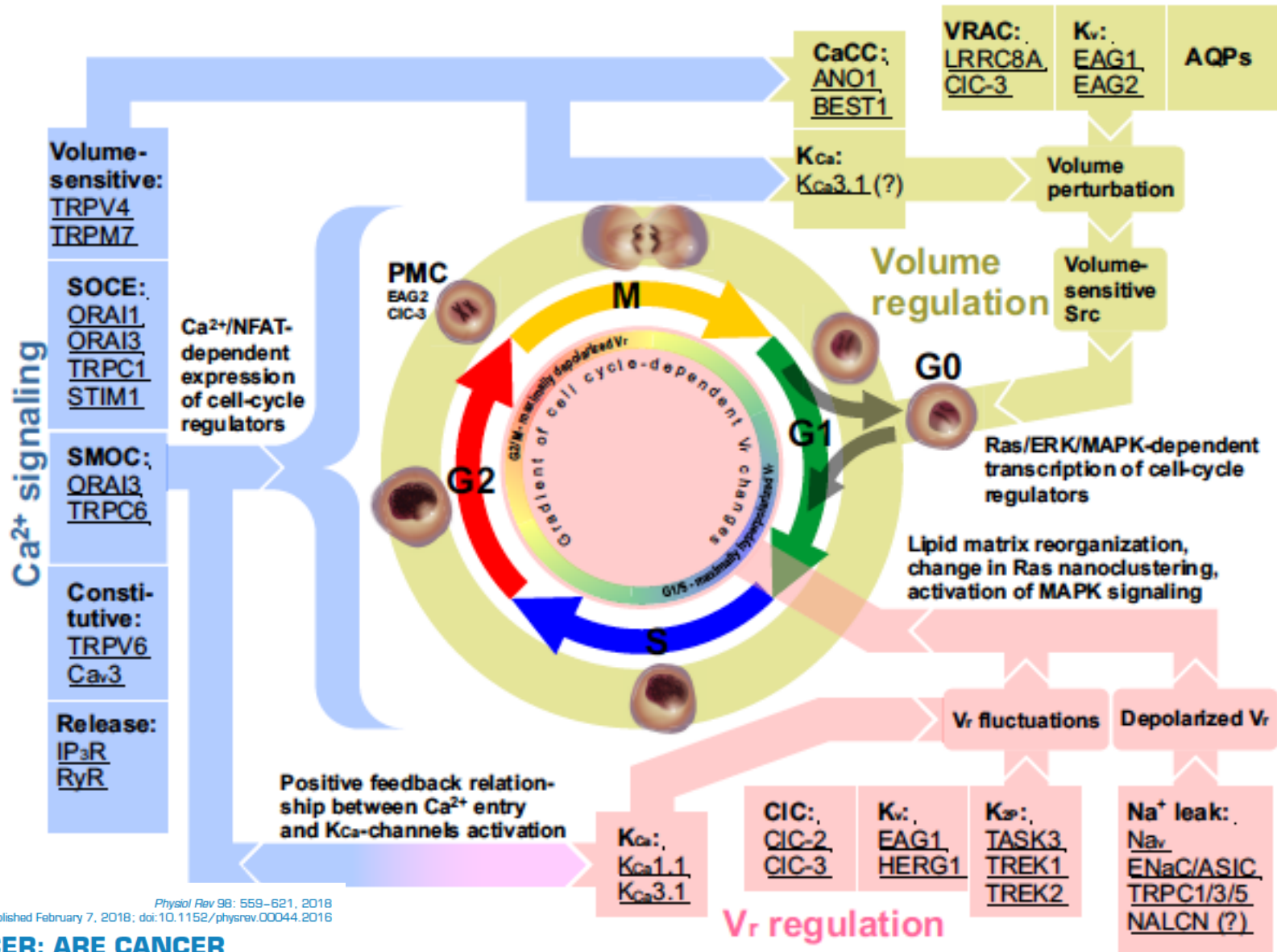


# Hallmarks of Cancer: The Next Generation

Hanahan and Weimberg, Cell, 2011



# Contribution of ion channels to proliferation and cell cycle progression of cancer cells



Physiol Rev 98: 559–621, 2018  
Published February 7, 2018; doi:10.1152/physrev.00044.2016

## ION CHANNELS IN CANCER: ARE CANCER HALLMARKS ONCOCHANNELOPATHIES?

## Some channel-mediated mechanisms in cell proliferation

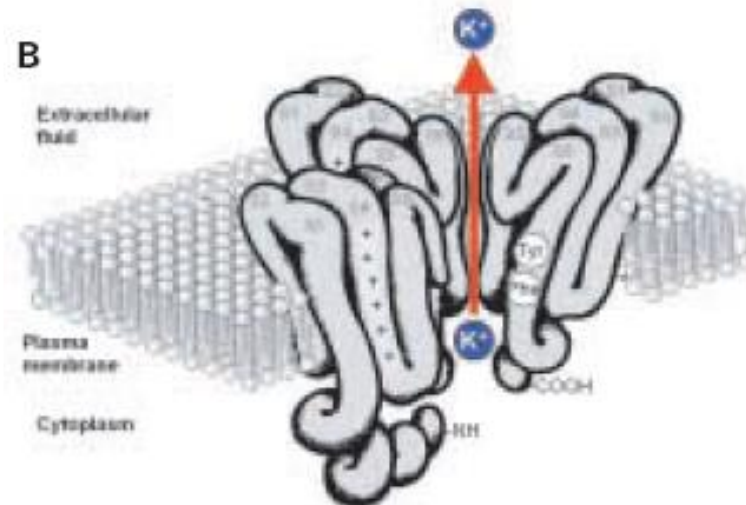
- Direct modulation of the cell cycle machinery (by e.g.,  $\text{Ca}^{2+}$ , pH).
- Depolarization +  $\text{Ca}^{2+}$  regulate exocytosis of hormones and paracrine factors (GFs, cytokines, etc.).
- Regulation of cell volume ( $\text{K}^+$  plus  $\text{Cl}^-$  channels, and KCC transporters for solute extrusion; mainly  $\text{Na}^+$  plus  $\text{Cl}^-$  channels, and NKCC transporters, for solute absorption).
- **Regulation of cell adhesion and GF release by non-conductive mechanisms (stimulation of intracellular cascades and conformational coupling with other membrane proteins)**

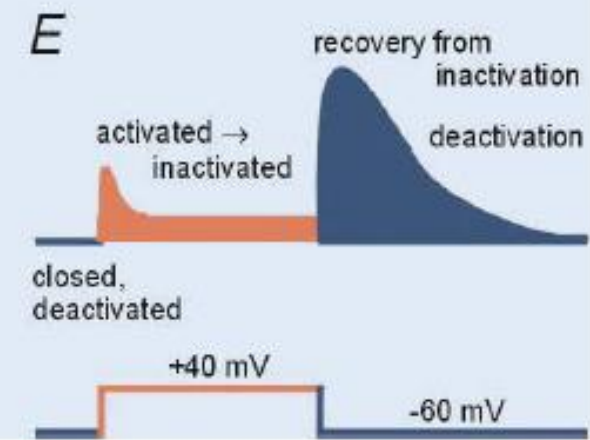
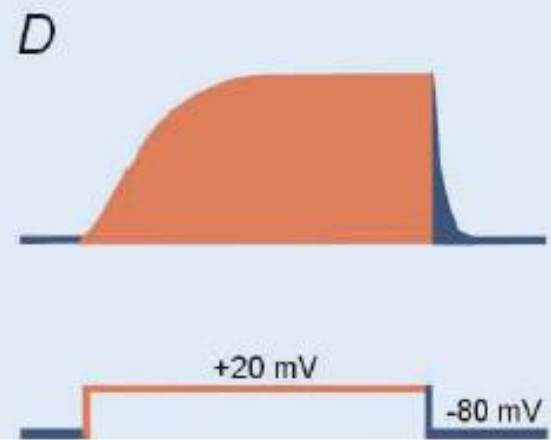
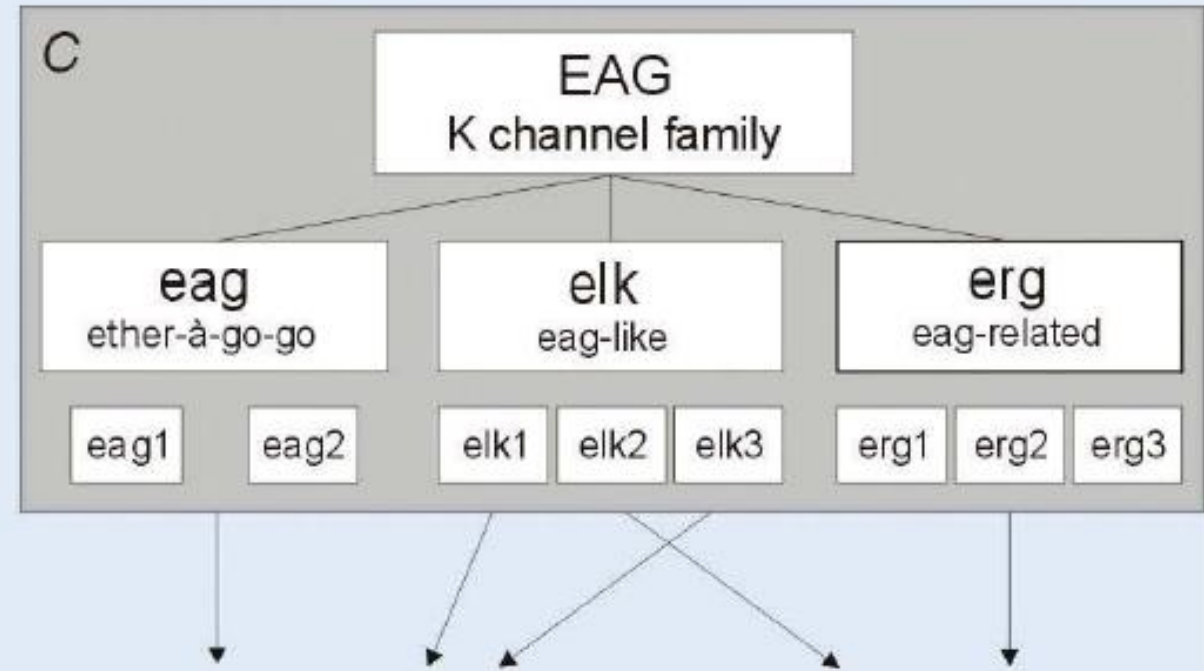
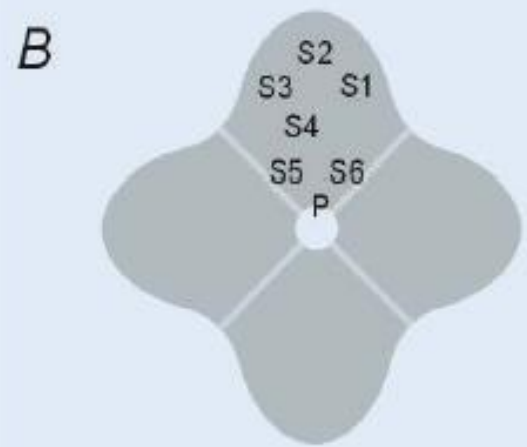
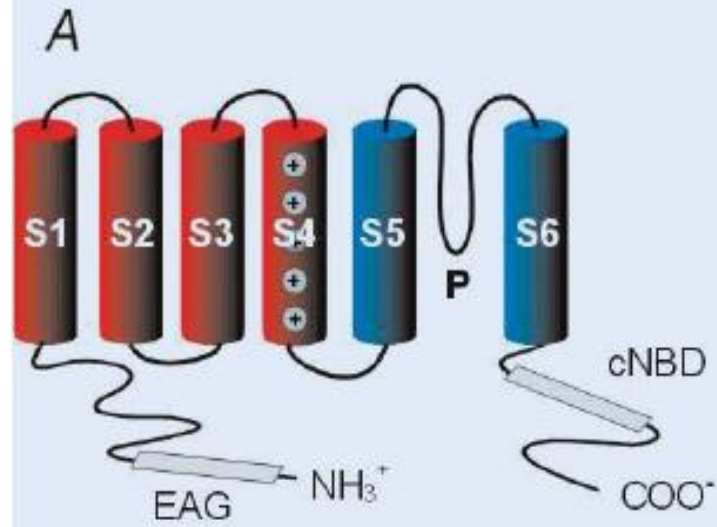
## Non conductive roles of ion channels

- Regulatory domains (e.g. kinase domains).
- **Formation and modulation of membrane multiprotein complexes (e.g. hERG or AMPA glutamate receptors with integrins).**
- Conformational coupling with cytosolic proteins (e.g. cytoskeleton).

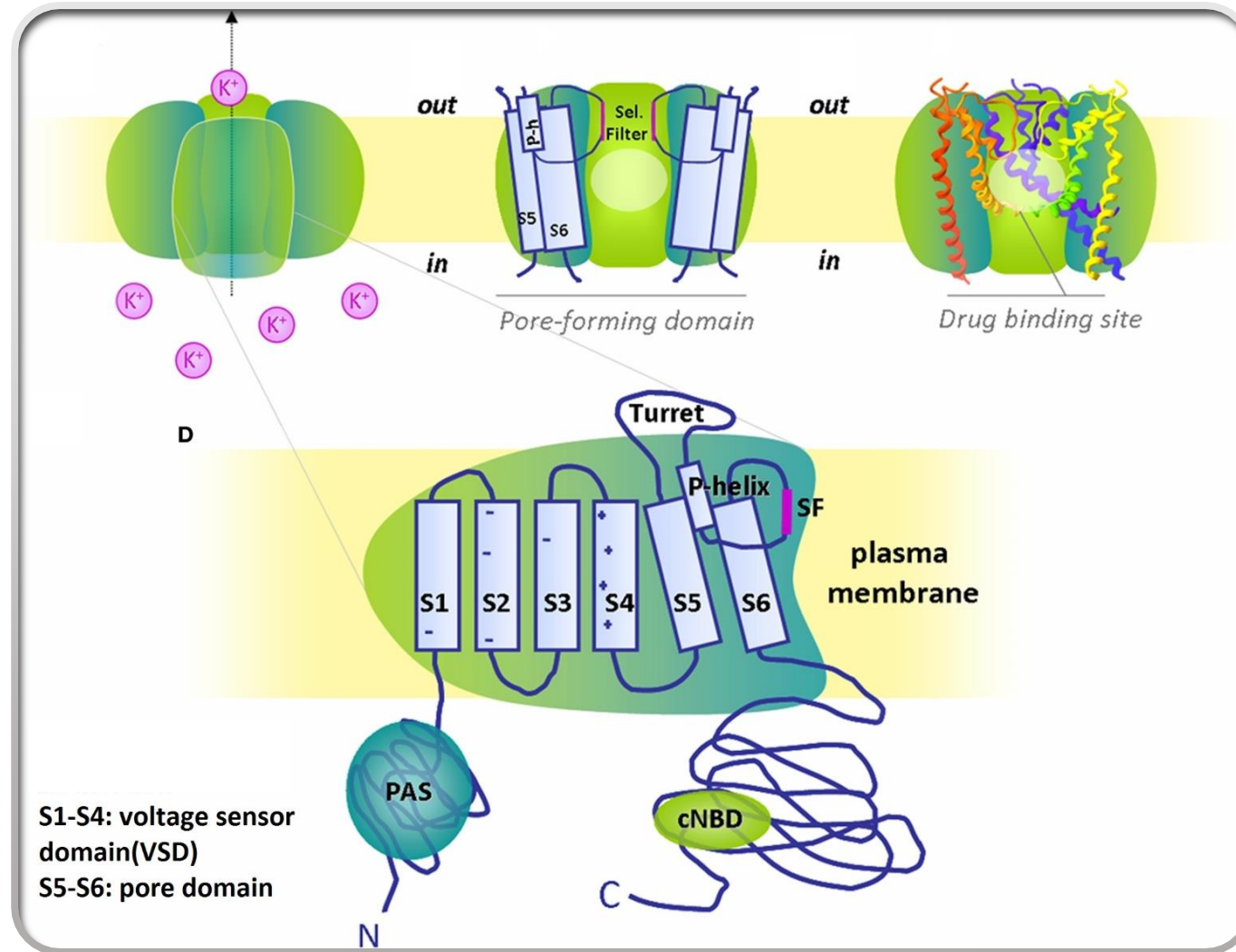


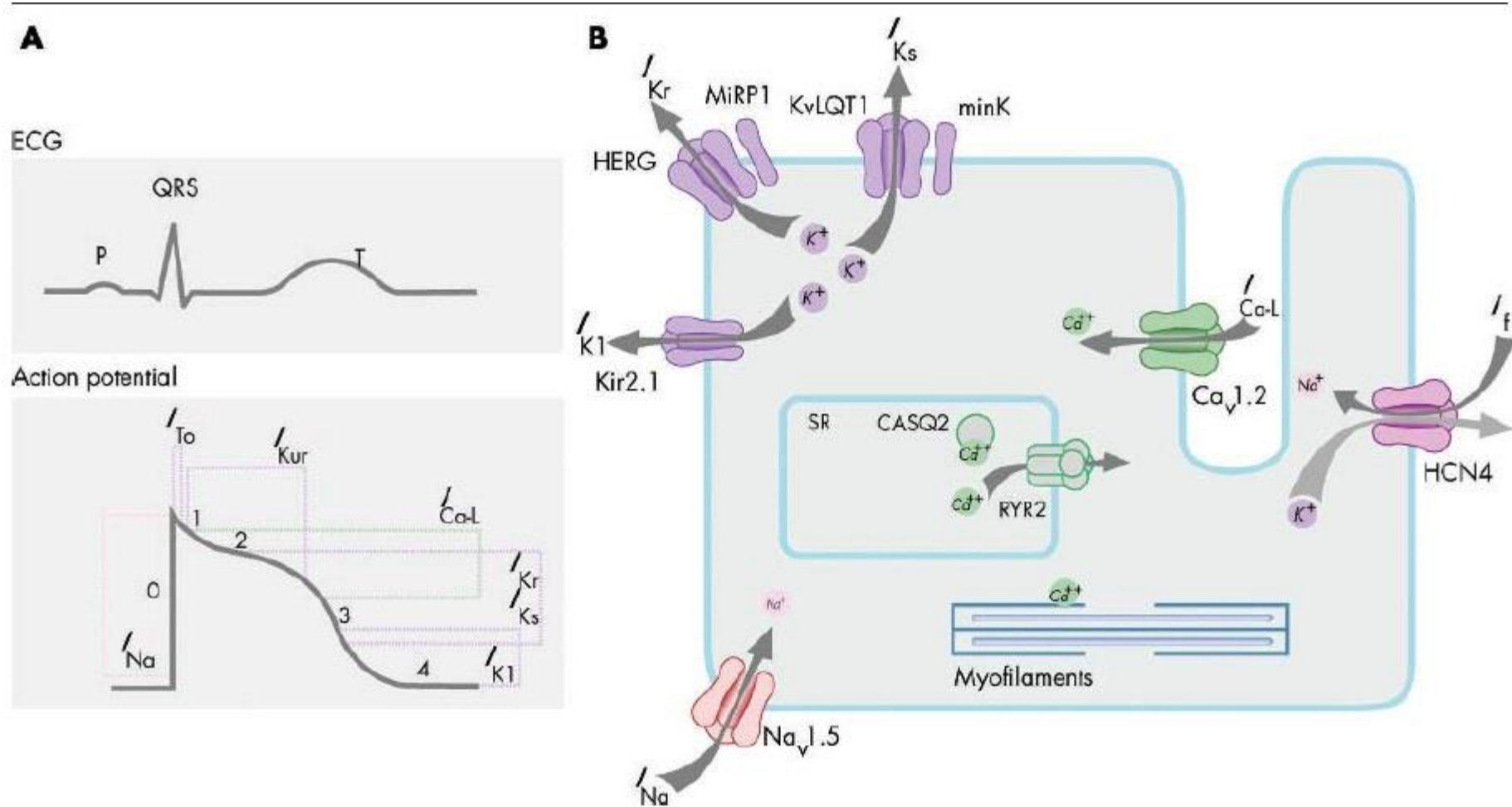
# Kv 11.1 (hERG1) CHANNELS IN CANCER





# hERG1

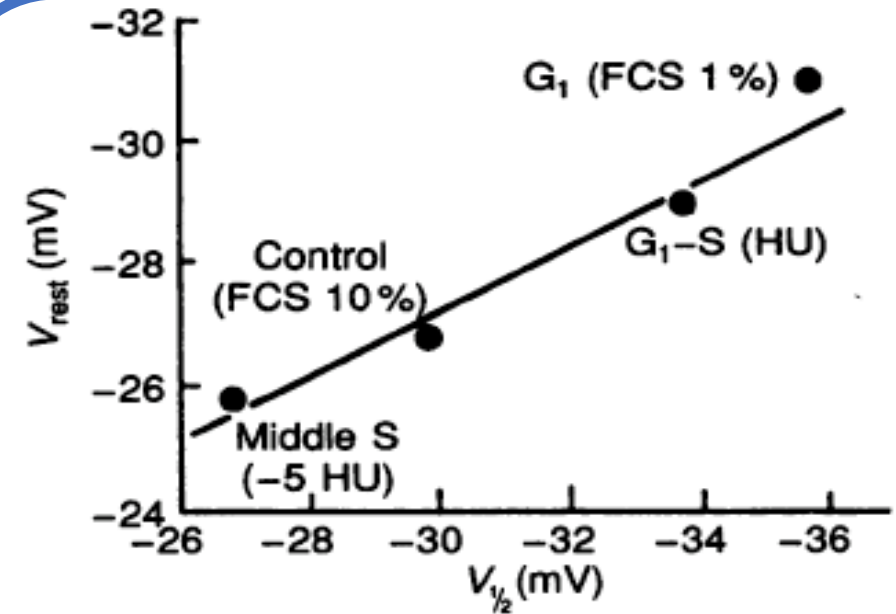
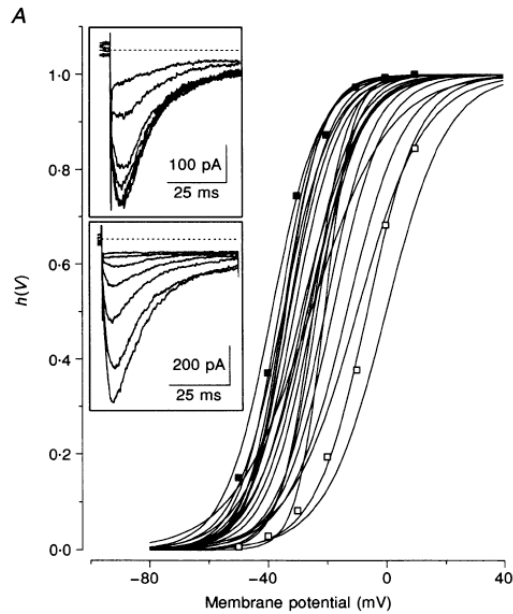




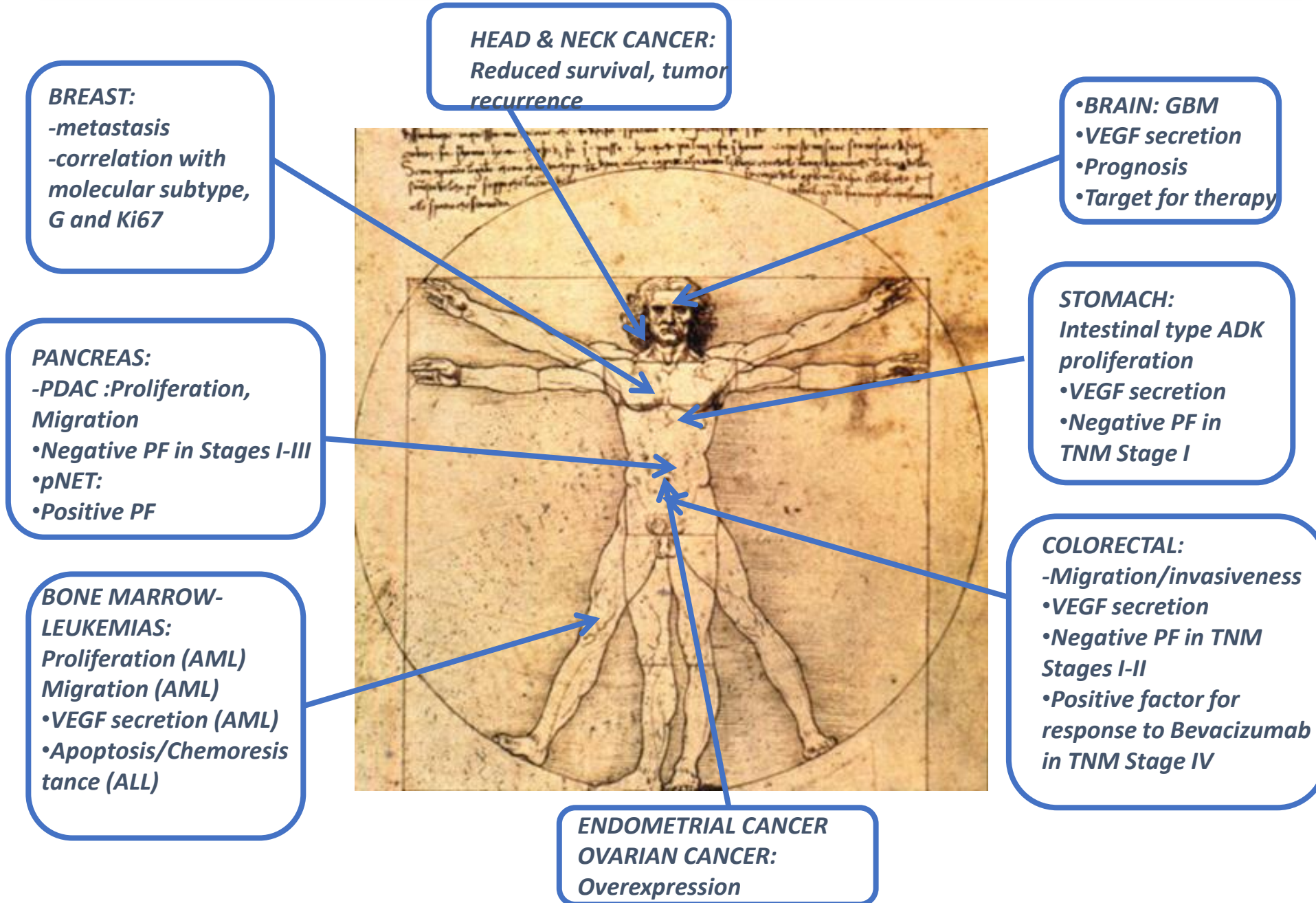
**Figure 1** Ionic currents contributing to the ventricular action potential (A) and schematic representation of a cardiomyocyte displaying (only) those proteins involved in the pathogenesis of inherited arrhythmia syndromes (B). In panel A, the action potential is aligned with its approximate time of action during the ECG. In panel B, ankyrin-B, an adapter protein involved in the long QT syndrome type 4, is not depicted.

## A novel inward-rectifying $K^+$ current with a cell-cycle dependence governs the resting potential of mammalian neuroblastoma cells

Annarosa Arcangeli\*, Laura Bianchi, Andrea Becchetti, Laura Faravelli, Marcella Coronello †, Enrico Mini †, Massimo Olivotto\* and Enzo Wanke ‡



# *hERG1 expression and role in human cancers*



# **hERG1 in Colorectal Cancer (CRC)**

## *herg1* Gene and HERG1 Protein Are Overexpressed in Colorectal Cancers and Regulate Cell Invasion of Tumor Cells

Elena Lastraioli,<sup>1</sup> Leonardo Guasti,<sup>1</sup> Olivia Crociani,<sup>1</sup> Simone Polvani,<sup>1</sup> Giovanna Hofmann,<sup>1</sup> Harry Witchel,<sup>4</sup> Lapo Bencini,<sup>3</sup> Massimo Calistri,<sup>3</sup> Luca Messerini,<sup>2</sup> Marco Scatizzi,<sup>3</sup> Renato Moretti,<sup>3</sup> Enzo Wanke,<sup>5</sup> Massimo Olivetto,<sup>1</sup> Gabriele Mugnai,<sup>1</sup> and Annarosa Arcangeli<sup>1</sup>

Departments of <sup>1</sup>Experimental Pathology and Oncology, and <sup>2</sup>Human Pathology and Oncology, University of Firenze, Firenze, Italy; <sup>3</sup>First Division of General Surgery and Transplantation, Careggi Hospital, Firenze, Italy; <sup>4</sup>Department of Physiology and Cardiovascular Research Laboratories, School of Medical Sciences, University of Bristol, Bristol, United Kingdom; and <sup>5</sup>Department of Biotechnology and Biosciences, University of Milano Bicocca, Milan, Italy

SCIENTIFIC  
REPORTS



OPEN

SUBJECT AREAS:

COLORECTAL CANCER

INTEGRIN SIGNALLING

TUMOUR ANGIOGENESIS

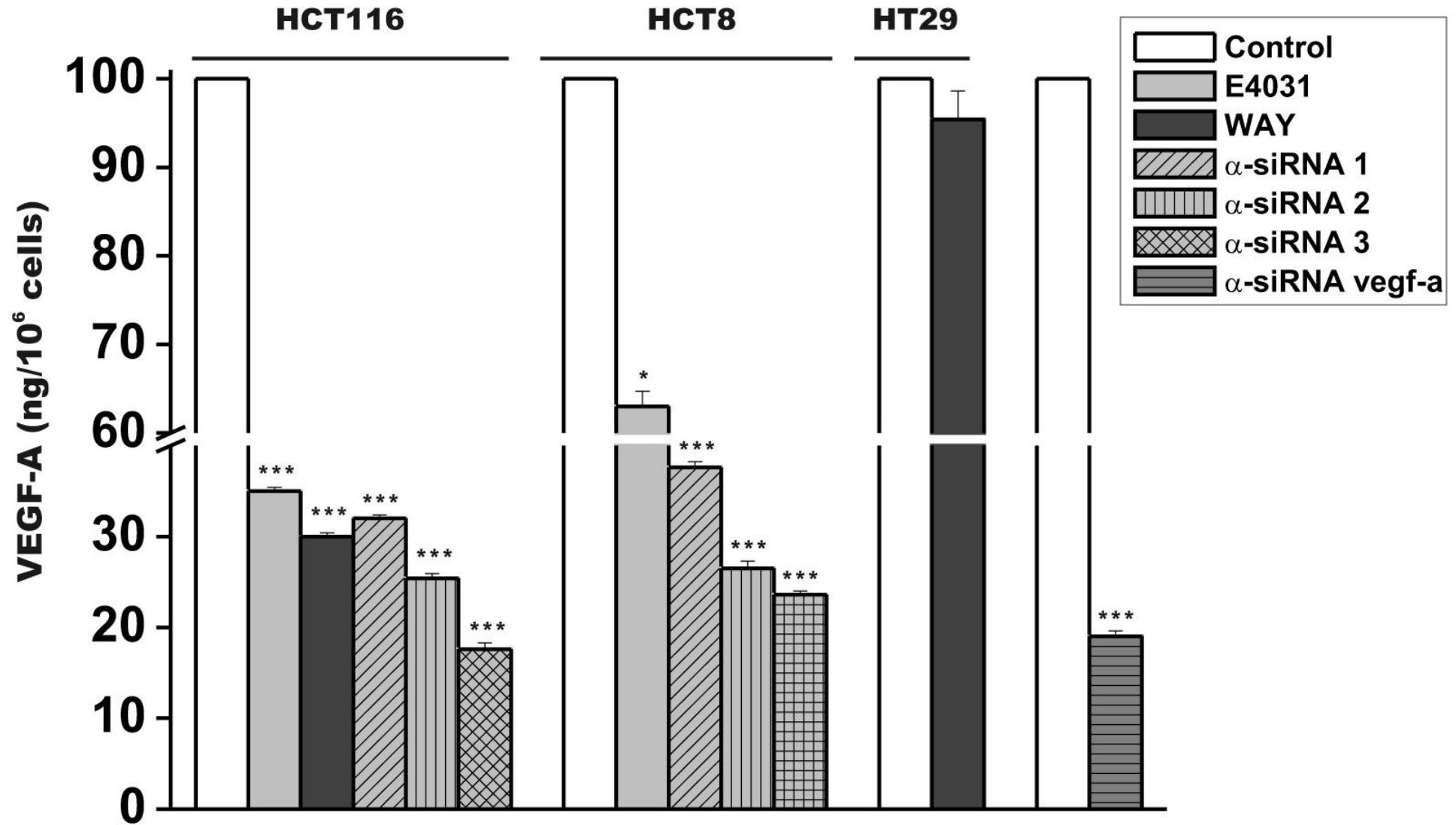
ION CHANNEL SIGNALLING

### hERG1 channels modulate integrin signaling to trigger angiogenesis and tumor progression in colorectal cancer

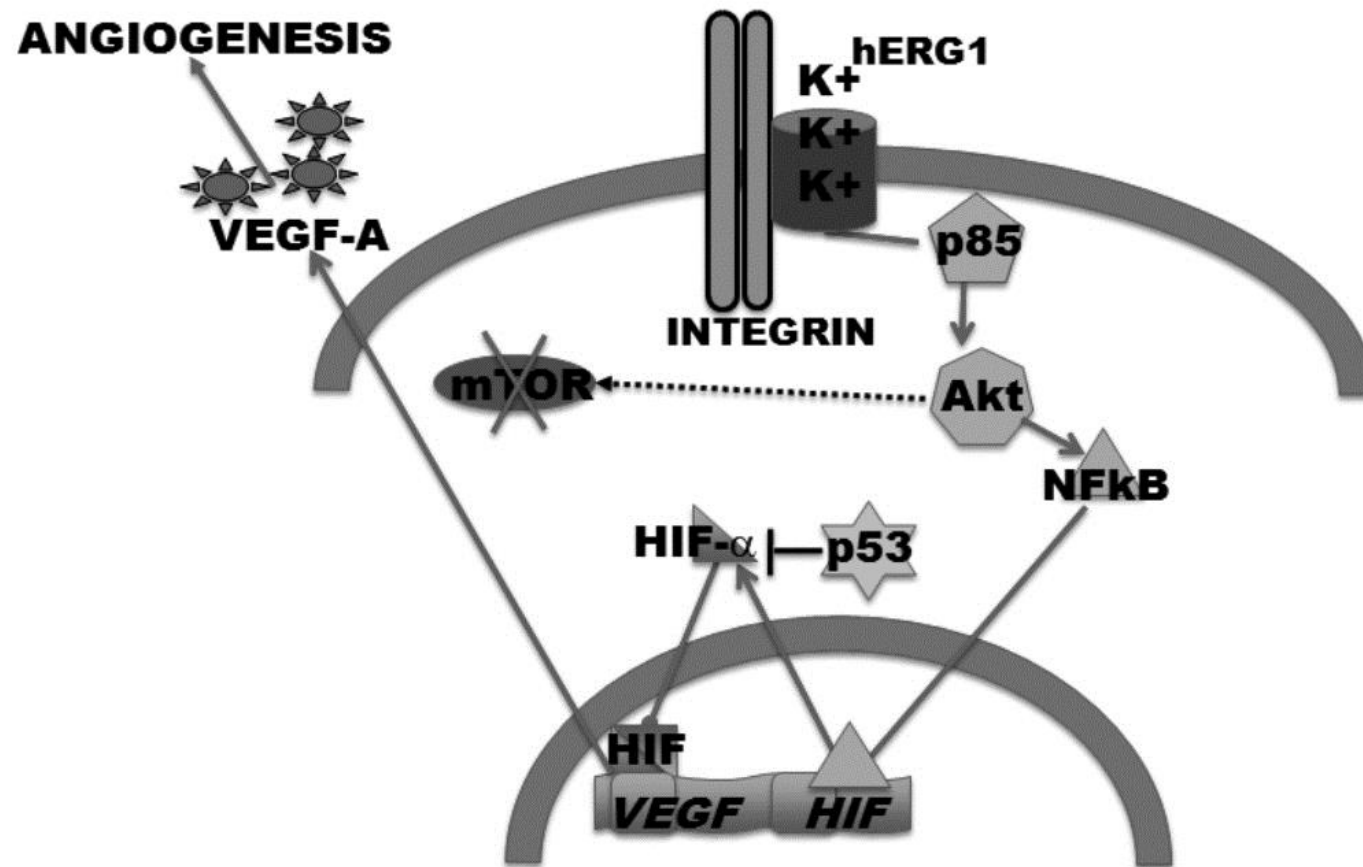
Olivia Crociani<sup>1</sup>, Francesca Zanieri<sup>1</sup>, Serena Pillozzi<sup>1</sup>, Elena Lastraioli<sup>1</sup>, Matteo Stefanini<sup>1</sup>, Antonella Fiore<sup>1</sup>, Angelo Fortunato<sup>1</sup>, Massimo D'Amico<sup>1</sup>, Marika Masselli<sup>1</sup>, Emanuele De Lorenzo<sup>1</sup>, Luca Gasparoli<sup>1</sup>, Martina Chiu<sup>2</sup>, Ovidio Bussolati<sup>2</sup>, Andrea Becchetti<sup>2</sup> & Annarosa Arcangeli<sup>1</sup>



# hERG1 regulates VEGF-A expression and secretion

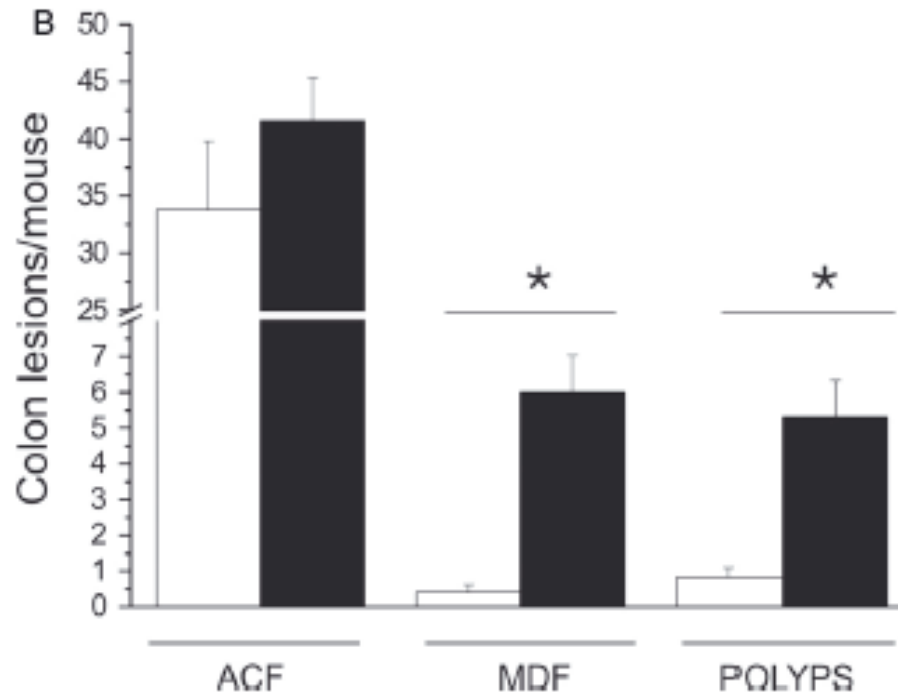
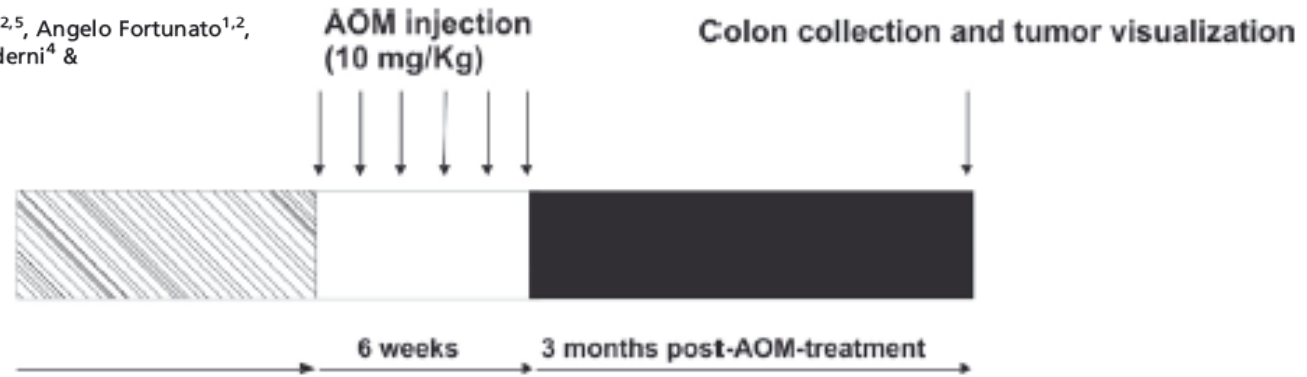


# The hERG1-centered pro-angiogenic signaling pathway in CRC



**Characterization of hERG1 channel role in mouse colorectal carcinogenesis**

Antonella Fiore<sup>1,2,a</sup>, Laura Carraresi<sup>3,a</sup>, Angela Morabito<sup>1,2</sup>, Simone Polvani<sup>1,2,5</sup>, Angelo Fortunato<sup>1,2</sup>, Elena Lastraioli<sup>1,2</sup>, Angelo P. Femia<sup>4</sup>, Emanuele De Lorenzo<sup>1,6</sup>, Giovanna Caderni<sup>4</sup> & Annarosa Arcangeli<sup>1,2</sup>



White bars= WT mice

Black bars = hERG1 TG (over-expressing) mice

# hERG1 positivity and Glut-1 negativity identifies high-risk TNM stage I/II Colorectal Cancer patients

Translational Oncology

www.transonc.com

Volume 5 Number 2 April 2012 pp. 105-112 105

## hERG1 Channels and Glut-1 as Independent Prognostic Indicators of Worse Outcome in Stage I and II Colorectal Cancer: A Pilot Study<sup>1</sup>

Elena Lestrali<sup>1\*</sup>, Lupo Bendini<sup>1</sup>, Elisa Bianchini<sup>1</sup>, Maria Raffaella Romoli<sup>2</sup>, Olivia Crociani<sup>2</sup>, Elisa Giommoni<sup>2</sup>, Luca Messerini<sup>2</sup>, Silvia Gasperoni<sup>2</sup>, Renato Morotti<sup>2</sup>, Francesco Di Costanzo<sup>3</sup>, Luca Boni<sup>1,2</sup> and Annarosa Arzangeli<sup>1,2</sup>

<sup>1</sup>Department of Experimental Pathology and Oncology, University of Florence, Istituto Toscano Tumori, Florence, Italy; <sup>2</sup>General Surgery and Surgical Oncology, Azienda Ospedaliero-Universitaria, Careggi, Florence, Italy;

<sup>3</sup>Clinical Trials Coordinating Center, Azienda Ospedaliero-Universitaria, Careggi, Florence, Italy

<sup>\*</sup>Correspondence: Elena Lestrali, Department of Experimental Pathology and Oncology, University of Florence, Via Sesto 49, 50132 Florence, Italy. E-mail: lestrali@unifi.it

OncoTargets and Therapy

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ORIGINAL RESEARCH

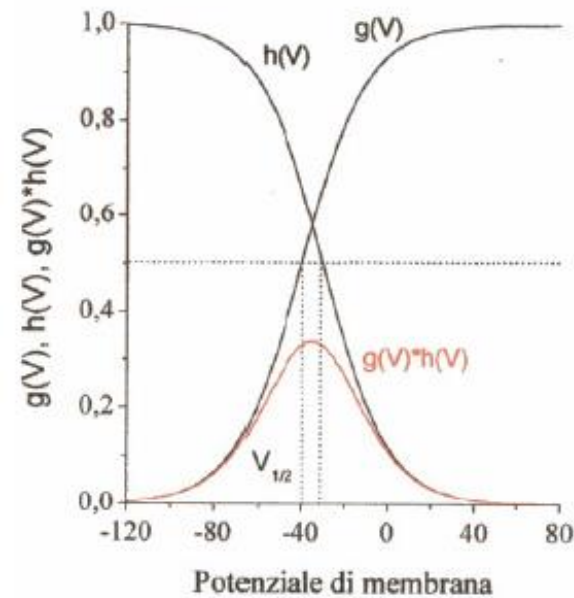
## hERG1 positivity and Glut-1 negativity identifies high-risk TNM stage I and II colorectal cancer patients, regardless of adjuvant chemotherapy

# **The dual mode of action of hERG1: ion flux vs non-conductive mechanisms**

# *hERG1 role in human cancers*

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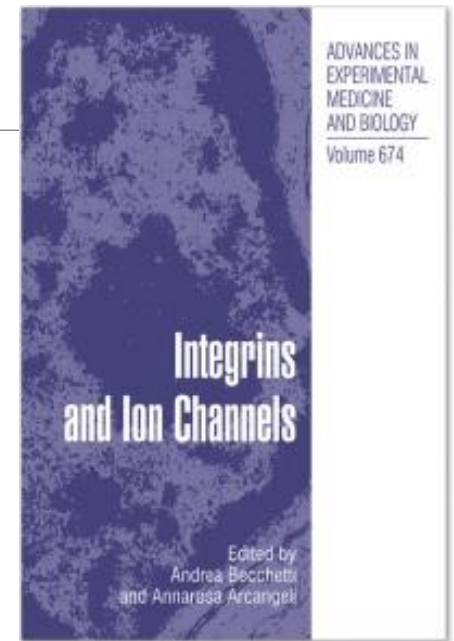
- ✓ How can hERG1 exert different roles?



Membrane potential

- ✓ Regulation of  $V_{rest}$
- ✓ Regulation of intracellular signaling

# *hERG1 and integrins*



Review

TRENDS in Cell Biology Vol.16 No.12

Full text provided by [www.sciencedirect.com](http://www.sciencedirect.com)



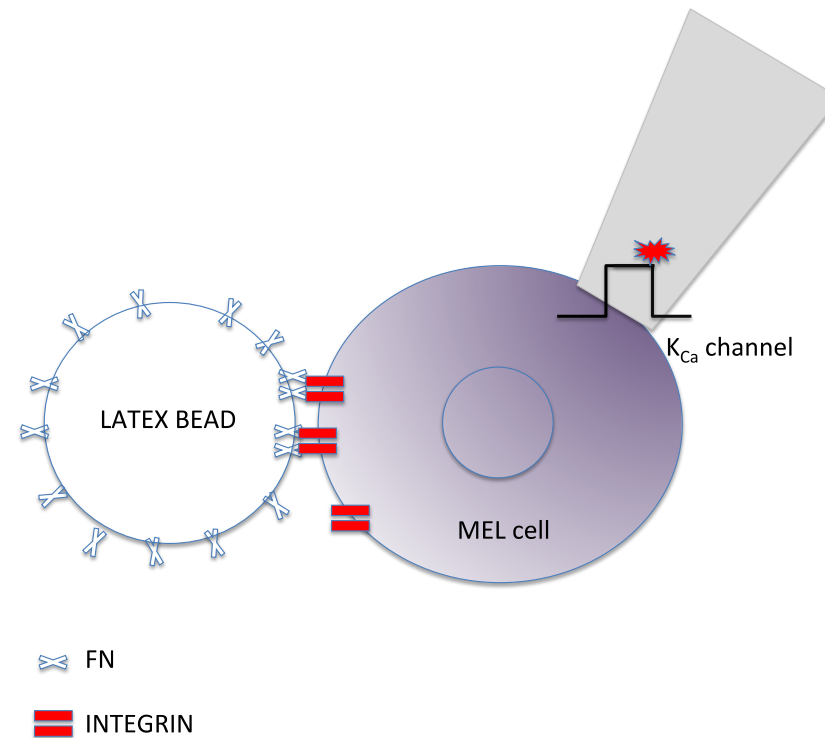
## Complex functional interaction between integrin receptors and ion channels

Annarosa Arcangeli<sup>1</sup> and Andrea Becchetti<sup>2</sup>

<sup>1</sup> Department of Experimental Pathology and Oncology, University of Firenze, Viale G.B. Morgagni 50, 50134 Firenze, Italy

<sup>2</sup> Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy

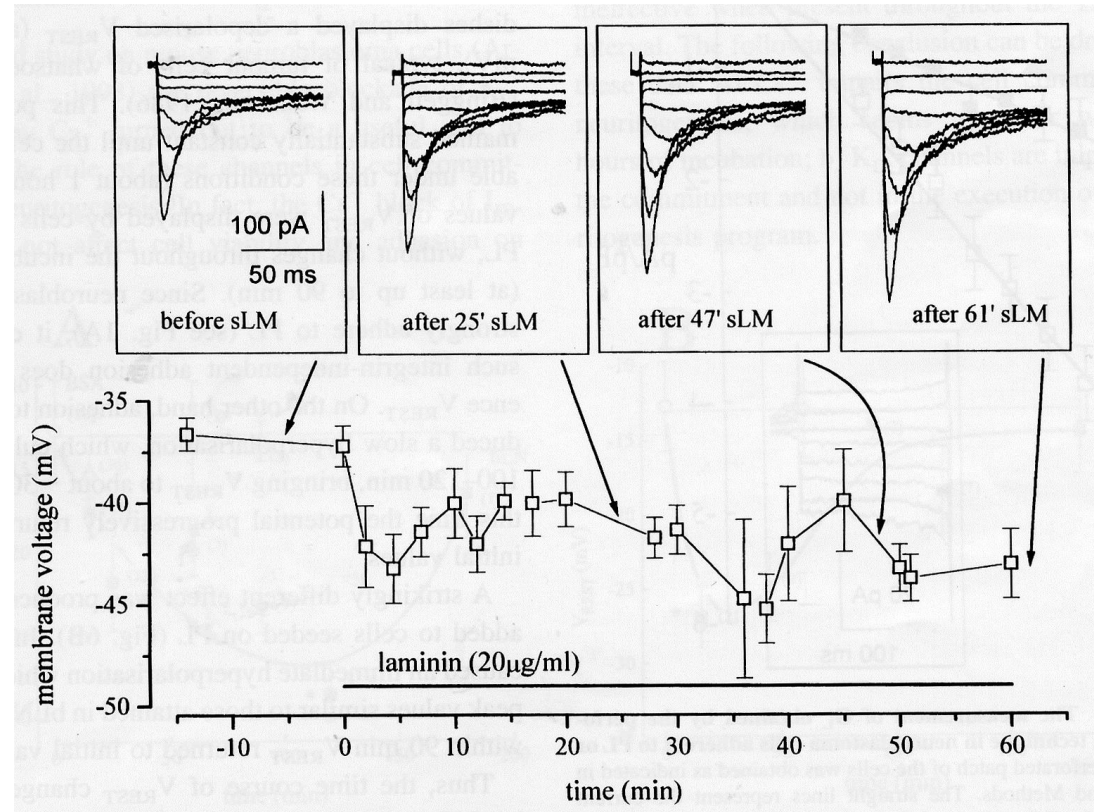
# Engagement of the $\beta_1$ integrin activates $\text{Ca}^{2+}$ -dependent $\text{K}^+$ channels in murine erythroleukemia (MEL) cells



Arcangeli, A., et al., *Biochem. Biophys. Res. Commun.*, 146, 1450-1457, 1987.  
Arcangeli A. et al., *Biochem. Biophys. Res. Commun.*, 177, 1266-1272, 1991.  
Becchetti A. et al., *Proc. Royal Soc. Ser. B*, 248, 235-240, 1992



# Engagement of the $\beta_1$ integrin activates hERG1 K<sup>+</sup> channels in neuroblastoma cells.

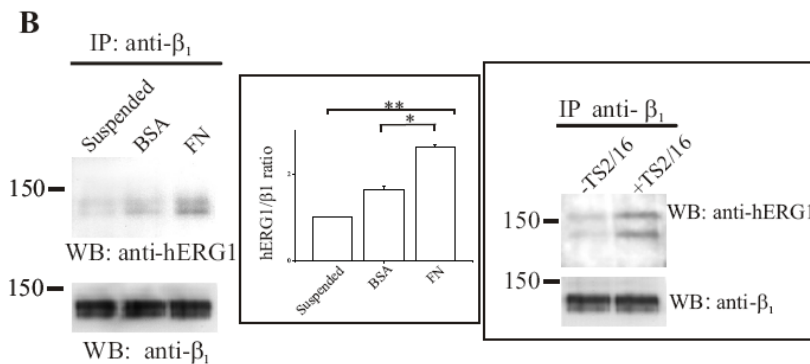
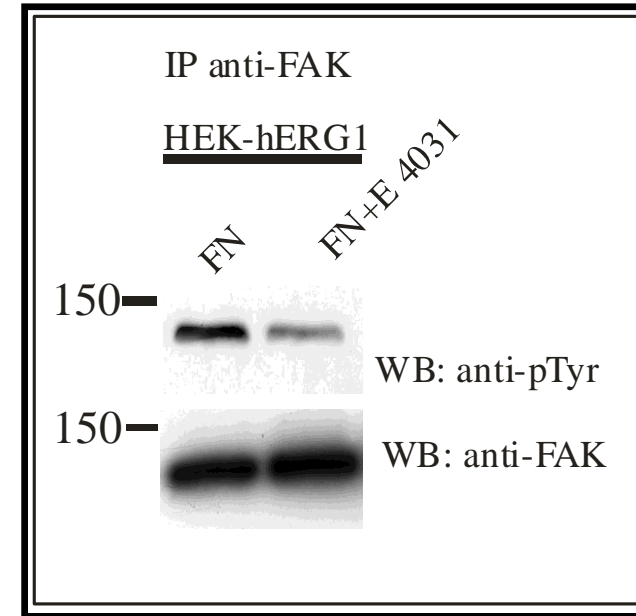
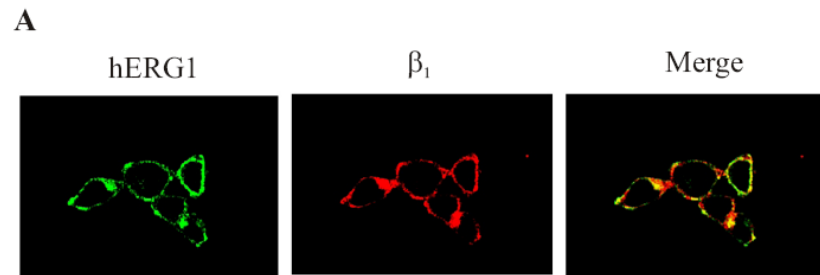
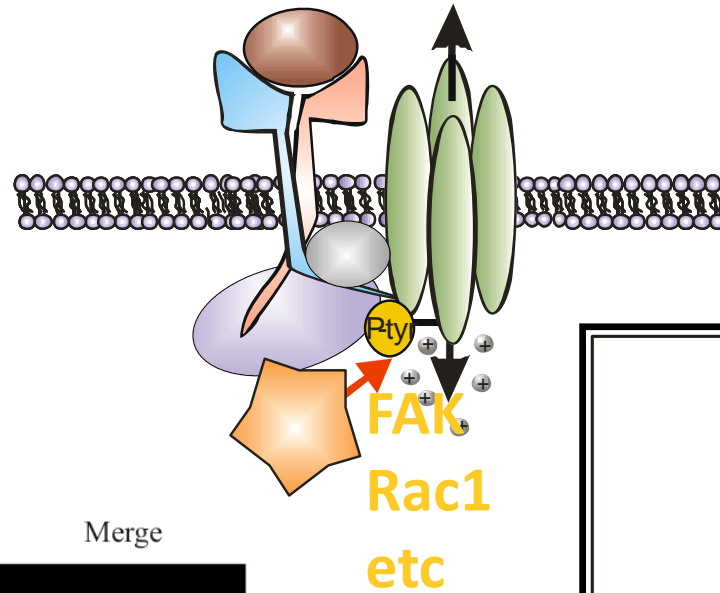


Arcangeli A. et al., *J. Cell. Biol.*, 122: 1131-1143, 1993

Arcangeli, A. et al., *J. Physiol.*, 489, 455-471, 1995

Arcangeli A., et al., *Cell Adhesion Commun.*, 4: 369-384, 1996

# Integrin-channel complex: hERG1/ $\beta_1$



**Cherubini A. et al., Mol.Biol.Cell, 2005**

# The hERG1/ $\beta$ 1 integrin complex in CRC

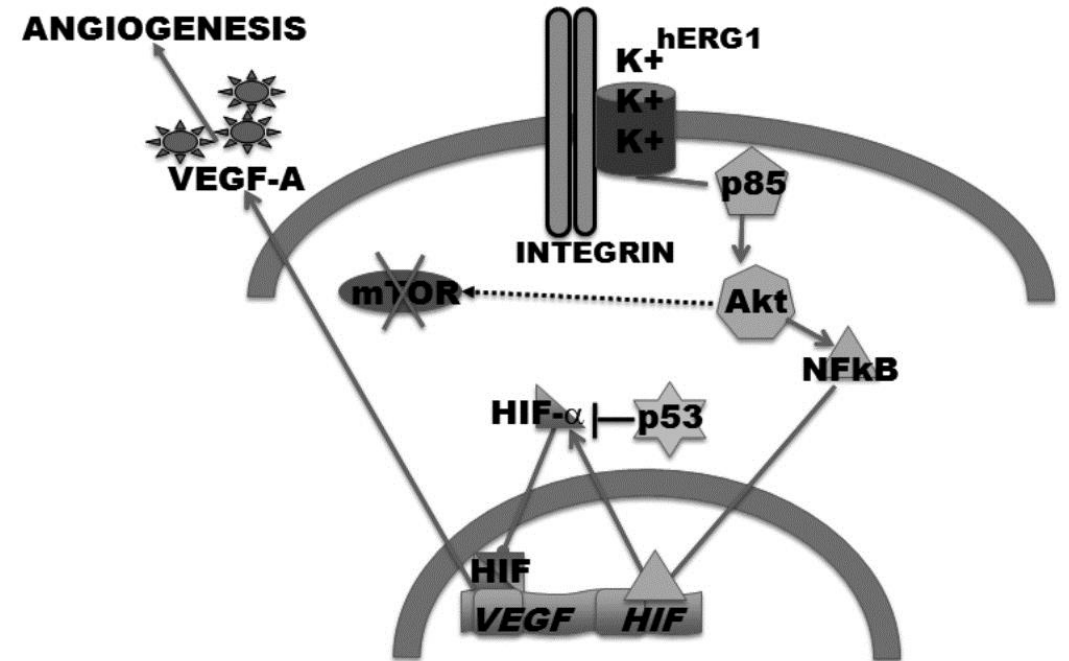


OPEN

hERG1 channels modulate integrin signaling to trigger angiogenesis and tumor progression in colorectal cancer

SUBJECT AREAS:  
COLORECTAL CANCER  
INTEGRIN SIGNALLING  
TUMOUR ANGIOGENESIS  
ION CHANNEL SIGNALLING

Olivia Crociani<sup>1</sup>, Francesca Zanieri<sup>1</sup>, Serena Pillozzi<sup>1</sup>, Elena Lastraioi<sup>1</sup>, Matteo Stefanini<sup>1</sup>, Antonella Fiore<sup>1</sup>, Angelo Fortunato<sup>1</sup>, Massimo D'Amico<sup>1</sup>, Marika Masselli<sup>1</sup>, Emanuele De Lorenzo<sup>1</sup>, Luca Gasparoli<sup>1</sup>, Martina Chiu<sup>2</sup>, Ovidio Bussolati<sup>2</sup>, Andrea Becchetti<sup>2</sup> & Annarosa Arcangeli<sup>1</sup>



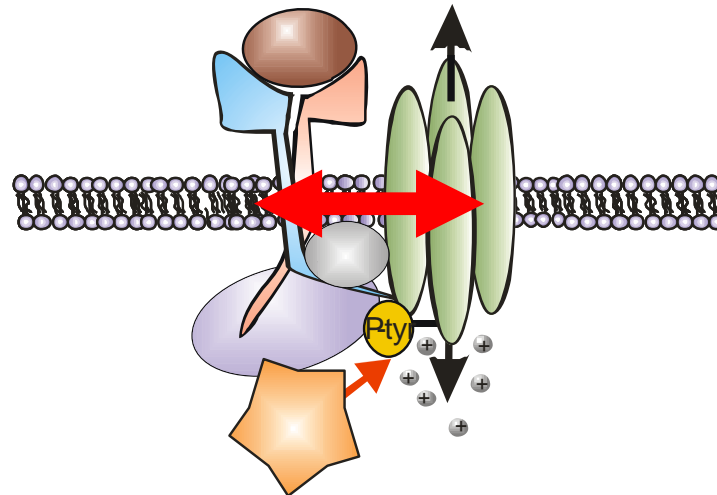
## CANCER

# The conformational state of hERG1 channels determines integrin association, downstream signaling, and cancer progression

Andrea Becchetti,<sup>1</sup> Silvia Crescioli,<sup>2</sup> Francesca Zanieri,<sup>2</sup> Giulia Petroni,<sup>2</sup> Raffaella Mercatelli,<sup>3</sup> Stefano Coppola,<sup>4</sup> Luca Gasparoli,<sup>2</sup> Massimo D'Amico,<sup>5</sup> Serena Pillozzi,<sup>2</sup> Olivia Crociani,<sup>2</sup> Matteo Stefanini,<sup>5</sup> Antonella Fiore,<sup>2</sup> Laura Carraresi,<sup>5</sup> Virginia Morello,<sup>6\*</sup> Sagar Manoli,<sup>2</sup> Maria Felice Brizzi,<sup>7</sup> Davide Ricci,<sup>8</sup> Mauro Rinaldi,<sup>8</sup> Alessio Masi,<sup>2†</sup> Thomas Schmidt,<sup>4</sup> Franco Quercioli,<sup>3</sup> Paola Defilippi,<sup>4</sup> Annarosa Arcangeli<sup>2‡</sup>

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The association between the channel and the integrin occurs through the TM domains of either proteins



# The *hERG1* conformational state determines (the closed state favours) integrin association

***hERG1* mutants:**

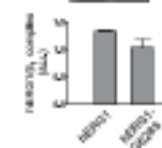
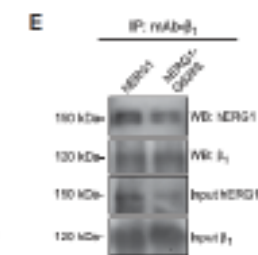
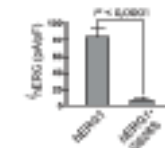
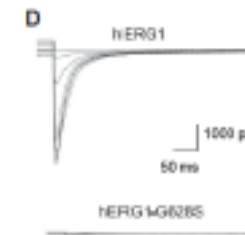
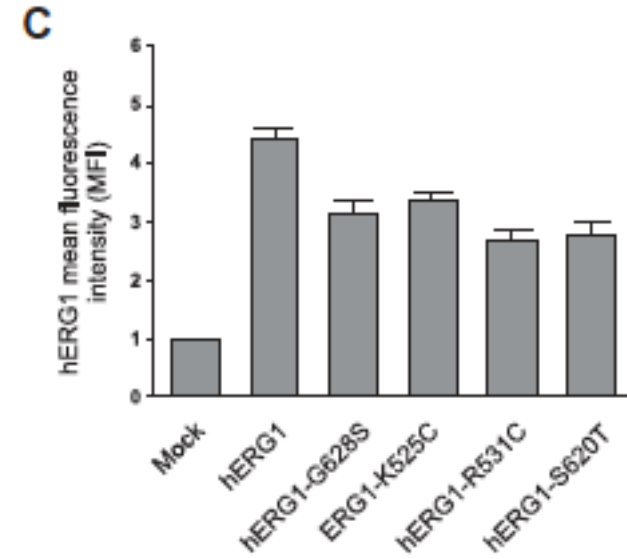
***G628S*: non conductive**

***S620T*: non inactivating**

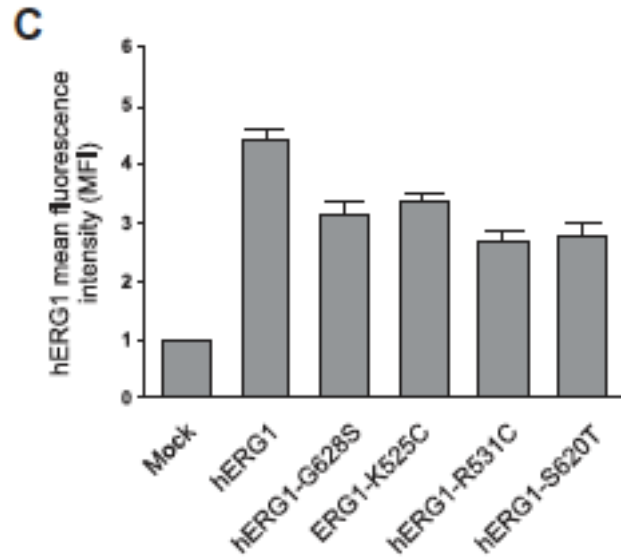
***K525C*: S4 (voltage sensor) mutant\*  
preferentially in the open state**

***R531C*: S4 (voltage sensor) mutant\*  
preferentially in the closed state**

**\*=alterations of gating**



# The *hERG1* conformational state determines (the closed state favours) integrin association



*hERG1* mutants:

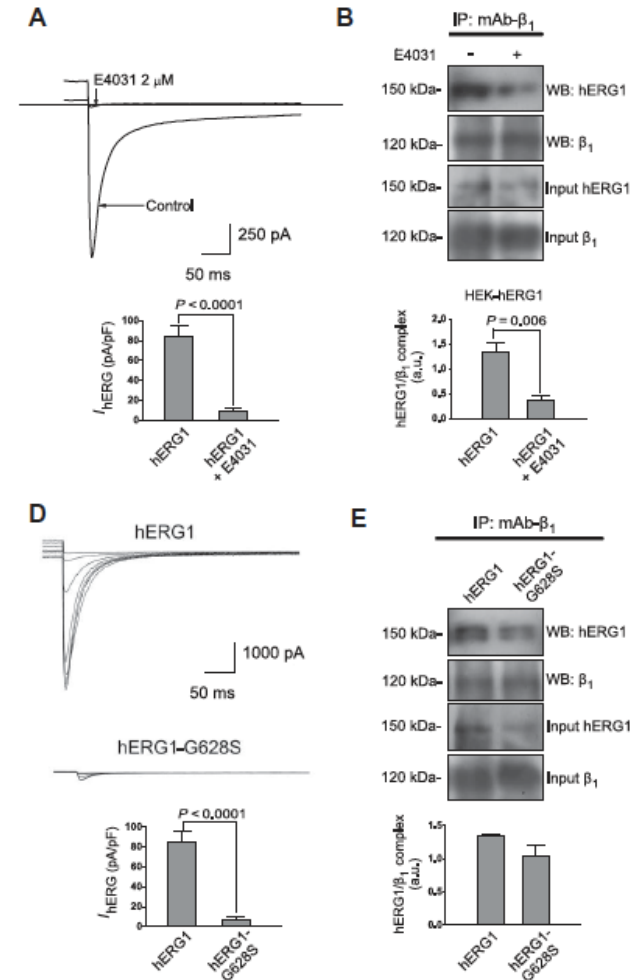
G628S: non conductive

S620T: non inactivating

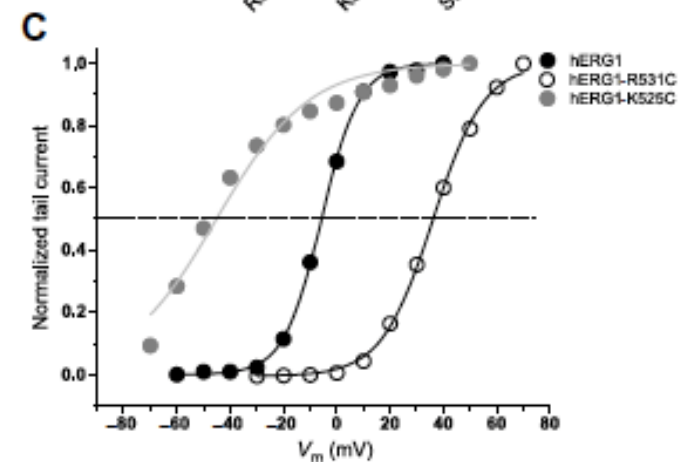
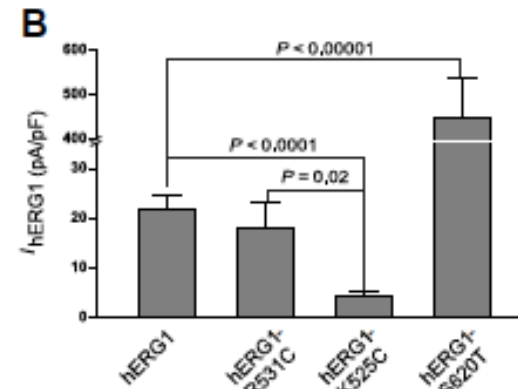
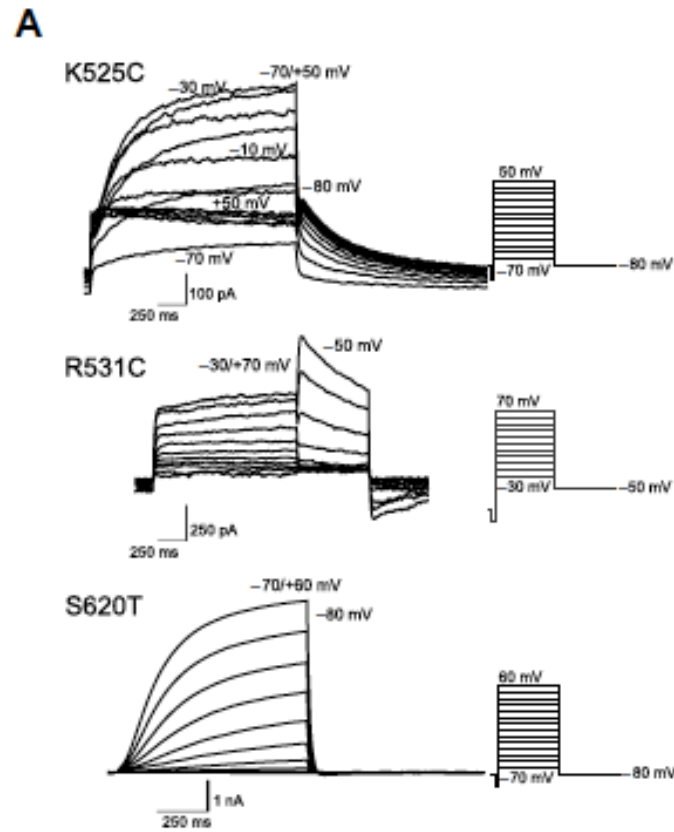
K525C: S4 (voltage sensor) mutant\*

R531C: S4 (voltage sensor) mutant\*

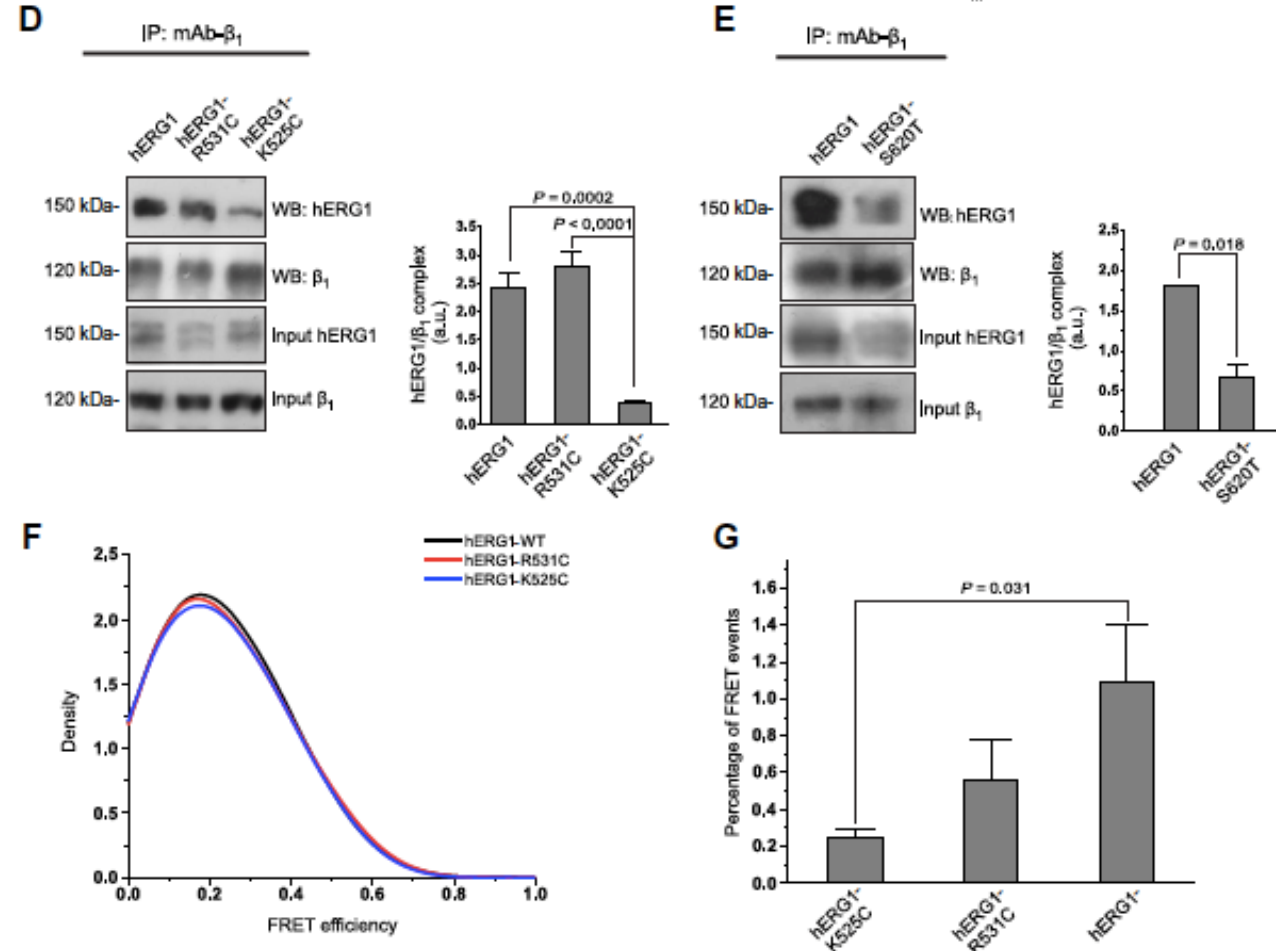
\*=alterations of gating



# The hERG1 conformational state determines (the closed state favours) integrin association

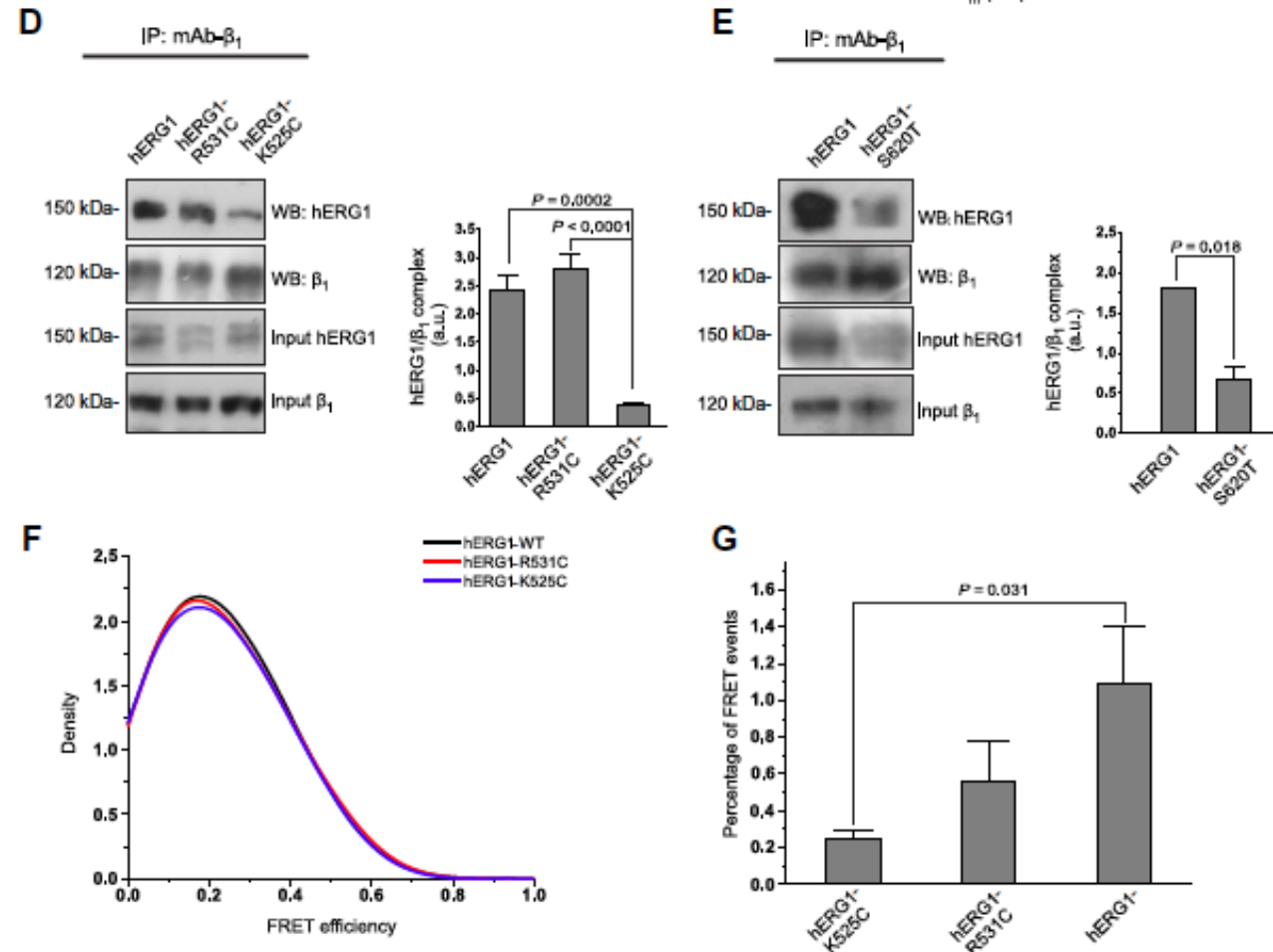


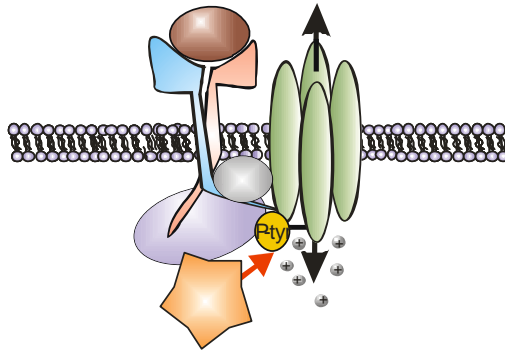
# The hERG1 conformational state determines (the closed state favours) integrin association





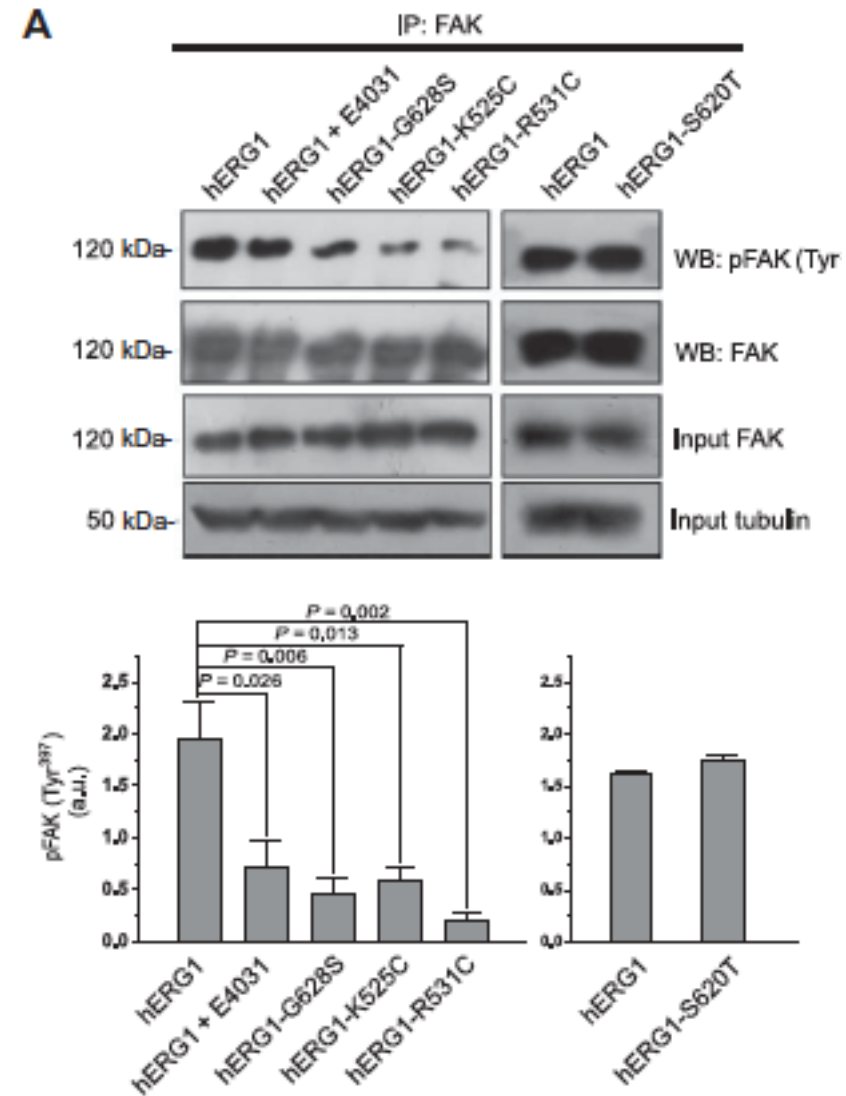
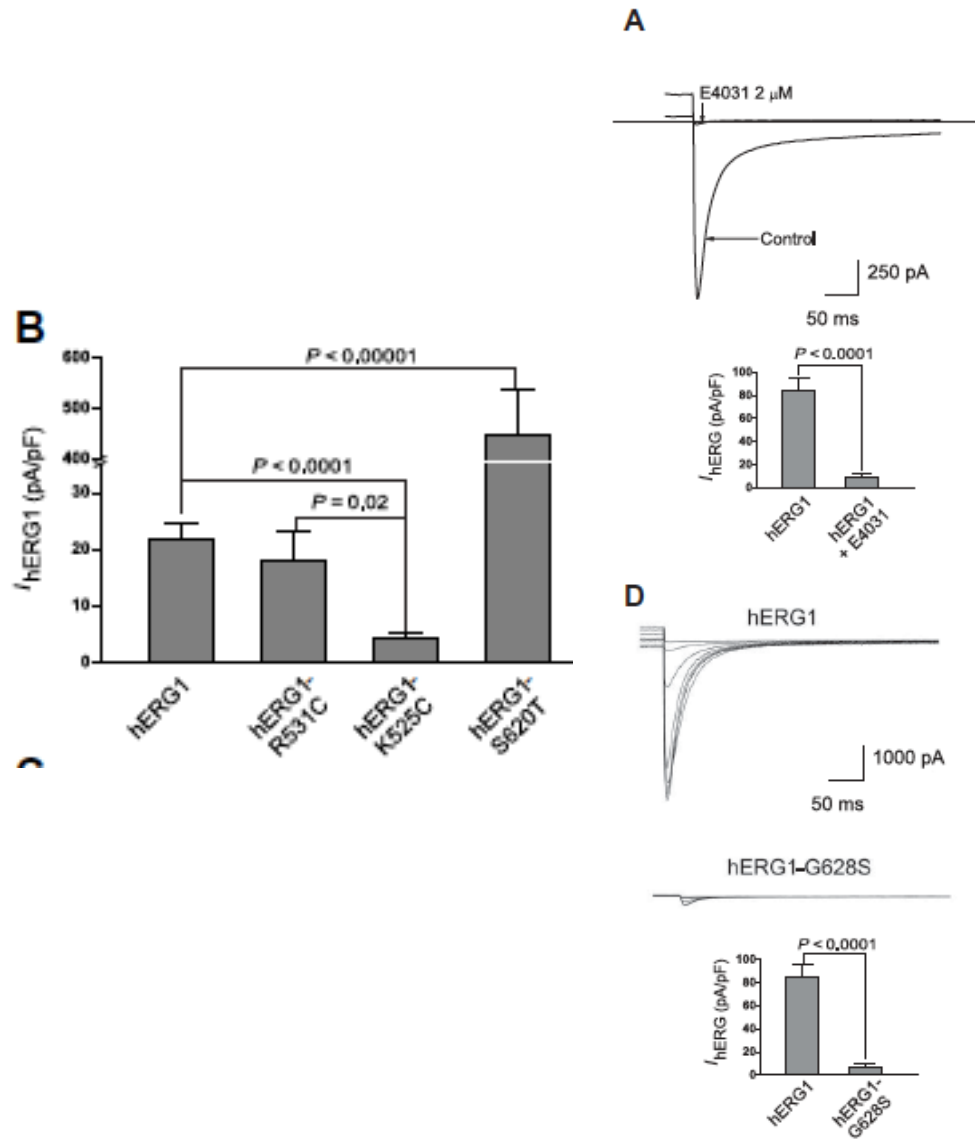
# The *hERG1* conformational state determines (the closed state favours) integrin association



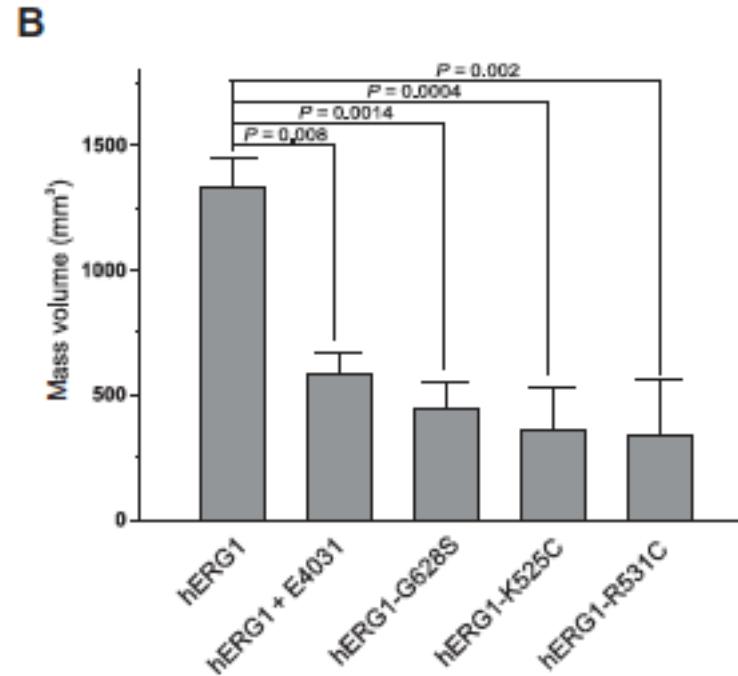


| <b>hERG1 mutant</b> | <b>hERG1 conformational state</b>              | <b>hERG1 currents</b> | <b>hERG1/b1 integrin complex</b> |
|---------------------|--|-----------------------|----------------------------------|
| <b>WT</b>           | <b>Open/Closed</b>                             | <b>+++</b>            | <b>+++</b>                       |
| <b>G628S</b>        | <b>Non Conductive</b>                          | <b>0</b>              | <b>++/+++</b>                    |
| <b>R531C</b>        | <b>Closed</b>                                  | <b>+</b>              | <b>+++ /++++</b>                 |
| <b>K525C</b>        | <b>Open</b>                                    | <b>+</b>              | <b>+</b>                         |
| <b>S620T</b>        | <b>No inactivation</b>                         | <b>++++</b>           | <b>+/++</b>                      |
| <b>E4031</b>        | <b>Blocked<br/>(binding in the open state)</b> | <b>0</b>              | <b>+/++</b>                      |

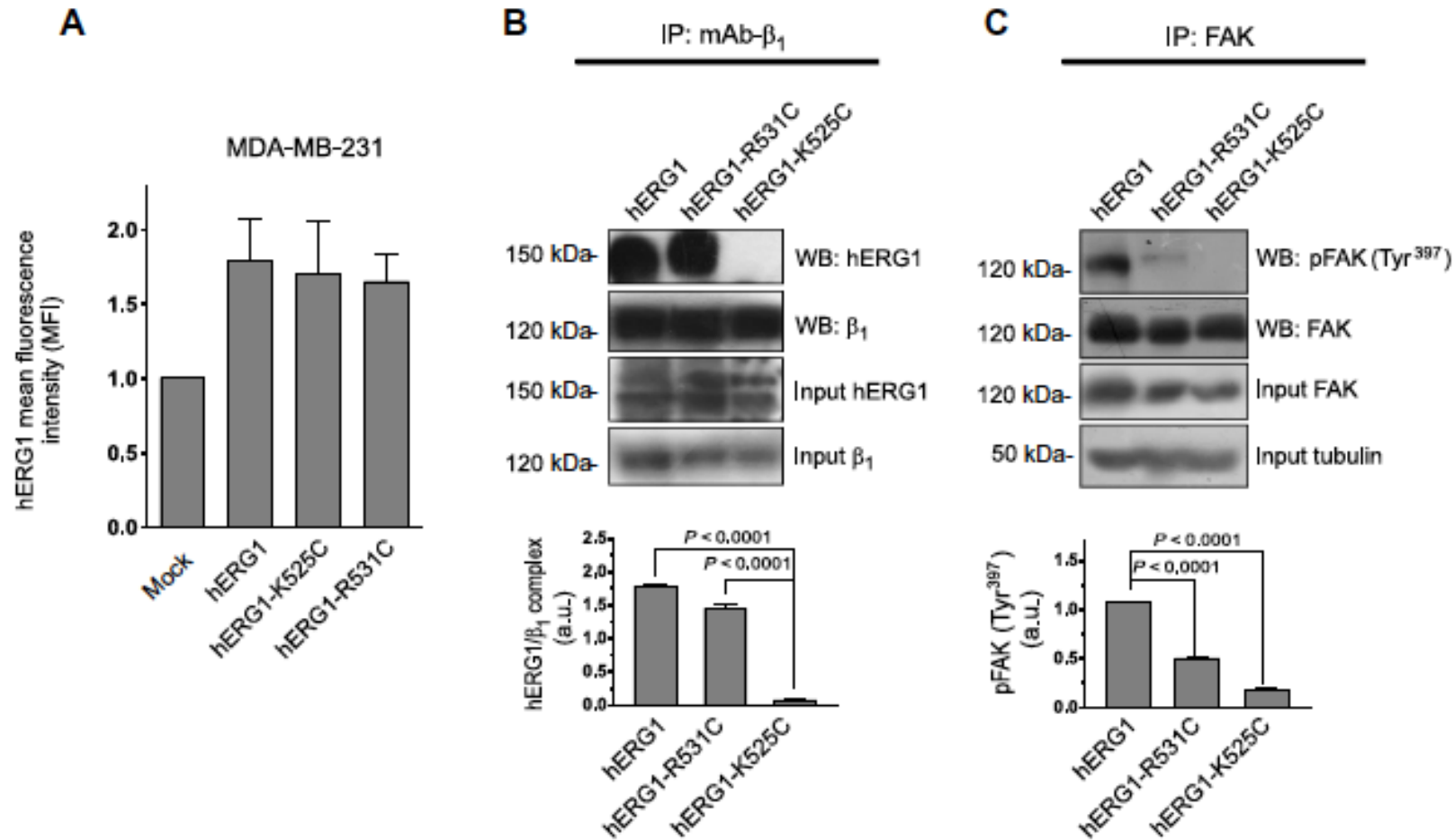
# hERG1 currents (ion flux) regulate FAK phosphorylation



# hERG1 currents (ion flux) regulate FAK phosphorylation.....and local tumor growth



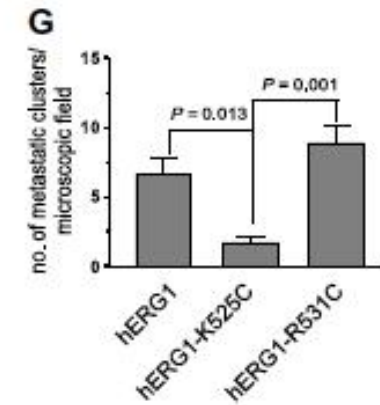
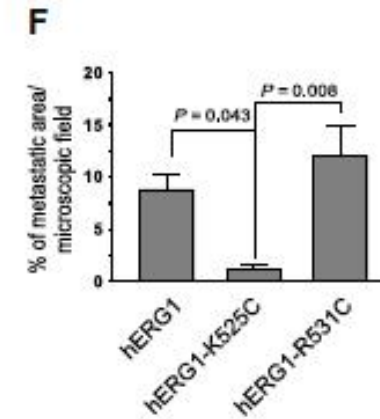
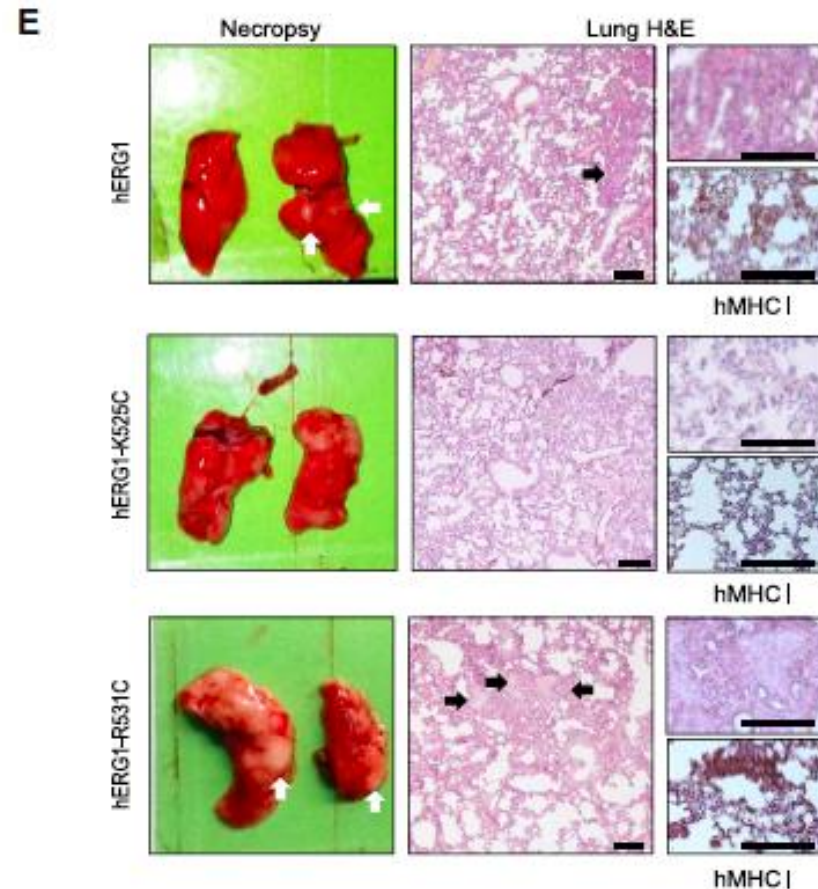
# The hERG1 conformational state determines (the closed state favours) integrin association: MDA-MB-231 breast cancer cells



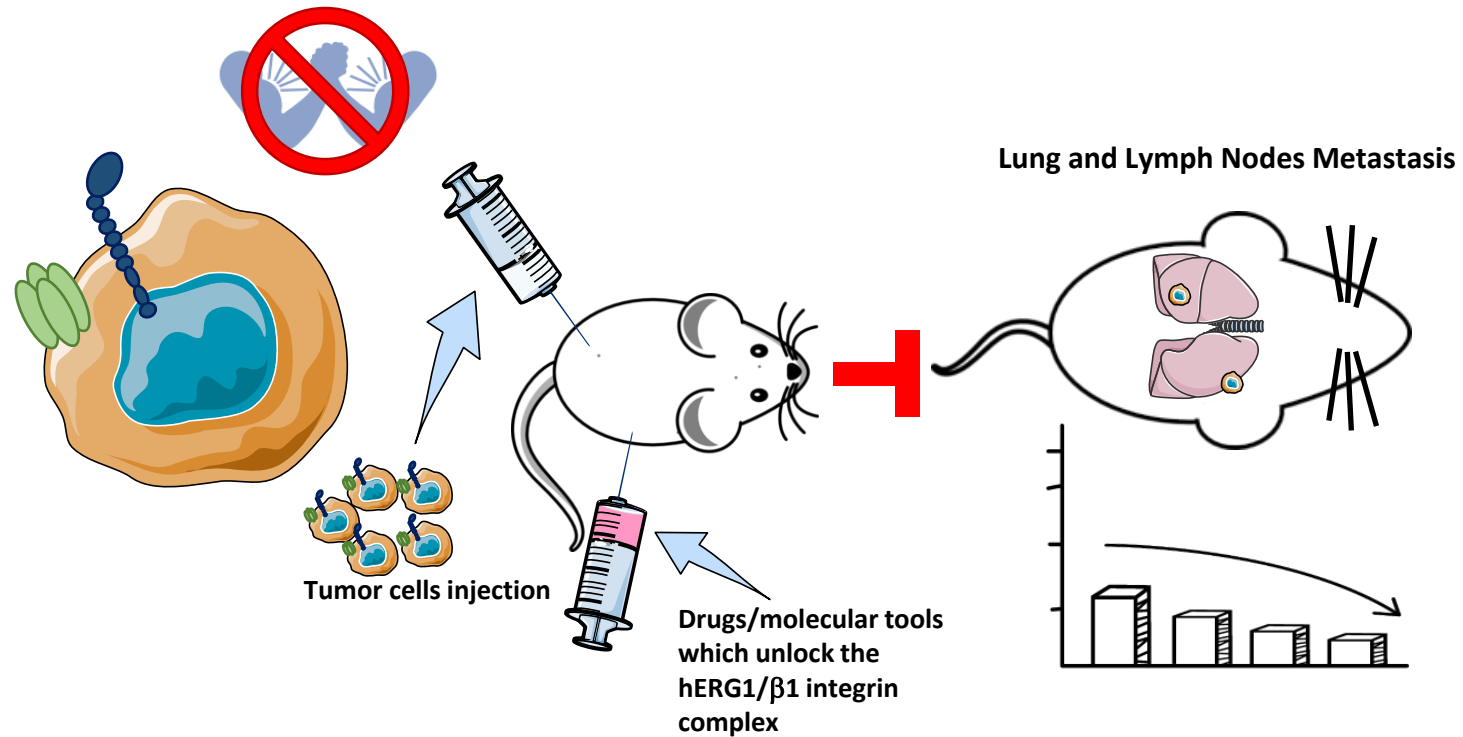
**The hERG1 conformational state determines (the closed state favours) integrin association.....and tumor metastasis (MDA-MB-231 breast cancer cells)**

|  | MDA-MB-231 | hERG1        | hERG1-K525C  | hERG1-R531C  |
|--|------------|--------------|--------------|--------------|
| <b>Local tumor growth</b>                      |            |              |              |              |
| Number of tumor masses (%)                     |            | 9/10 (90%)   | 10/10 (100%) | 9/10 (90%)   |
| Median tumor volume (mm <sup>3</sup> )         |            | 150 (19–300) | 122 (33–300) | 212 (33–300) |
| <b>Metastases</b>                              |            |              |              |              |
| <b>Inguinal lymph nodes</b>                    |            |              |              |              |
| Number of mice with macroscopic metastases (%) |            | 2/5 (40%)    | 0/5 (0%)     | 3/5 (60%)    |
| <b>Lung</b>                                    |            |              |              |              |
| Number of mice with macroscopic metastases (%) |            | 2/5 (40%)    | 0/5 (0%)     | 4/5 (80%)    |

# The hERG1 conformational state determines (the closed state favours) integrin association.....and tumor metastasis (MDA-MB-231 breast cancer cells)



# Disrupting the hERG1/ $\beta$ 1 integrin complex inhibits tumor metastasis

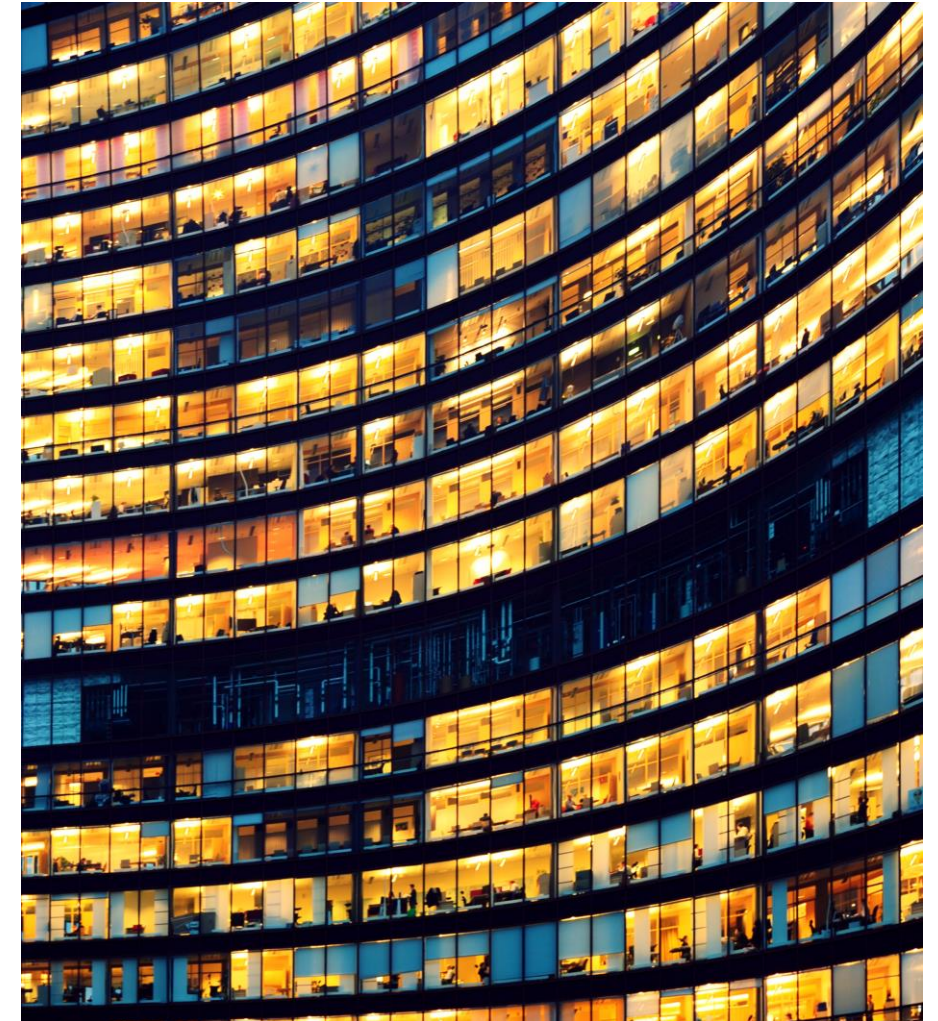
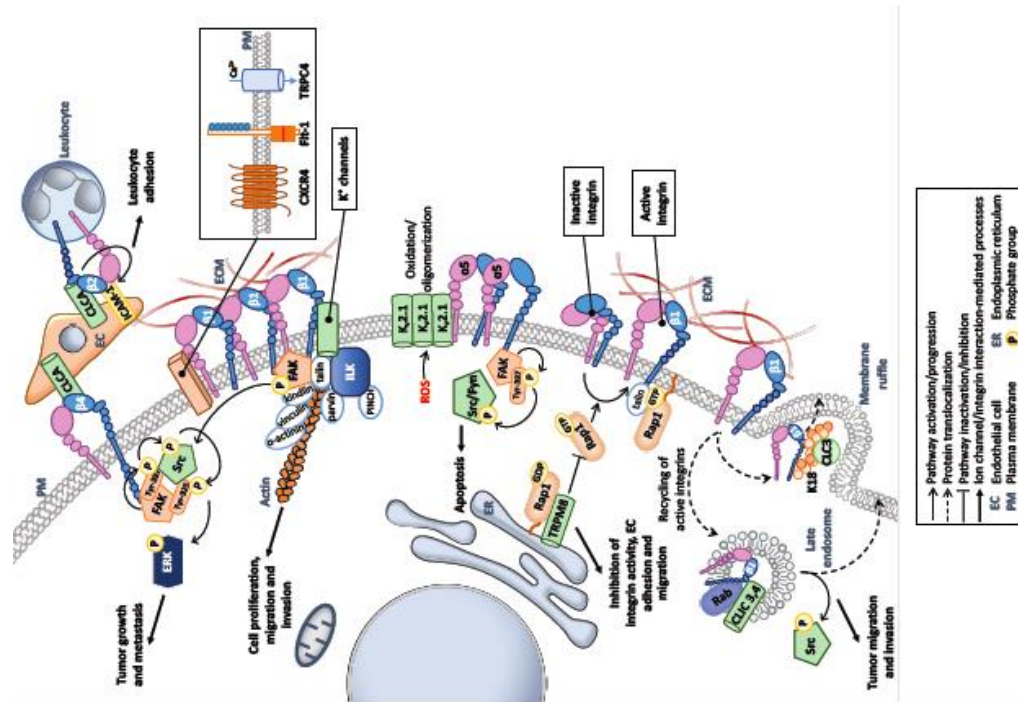


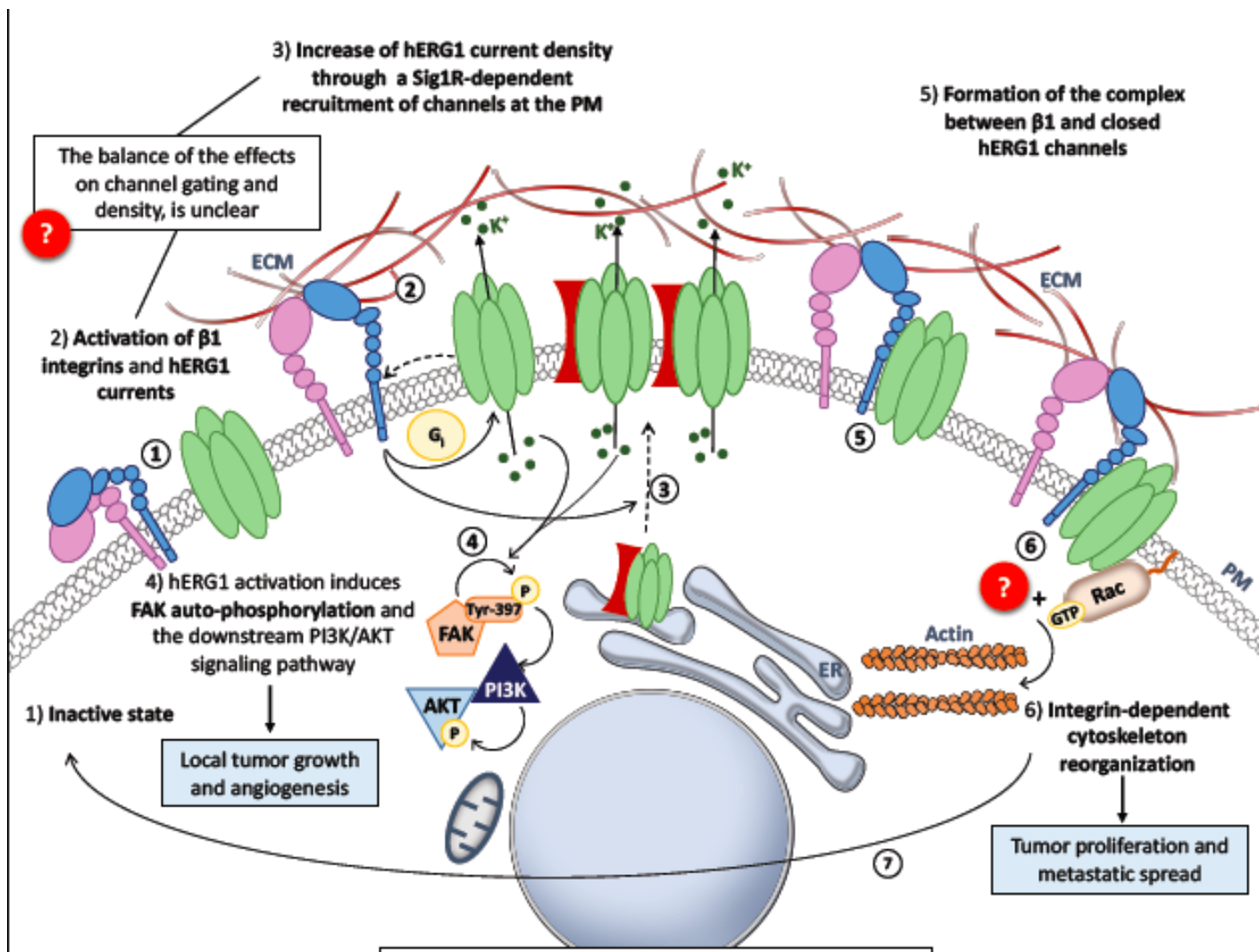


Opinion

# Ion Channel Conformations Regulate Integrin-Dependent Signaling

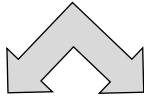
Andrea Becchetti,<sup>1,\*</sup> Giulia Petroni,<sup>2</sup> and Annarosa Arcangeli<sup>2</sup>





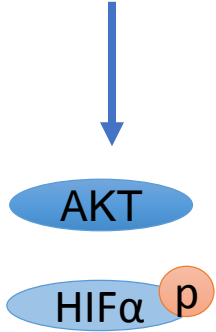
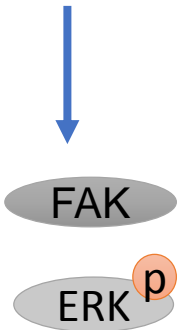
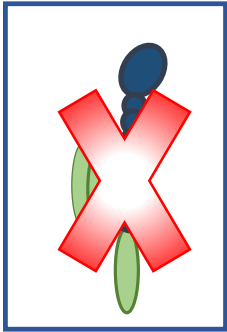
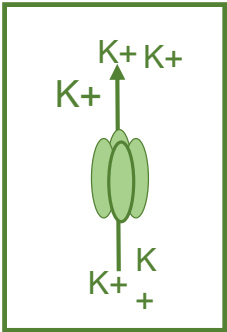
→ Pathway activation/progression  
 - - - - - Protein translocalization  
 → Upregulated cellular processes  
 P Phosphate group      G<sub>i</sub> Inhibitory G protein  
 ER Endoplasmic reticulum      PM Plasma membrane  
 $\beta 1$  integrin      hERG1 channel      Sigma 1 Receptor (Sig1R)

# TUMOR



OPEN CHANNEL

CLOSED CHANNEL LINKED TO AN ACTIVE INTEGRIN

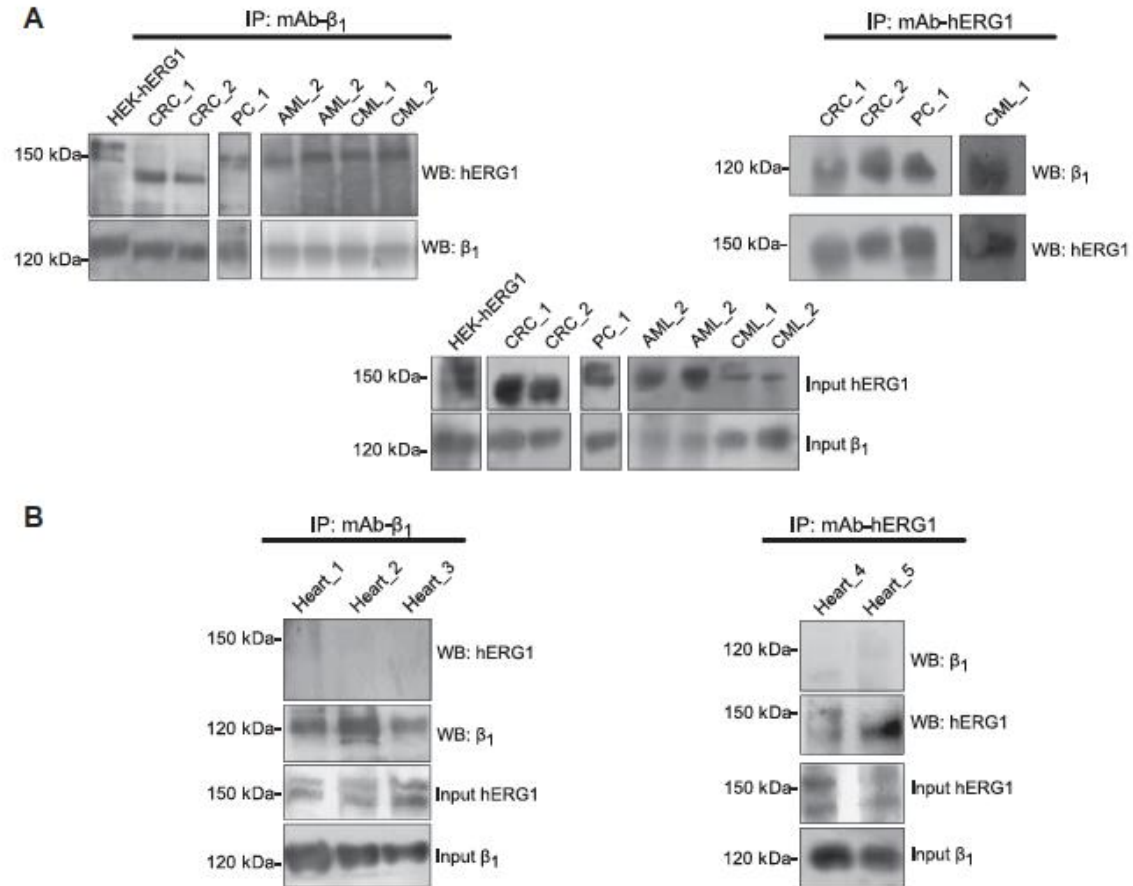


*Proliferation  
Survival*

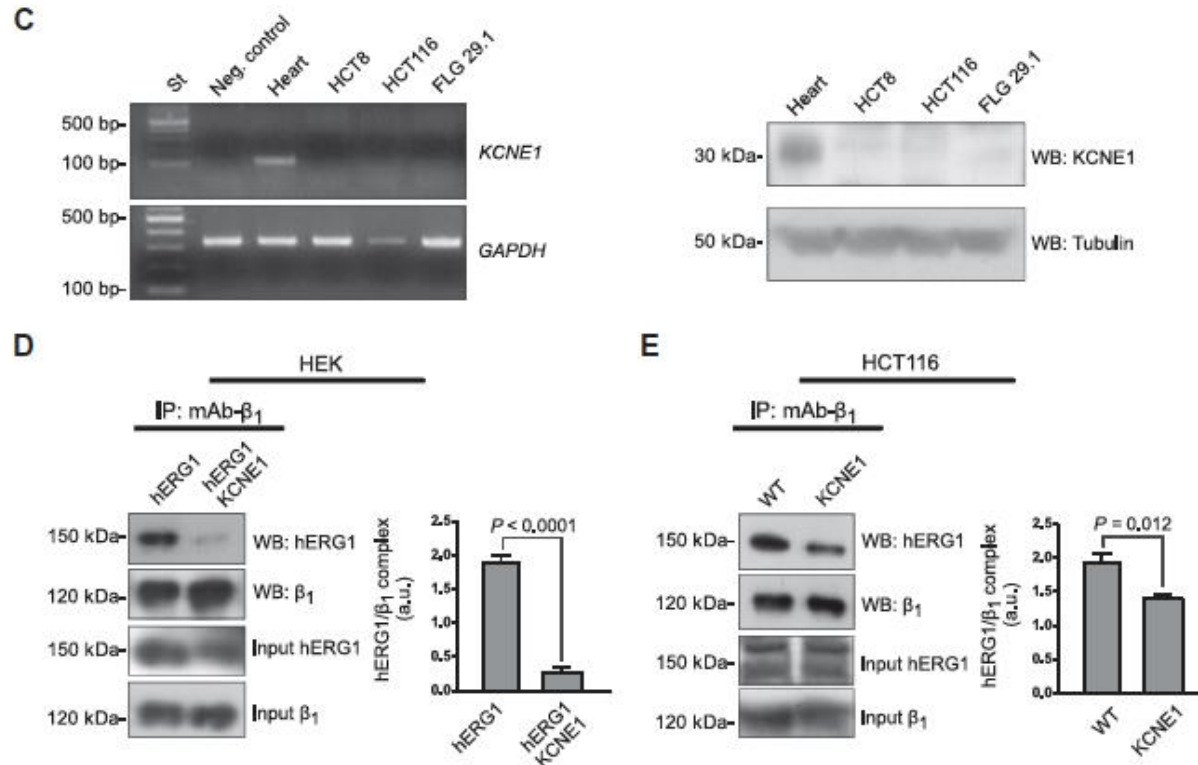
*Invasiveness,  
Angiogenesis  
Metastasis*

*The hERG1/ $\beta$ 1 complex occurs in tumour cells, but not in the heart*

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... because tumour cells do not express “canonical”  
(KCNE1) beta subunits



TUMOR

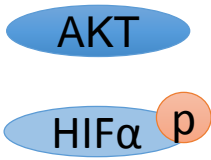
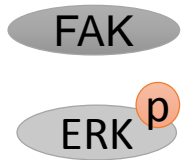
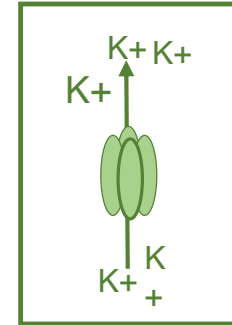
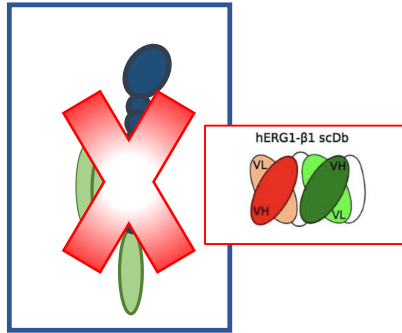
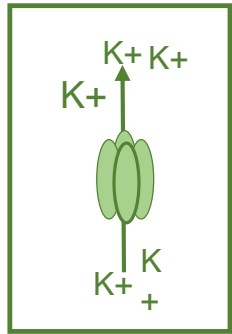
HEART

hERG1

OPEN CHANNEL

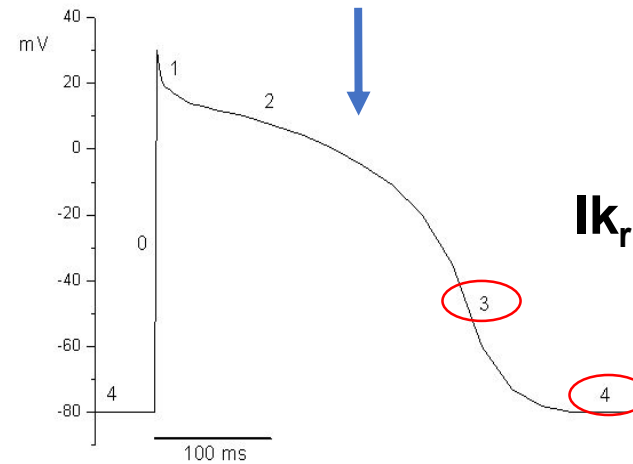
CLOSED CHANNEL LINKED TO AN ACTIVE INTEGRIN

OPEN CHANNEL ( $\Delta V$ )



Proliferation  
Survival

Invasiveness,  
Angiogenesis  
Metastasis



# Different physiological roles of hERG

- **Cardiac cells: regulates repolarization**
- **Other excitable cells: regulates  $V_{rest}$  and firing**
- **Cancer cells: regulates slowly changing  $V_{rest}$  and exerts non-conductive roles (e.g. molecular complexes with integrin receptors)**

# *Targeting hERG1 in cancer*

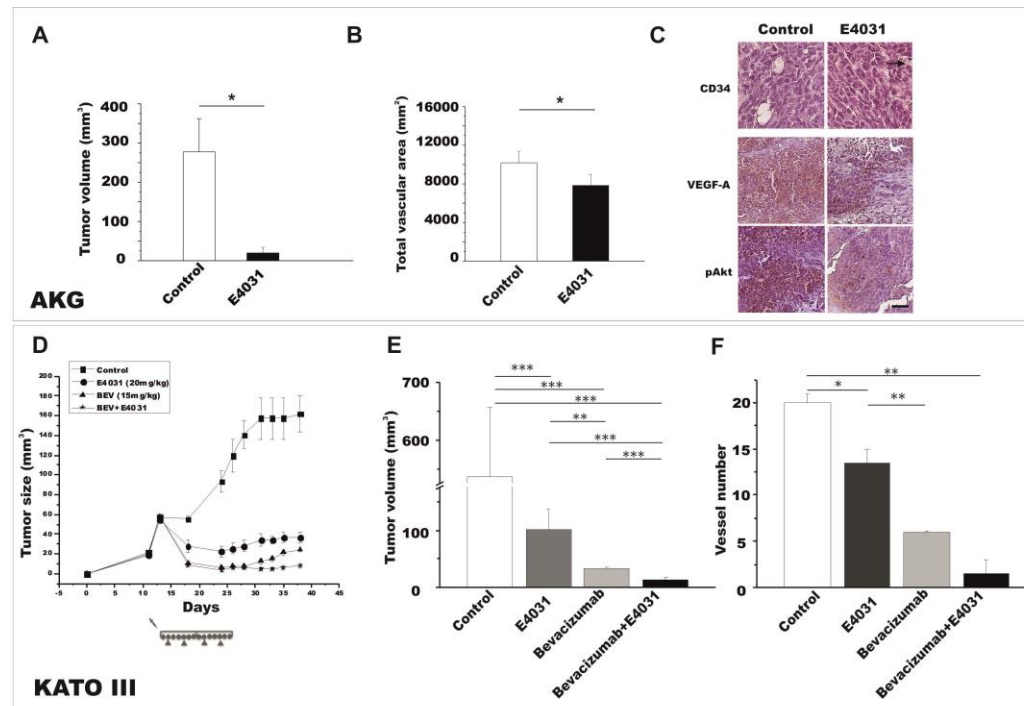
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# *hERG1 in cancer*

- Several preclinical data (mouse xenografts) indicate that blocking hERG1 inhibits tumour growth and metastatic spread



*Pillozzi et al., Blood, 2007;*

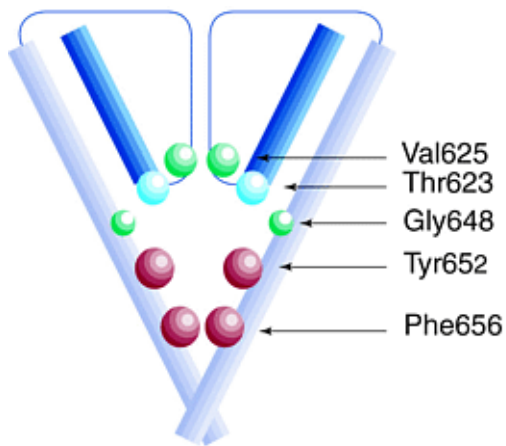
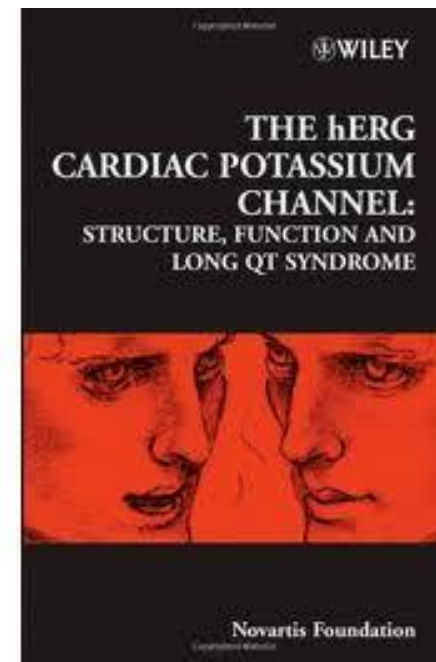
*Pillozzi et al., Blood, 2011;*

*Crociani et al., Sci.Rep.2013;*

*Crociani et al., Clin. Cancer Res., 2014;*

*Becchetti et al., Sci. Signal., 2017*

hERG1 is considered an antitarget!  
hERG1 blockers can induce LQT syndrome and TdP



TRENDS in Pharmacological Sciences

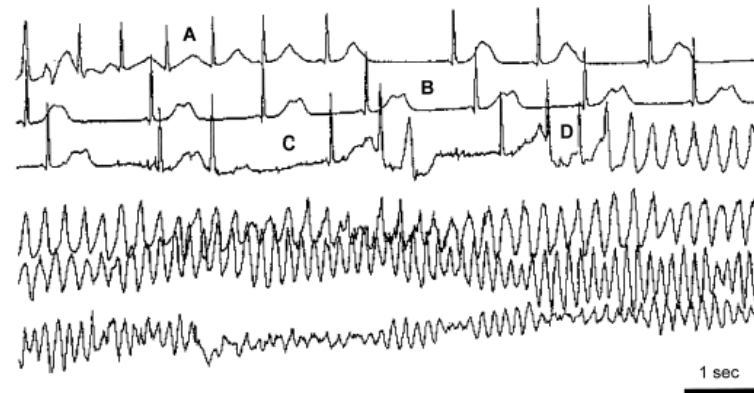


Fig. 2 Part of a continuous single-channel Holter recording from a patient with long QT syndrome. In this record, the characteristic prolonged QT interval (A), followed by giant late-repolarization "T-wave humps" (B), led to premature beats with a bigeminal pattern with short-long-short sequences of R-R intervals (C) before onset of *torsade de pointes* (D). The episode progressed into ventricular fibrillation before spontaneously resolving into sinus rhythm; the patient was later treated successfully with pacing and beta-blockers (Reproduced with permission from Benhorin and Medina.<sup>179</sup>)

# Strategies to target hERG1 in cancer

✓ *Use of non cardiotoxic hERG1 blockers*

✓ *Targeting the molecular differences between “tumour” and “cardiac” hERG1:*

CCR Translations

Clinical  
Cancer  
Research

## hERG Channels: From Antitargets to Novel Targets for Cancer Therapy

Annarosa Arcangeli<sup>1</sup> and Andrea Becchetti<sup>2</sup>

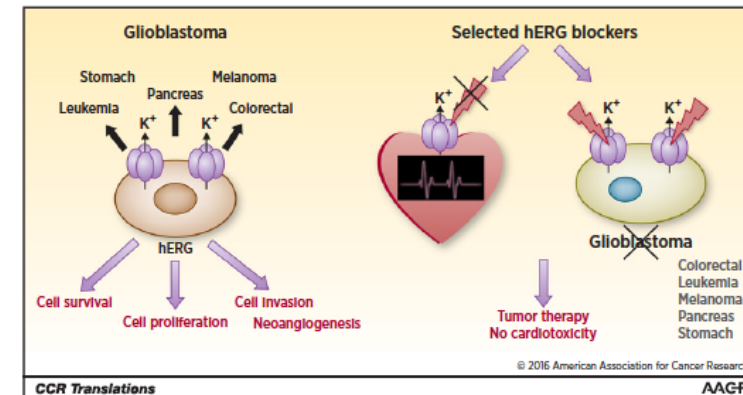


Figure 1.

Left, hERG is often overexpressed on the plasma membrane of different human cancer cells. It regulates tumor cell proliferation, survival, migration/invasiveness, and neoangiogenesis. Right, inhibiting hERG in different types of cancer cells (red lightning bolts) by using selective blockers that do not produce cardiac arrhythmia (as indicated by the black cross) is a possible strategy for anticancer therapy. The article by Pointer and colleagues (1) suggests that this is feasible in glioblastoma. Such a strategy may be effective in other cancers (shown in gray) in which hERG is overexpressed and has been shown to regulate neoplastic progression.

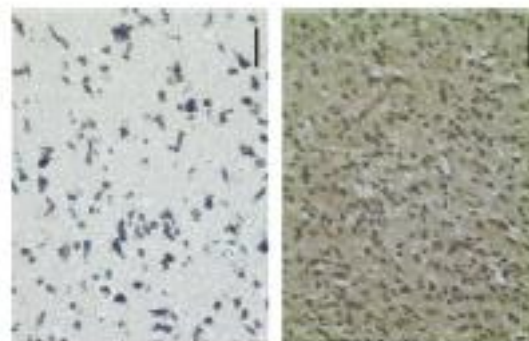
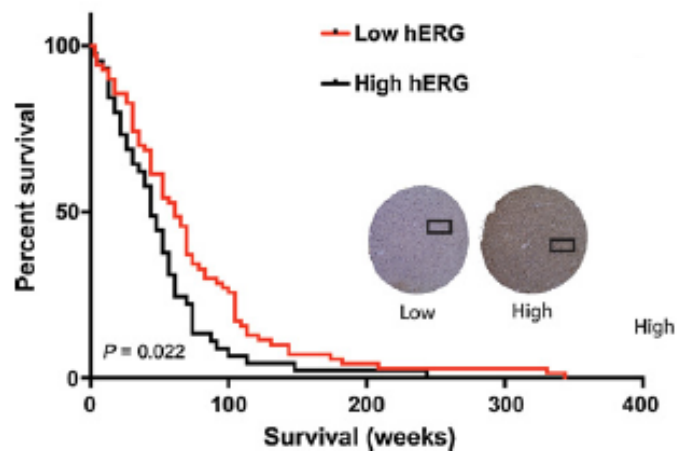
## *Strategies to target hERG1 in cancer*

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*✓ Use of non cardiotoxic (non torsadogenic) hERG1 blockers*

## Administration of Non-Torsadogenic human Ether-à-go-go-Related Gene Inhibitors Is Associated with Better Survival for High hERG-Expressing Glioblastoma Patients

Kelli B. Pointer<sup>1,2,3,4</sup>, Paul A. Clark<sup>1</sup>, Kevin W. Eliceiri<sup>4,5</sup>, M. Shahriar Salamat<sup>6</sup>, Gail A. Robertson<sup>7,8</sup>, and John S. Kuo<sup>1,2,3,5,9</sup>



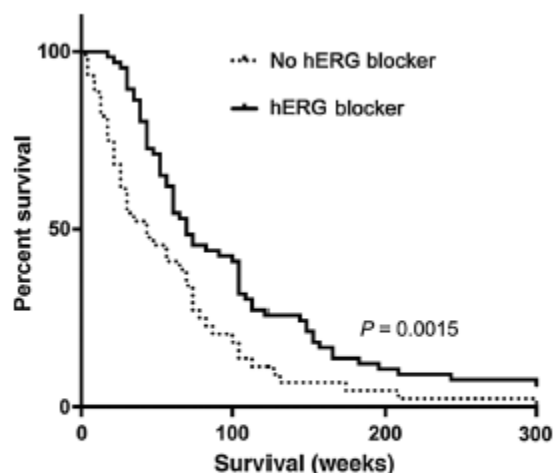
**Table 1.** Univariate and multivariate Cox regression analyses of glioblastoma hERG expression and confounding variables

|                 | Univariate analysis |                                 |              | Multivariate analysis |        |
|-----------------|---------------------|---------------------------------|--------------|-----------------------|--------|
|                 | N                   | Median survival, weeks (95% CI) | P (log-rank) | HR (95% CI)           | P      |
| hERG expression |                     |                                 | 0.022        | 2.122 (1.247–3.610)   | 0.003  |
| High            | 45                  | 43.5 (33.8–53.1)                |              |                       |        |
| Low             | 71                  | 60.9 (48.2–73.6)                |              |                       |        |
| Gender          |                     |                                 | 0.463        |                       | NS     |
| Male            | 81                  | 52.2 (44.5–59.9)                |              |                       |        |
| Female          | 35                  | 56.5 (34.5–78.5)                |              |                       |        |
| Age             |                     |                                 | 0.957        |                       | NS     |
| ≤55             | 51                  | 52.2 (39.5–64.8)                |              |                       |        |
| >55             | 58                  | 52.2 (44.1–60.3)                |              |                       |        |
| KPS             |                     |                                 | 0.024        | 1.143 (0.769–1.700)   | 0.508  |
| ≤70             | 55                  | 60.9 (48.3–73.5)                |              |                       |        |
| >70             | 60                  | 43.5 (29.7–57.3)                |              |                       |        |
| Temozolomide    |                     |                                 | <0.001       | 2.845 (1.849–4.378)   | <0.001 |
| Yes             | 66                  | 69.6 (61.6–77.6)                |              |                       |        |
| No              | 50                  | 34.8 (27.3–42.3)                |              |                       |        |
| Radiation       |                     |                                 | <0.001       | 2.122 (1.247–3.610)   | 0.006  |
| Yes             | 95                  | 60.9 (9.7–25.1)                 |              |                       |        |
| No              | 21                  | 17.4 (4.96–72.2)                |              |                       |        |
| Tobacco use     |                     |                                 | 0.519        |                       | NS     |
| Yes             | 46                  | 52.2 (41.4–63.0)                |              |                       |        |
| No              | 70                  | 52.2 (37.9–66.5)                |              |                       |        |

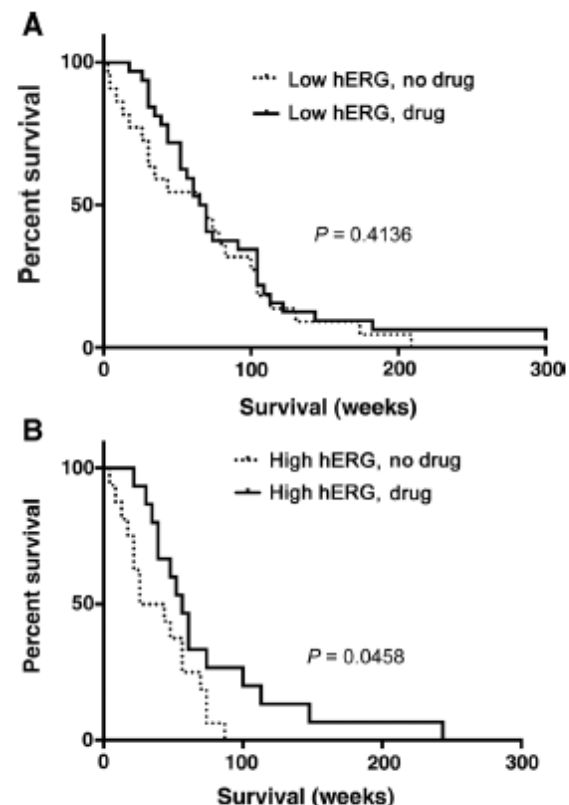
Abbreviation: CI, confidence interval; NS, not significant.

## Administration of Non-Torsadogenic human Ether-à-go-go-Related Gene Inhibitors Is Associated with Better Survival for High hERG-Expressing Glioblastoma Patients

Kelli B. Pointer<sup>1,2,3,4</sup>, Paul A. Clark<sup>1</sup>, Kevin W. Eliceiri<sup>4,5</sup>, M. Shahriar Salamat<sup>6</sup>, Gail A. Robertson<sup>7,8</sup>, and John S. Kuo<sup>1,2,3,5,9</sup>



**Figure 4.** Receipt of hERG blockers correlates with a better glioblastoma patient survival. Patients who received more than one hERG blocker were compared with patients who had not received hERG blockers, and there was a statistically significant difference in their survival ( $P = 0.0015$ ).

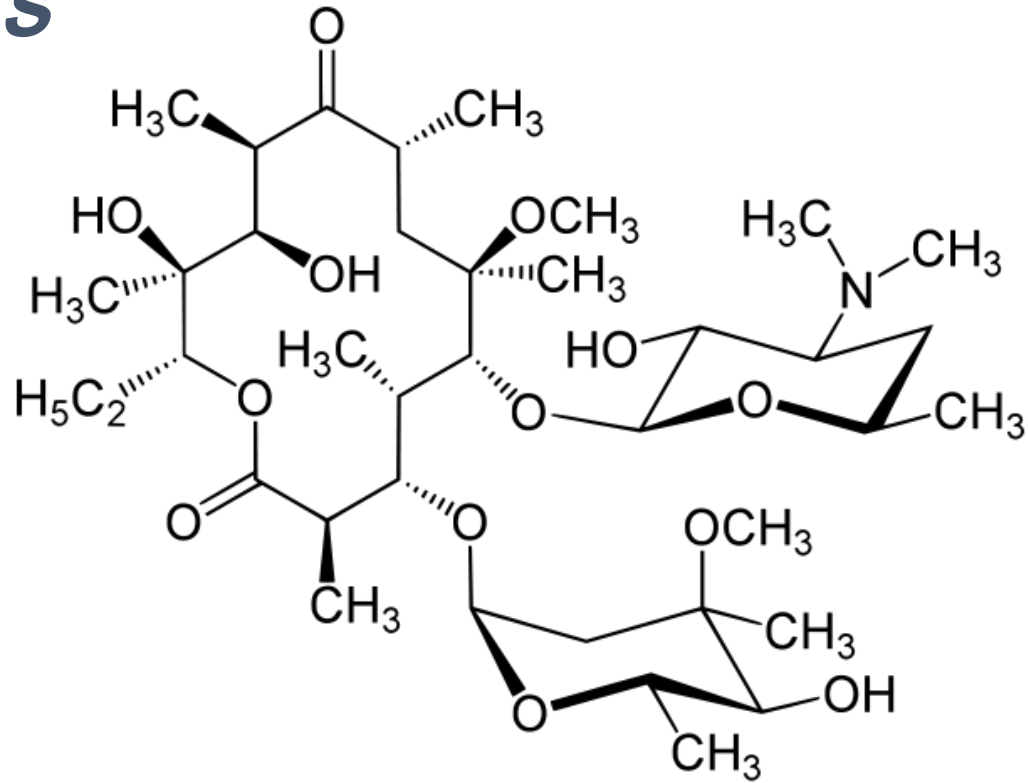


**Figure 5.** Patient hERG expression levels correlate with the benefit of receiving an hERG. Patients were stratified based on whether or not they had high or low hERG expression levels. In each group, patients were further stratified on the basis of whether they had received an hERG blocker. **A**, No statistically significant difference was found in survival between patients who had low hERG expression levels ( $P = 0.4136$ ). **B**, There was a statistically significant difference in survival for patients with high hERG expression levels based on whether they received an hERG blocker ( $P = 0.0458$ ).

# Strategies to target *hERG1* in cancer

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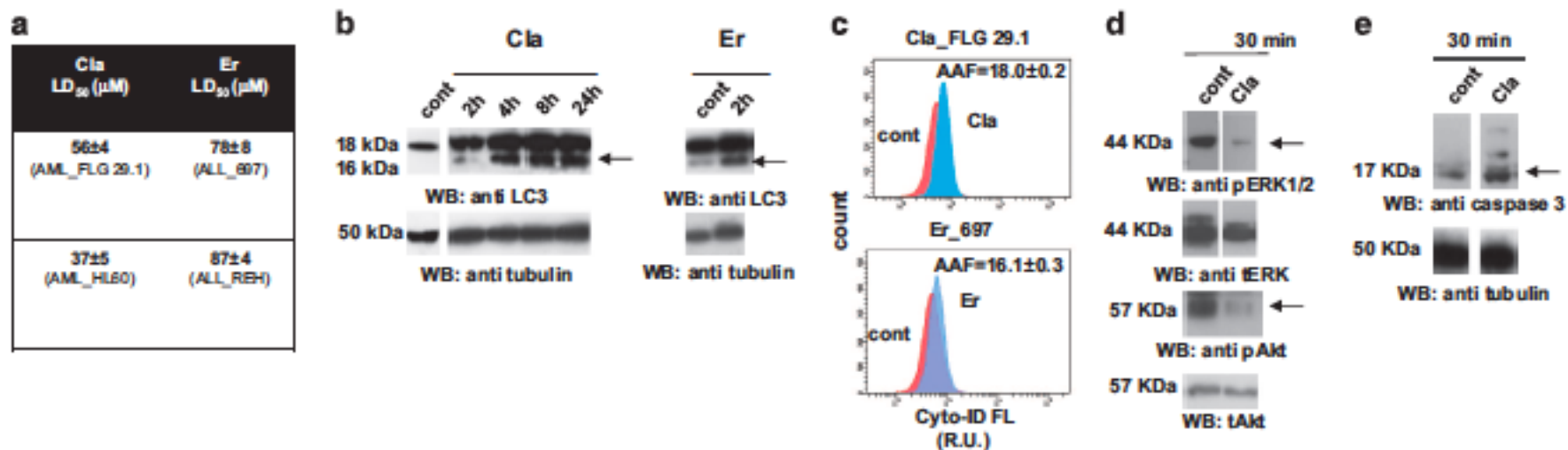
✓ *Use of non cardiotoxic (non torsadogenic) hERG1 blockers*



Clarythromycin

## LETTER TO THE EDITOR

## Macrolide antibiotics exert antileukemic effects by modulating the autophagic flux through inhibition of hERG1 potassium channels





## *Strategies to target hERG1 in cancer*

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- ✓ Use of non cardiotoxic (non torsadogenic) hERG1 blockers*
- ✓ Targeting the molecular differences between “tumour” and “cardiac” hERG1:*
- ✓ Preferential expression of hERG1B in leukemias (CD160130)*

# High *hHERG1B* expression in leukemias

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## Cell Cycle-dependent Expression of *HERG1* and *HERG1B* Isoforms in Tumor Cells\*

Received for publication, October 22, 2001, and in revised form, November 12, 2001  
Published, JBC Papers in Press, November 12, 2002, DOI: 10.1074/jbc.M210780200

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Massimo Olivottoli<sup>1</sup>, Randy S. Wymore<sup>2</sup>, and Annarosa Arcangeli<sup>1,3\*</sup>

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2003

OPEN

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[www.nature.com/leu](http://www.nature.com/leu)



2014

LETTER TO THE EDITOR

Differential expression of *hERG1A* and *hERG1B* genes in pediatric acute lymphoblastic leukemia identifies different prognostic subgroups

*Pediatric Hematology and Oncology*, 32:182–192, 2015  
Copyright © Informa Healthcare USA, Inc.  
ISSN: 0888-0018 print / 1521-0669 online  
DOI: 10.3109/08880018.2014.949941

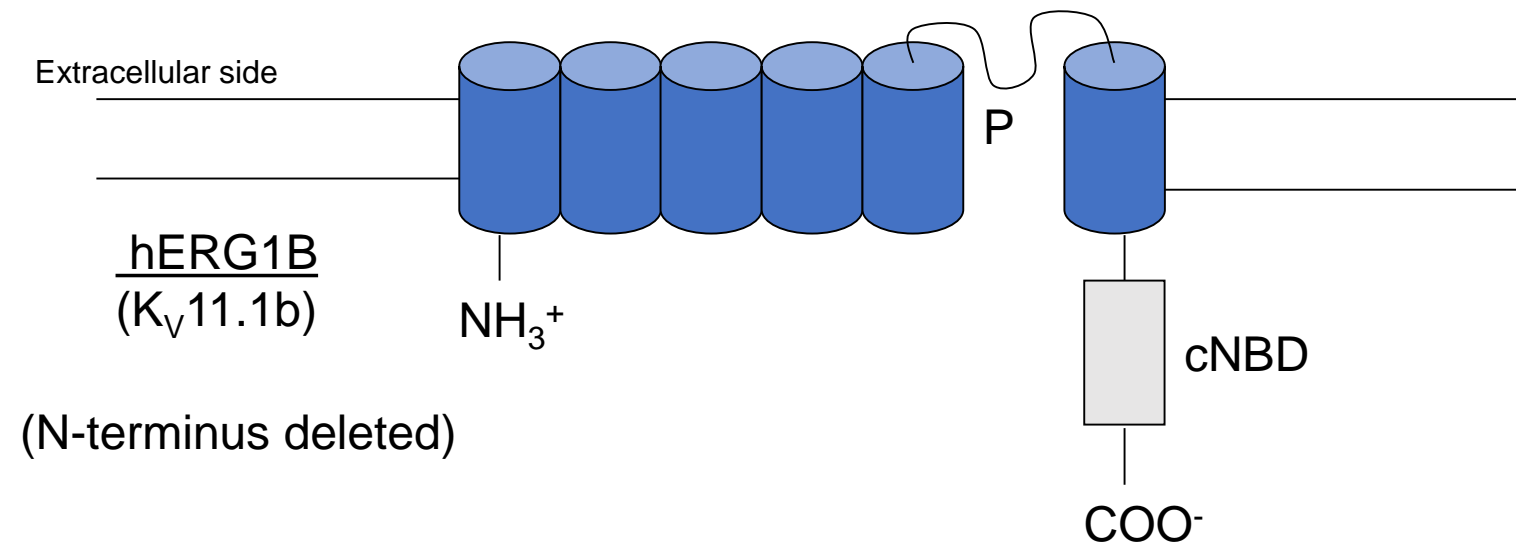
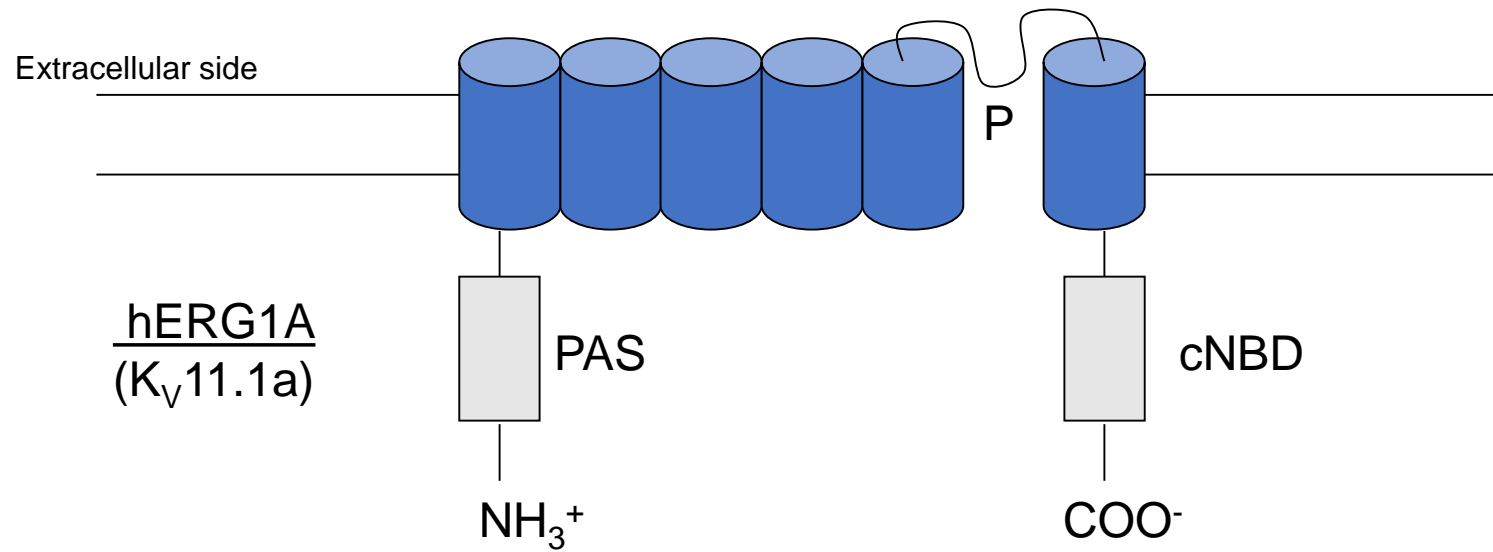
informa  
healthcare

ORIGINAL ARTICLE

## *herg1b* Expression as a Potential Specific Marker in Pediatric Acute Myeloid Leukemia Patients with *HERG* 897K/K Genotype

2015

Merve Erdem,<sup>1,\*</sup> Tugce Ayca Tekiner,<sup>1,2</sup> Arta Fejzullahu,<sup>1,3</sup> Gokce Akan,<sup>4</sup>  
Sema Anak,<sup>5</sup> Ebru Tugrul Saribeyoglu,<sup>5</sup> Ugur Ozbek,<sup>6</sup>  
and Fatmahan Atalar<sup>1</sup>

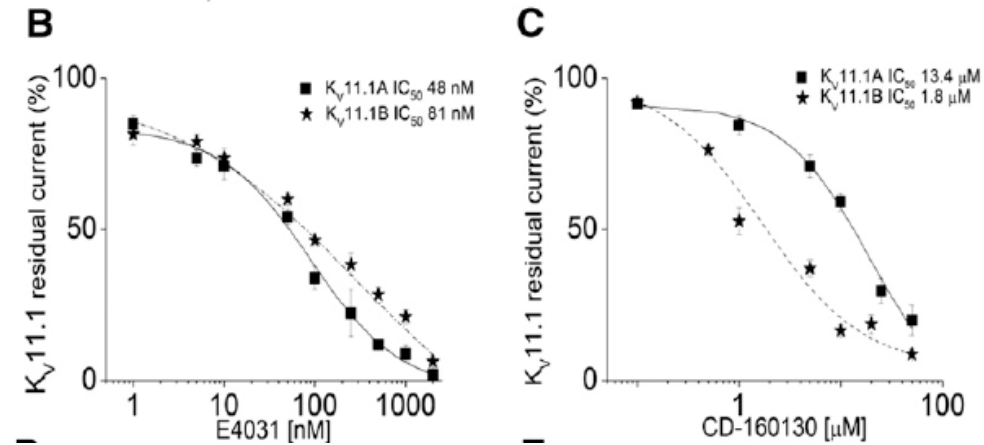


## New Pyrimido-Indole Compound CD-160130 Preferentially Inhibits the $K_v11.1B$ Isoform and Produces Antileukemic Effects without Cardiotoxicity<sup>ISI</sup>

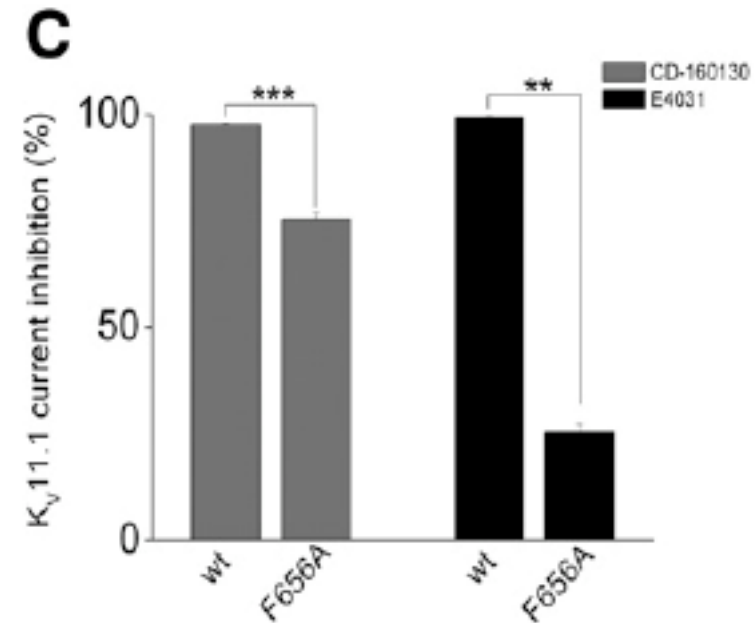
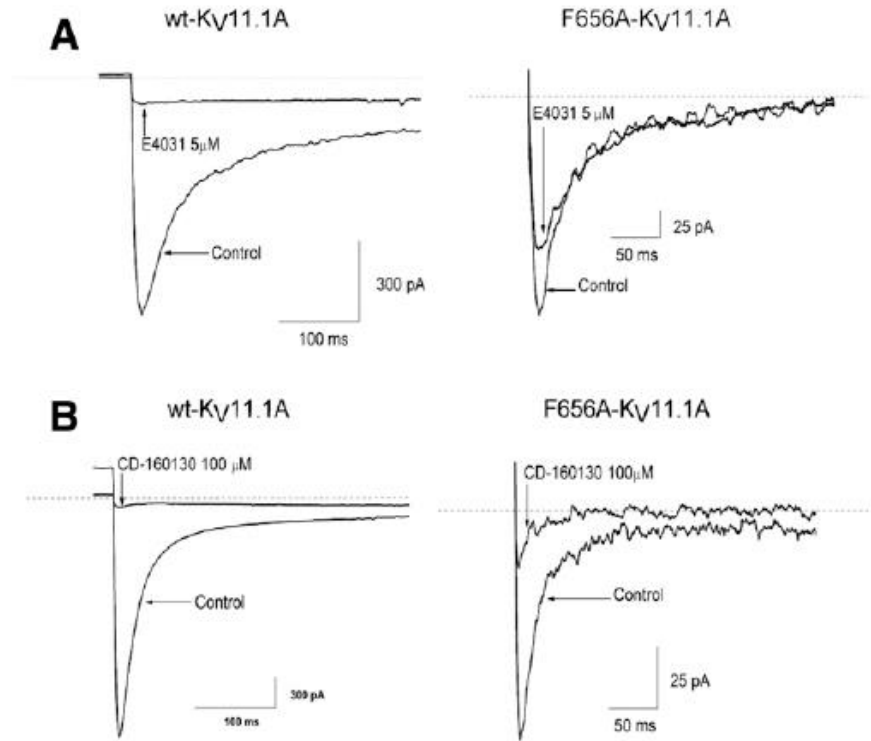
Luca Gasparoli, Massimo D'Amico, Marika Masselli, Serena Pillozzi, Rachel Caves, Rawan Khuwaileh, Wolfgang Tiedke, Kenneth Mugridge, Alessandro Pratesi, John S. Mitcheson, Giuseppe Basso, Andrea Becchetti, and Annarosa Arcangeli

*Department of Experimental and Clinical Medicine, University of Florence, Florence, Italy (L.G., S.P., A.A.); Department of Chemistry "Ugo Schiff," University of Florence, Florence, Italy (M.M., A.P.); DI.V.A.L. Toscana srl, Sesto Fiorentino, Italy (M.D.A., M.M.); Department of Cell Physiology and Pharmacology, University of Leicester, Leicester, United Kingdom (R.C., R.K., J.S.M.); BlackSwan Pharma GmbH, Leipzig, Germany (W.T., K.M.); Oncohematology Laboratory, Department of Woman and Child Health, University of Padova, Padova, Italy (G.B.); and Department of Biotechnologies and Biosciences, University of Milano-Bicocca, Milan, Italy (A.B.)*

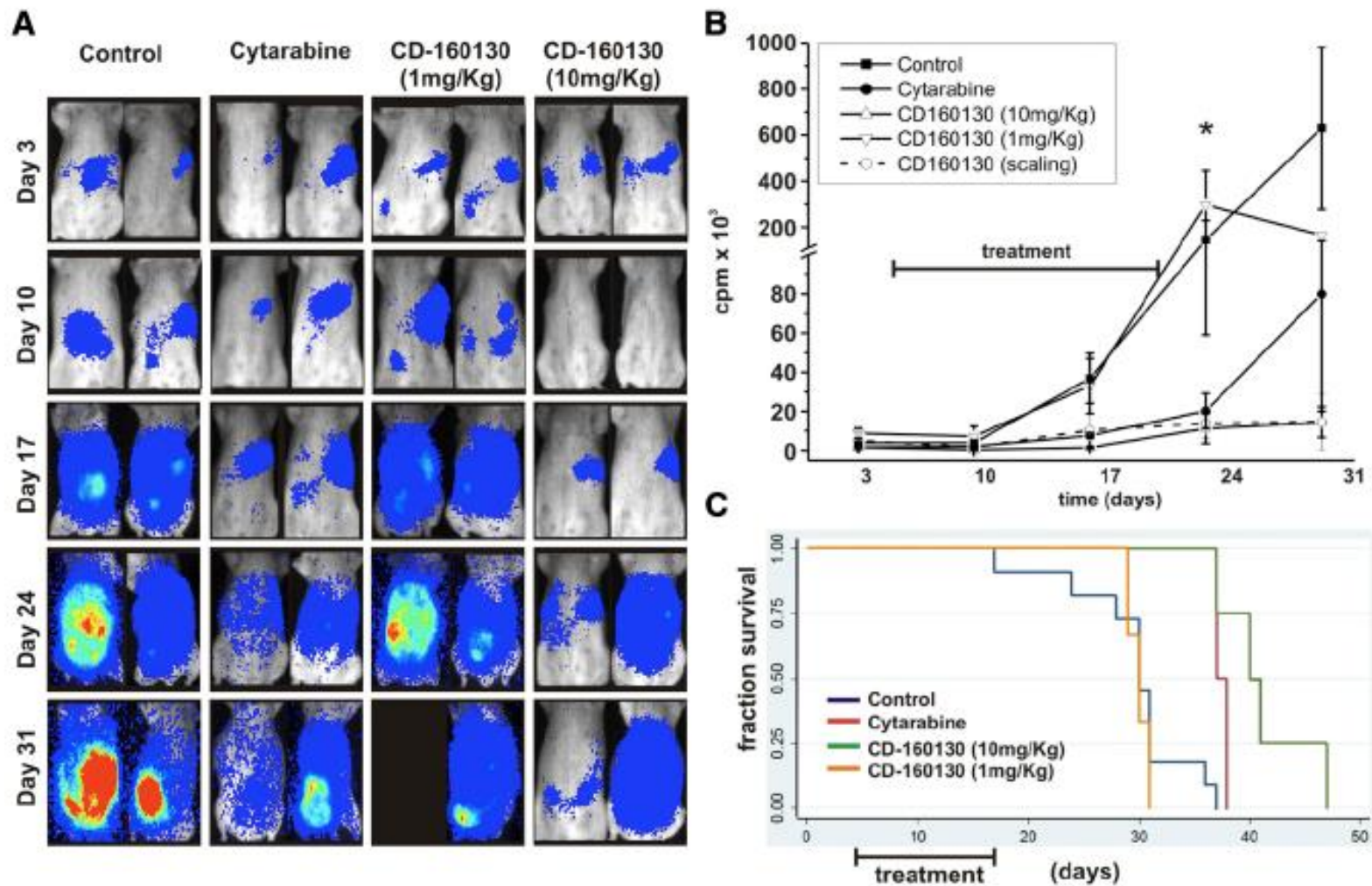
Received July 22, 2014; accepted November 19, 2014



# *CD 160130 does not bind the F656 “canonical” binding site in hERG1*



# CD 160130 hampers leukemia burden in vivo



*...without lengthening the QT interval*

*(in guinea pigs)*

---

**B**

**ECG parameters**

| CD-160130 (n=5)<br>10mg/kg | QT ± SEM (ms) | HR ± SEM<br>(beat/min) | QTc ± SEM    | ΔQTc<br>(% vs Pre-drug) |
|----------------------------|---------------|------------------------|--------------|-------------------------|
| Pre-drug                   | 129.0 ± 4.2   | 237.1 ± 3.4            | 305.1 ± 7.5  | -                       |
| 0 min                      | 131.4 ± 5.4   | 241.1 ± 5.2            | 308.4 ± 7.4  | 1.1 ± 3.5               |
| 5 min                      | 127.5 ± 7.0   | 249.1 ± 1.1            | 298.8 ± 12.9 | -2.1 ± 4.9              |
| 10 min                     | 129.0 ± 5.8   | 240.5 ± 3.4            | 300.9 ± 11.2 | -1.4 ± 4.4              |
| 15 min                     | 128.0 ± 6.7   | 240.5 ± 2.8            | 298.6 ± 11.8 | -2.2 ± 4.6              |
| Sotalol (n=5)<br>3mg/kg    | QT ± SEM (ms) | HR ± SEM<br>(beat/min) | QTc ± SEM    | ΔQTc<br>(% vs Pre-drug) |
| Pre-drug                   | 115.4 ± 3.6   | 240.3 ± 3.7            | 276.4 ± 6.8  | -                       |
| 0 min                      | 115.4 ± 3.8   | 240.4 ± 5.1            | 273.7 ± 6.7  | -1.0 ± 3.4              |
| 5 min                      | 132.7 ± 4.0   | 240.4 ± 1.2            | 301.1 ± 6.2  | 8.9 ± 3.5               |
| 10 min                     | 135.3 ± 4.2   | 235.0 ± 3.1            | 304.3 ± 6.1  | 10.1 ± 3.5              |
| 15 min                     | 133.0 ± 5.8   | 232.2 ± 4.5            | 299.5 ± 8.0  | 8.4 ± 3.9               |

**CD-160130**

# *Strategies to target hERG1 in cancer*

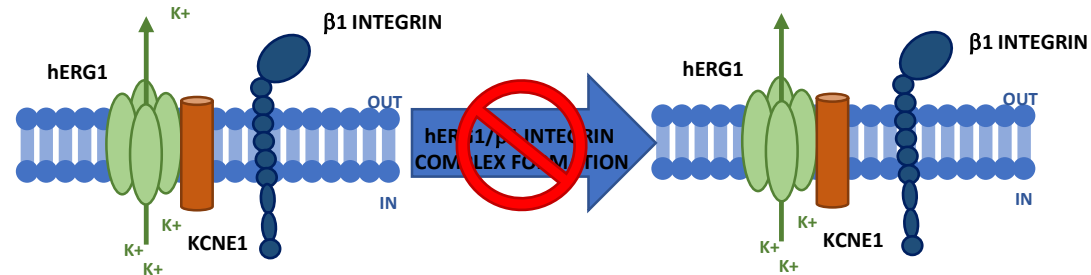
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- ✓ Use of non cardiotoxic (non torsadogenic) hERG1 blockers*
- ✓ Targeting the molecular differences between “tumour” and “cardiac” hERG1:*
- ✓ Preferential expression of hERG1B in leukemias (CD160130)*
- ✓ Formation of a hERG1/ $\beta$ 1 integrin complex in tumour cells*

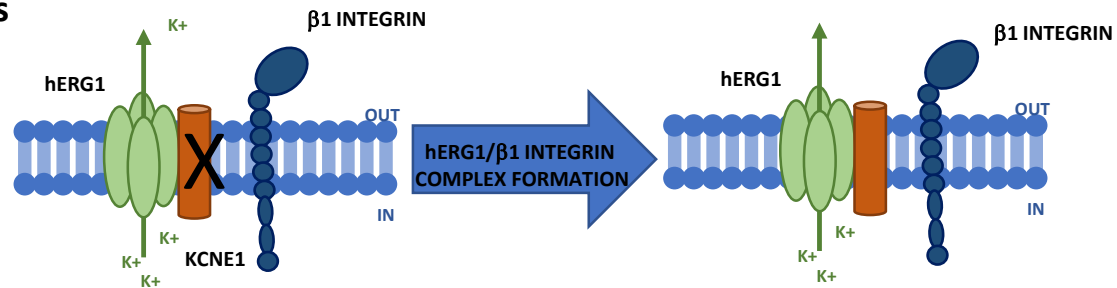


# hERG1 and $\beta$ 1 integrin associate in human cancer tissue but not cardiac tissue.

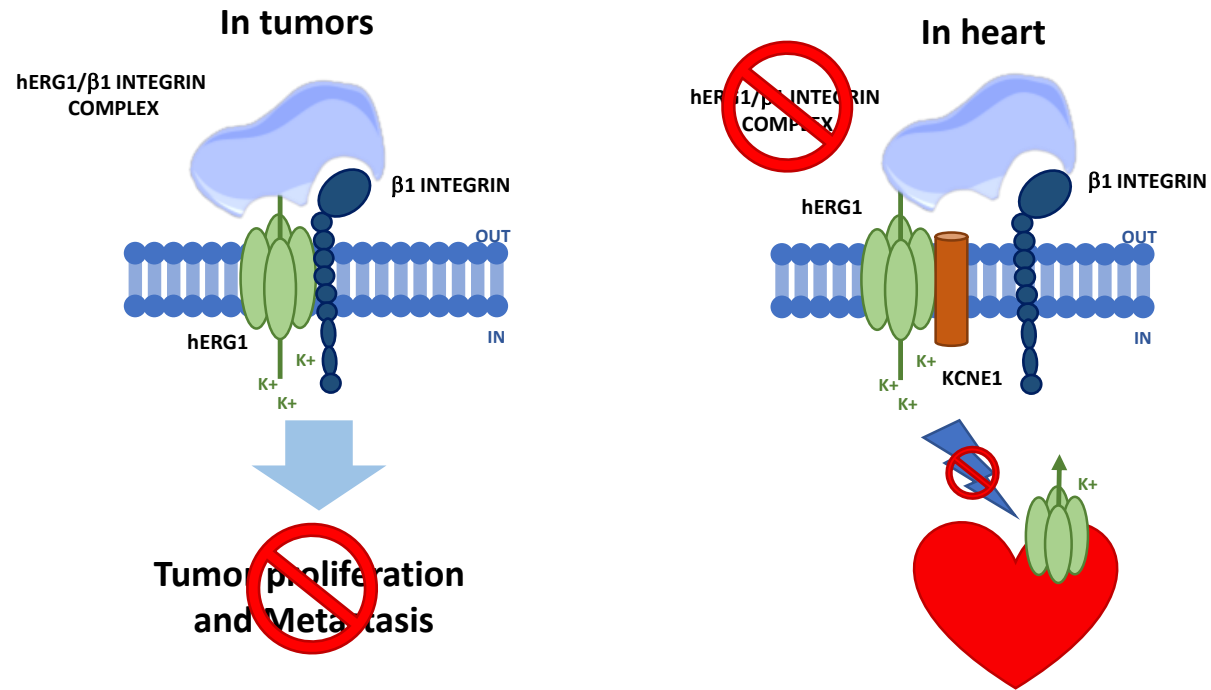
In heart



In tumors



# Future .... Blockade of the hERG1/ $\beta$ 1 integrin complex



TUMOR

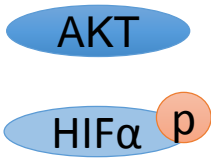
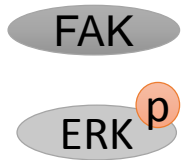
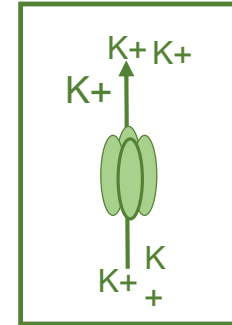
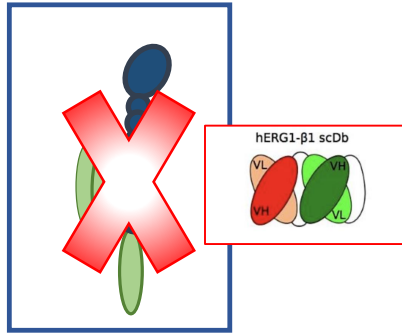
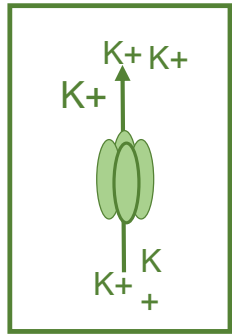
HEART

hERG1

OPEN CHANNEL

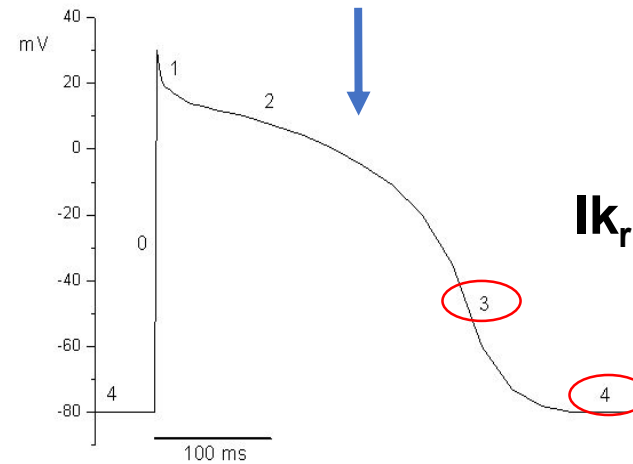
CLOSED CHANNEL LINKED TO AN ACTIVE INTEGRIN

OPEN CHANNEL ( $\Delta V$ )



Proliferation  
Survival

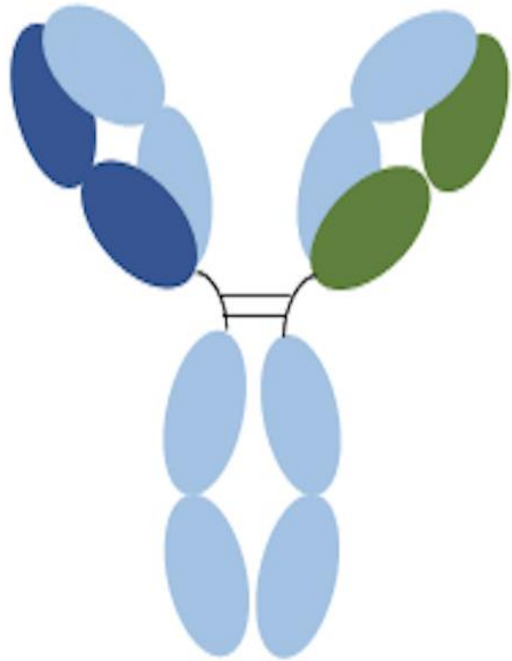
Invasiveness,  
Angiogenesis  
Metastasis



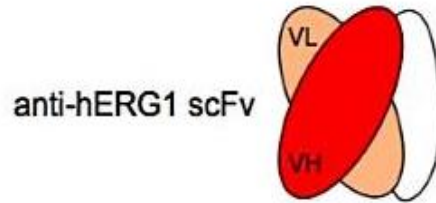
# PIPELINE DEVELOPMENT STRATEGY

DESIGNING NEXT GENERATION ANTIBODY MOLECULES

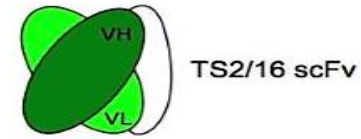
FROM BIG MONOCLONAL ANTIBODY TO SMALL BISPECIFIC DIABODY



**Anti-hERG1 Monoclonal Antibody**  
(M.W. 160 kDa)



**Anti-hERG1 Single-Chain Variable Fragment (Sc-FV)**

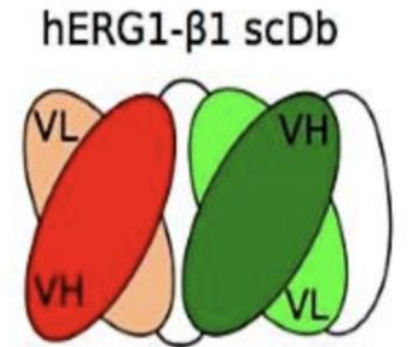


**Anti-integrin beta1 Single-Chain Variable –Fragment (Sc-fV)**



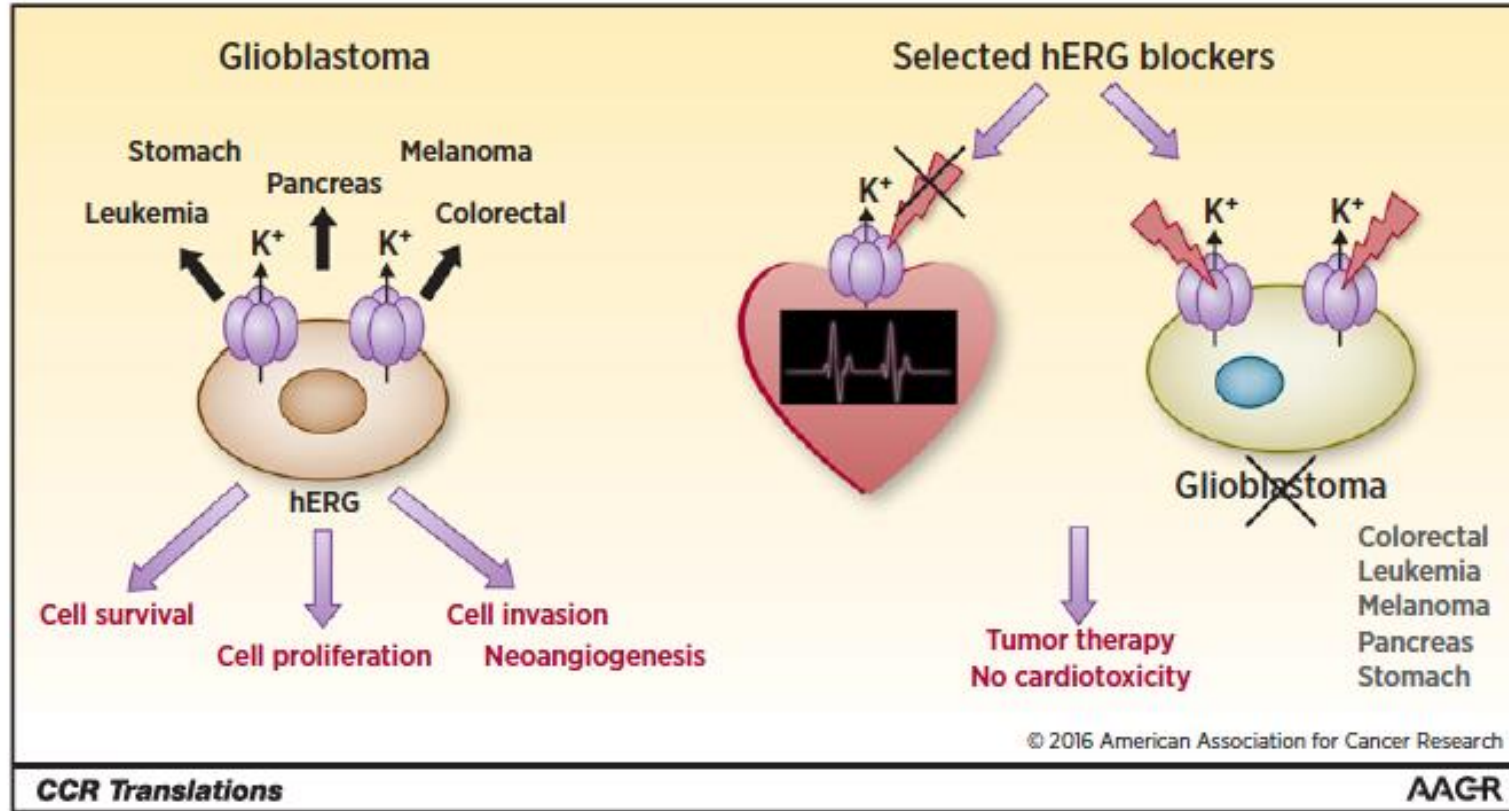
**Anti hERG1/integrin beta1 BISPECIFIC Antibody (DIABODY)**  
(M.W. 50 kDa)

**THERAPEUTIC ANTIBODY**  
**MCKAA2017THE01**



# hERG Channels: From Antitargets to Novel Targets for Cancer Therapy

Annarosa Arcangeli<sup>1</sup> and Andrea Becchetti<sup>2</sup>



# CONCLUSIONS

- ❖ **Ion channels are relevant in cancer (biomarkers!)**
- ❖ **Ion channels can exert both conductive and non conductive effects in cancer cells**
- ❖ **hERG1 has both conductive (ion flux-mediated) and non conductive (once bound to integrin receptors in the closed state) activities**
- ❖ **hERG1 mediates tumour progression (e.g. proliferation, invasion, angiogenesis, metastasis....)**
- ❖ **hERG1 can be considered a novel cancer biomarker**
- ❖ **The hERG1/beta1 integrin complex can be considered a therapeutic target in cancer and can be targeted through newly developed bispecific antibodies**

TUMOR

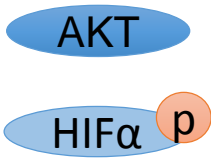
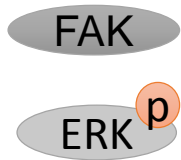
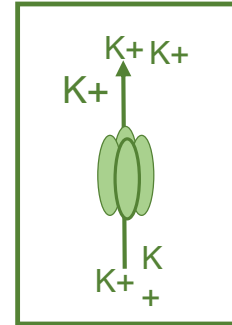
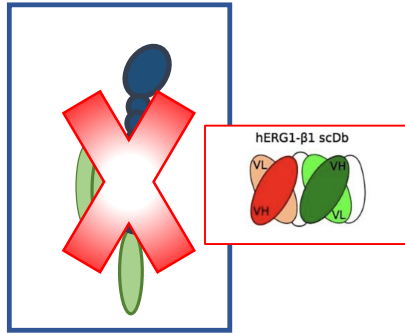
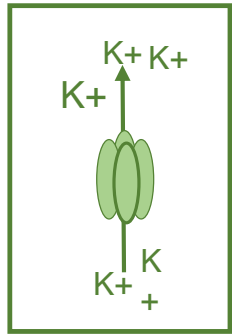
HEART

hERG1

OPEN CHANNEL

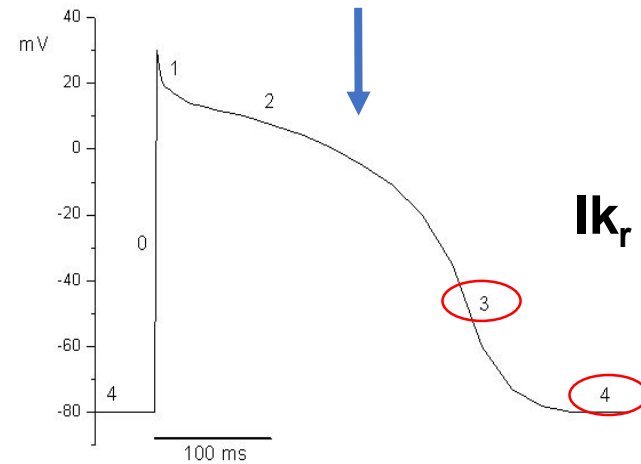
CLOSED CHANNEL LINKED TO AN ACTIVE INTEGRIN

OPEN CHANNEL ( $\Delta V$ )



Proliferation  
Survival

Invasiveness,  
Angiogenesis  
Metastasis



# Acknowledgements:



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Becchetti**  
University of  
Milano  
Bicocca, Italy



**Prof. F. Di  
Costanzo**  
Dept. Medical  
Oncology  
AOUC Firenze



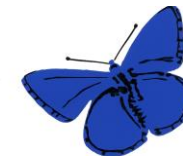
**Prof. R. Coppola**  
Dept. General  
Surgery  
Campus Biomedico  
Rome



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PER LA RICERCA SUL CANCRO**  
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# **TARGETING POTASSIUM CHANNELS TO OVERCOME CHEMORESISTANCE IN CRC**

# **(1) OVERCOMING CISPLATIN RESISTANCE BY TARGETING Kv11.1 AND KCa 3.1 CHANNELS**

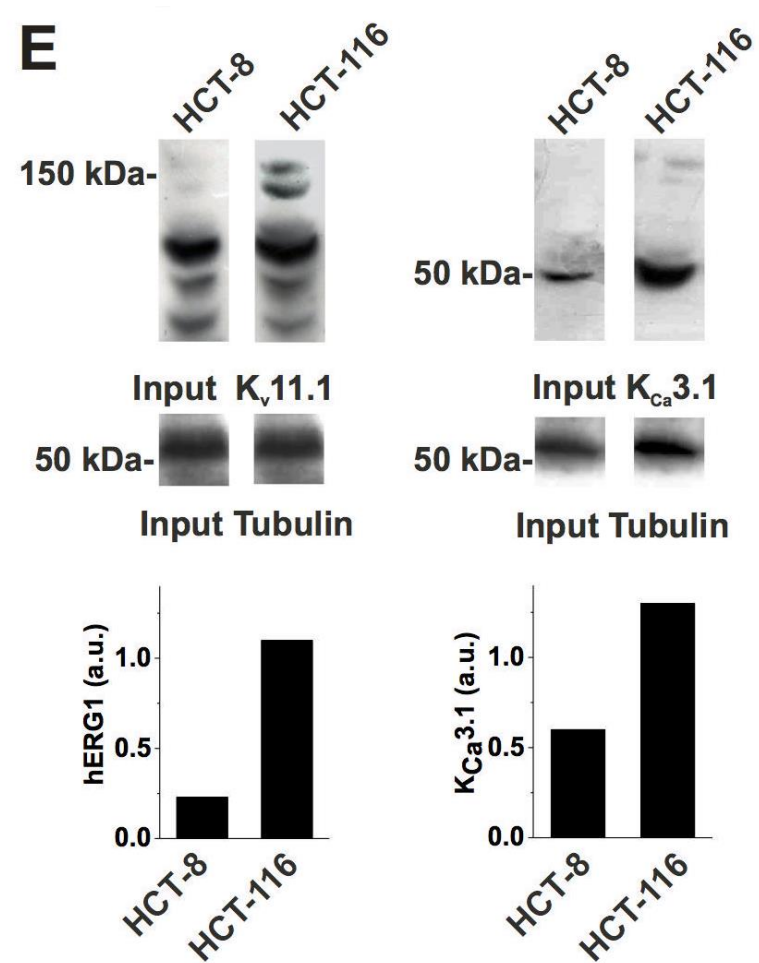
# The networking of Potassium channels



## **The combined activation of $K_{Ca}3.1$ and inhibition of $K_v11.1/hERG1$ currents contribute to overcome Cisplatin resistance in colorectal cancer cells**

Serena Pillozzi<sup>1,9</sup>, Massimo D'Amico<sup>2,9</sup>, Gianluca Bartoli<sup>1</sup>, Luca Gasparoli<sup>1</sup>, Giulia Petroni<sup>1</sup>, Olivia Crociani<sup>1</sup>, Tiziano Marzo<sup>3,4</sup>, Angela Guerriero<sup>1</sup>, Luigi Messori<sup>3</sup>, Mirko Severi<sup>5</sup>, Roberto Udisti<sup>5</sup>, Heike Wulff<sup>6</sup>, K George Chandy<sup>7</sup>, Andrea Becchetti<sup>8</sup> and Annarosa Arcangeli<sup>\*.1</sup>

# Cisplatin-resistant CRC cells (HCT116) express high levels of $K_{Ca}3.1$ and hERG1 channels



In resistant cells, KCa3.1 activators (SKA-31) and Kv11.1 inhibitors (E4031) had a synergistic action with Cisplatin in triggering apoptosis and inhibiting proliferation.

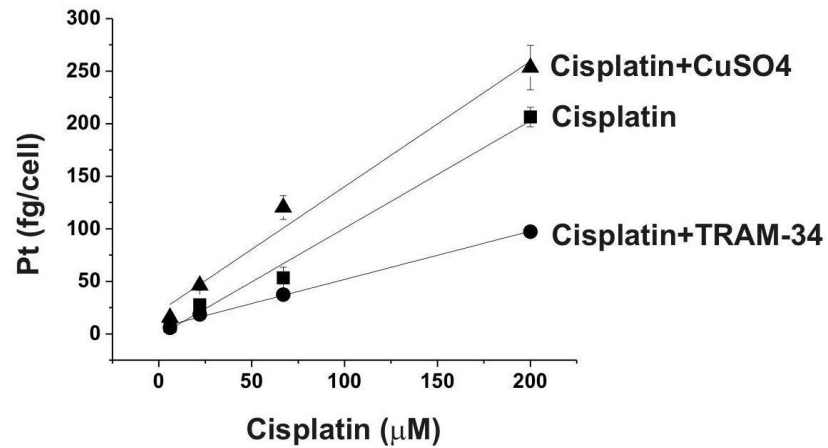
**Table 1B. Combination index and percentage (%) of apoptotic HCT-116 cells after different treatment combinations**

| Drug (concentration $\mu\text{M}$ )        | Combination index at $\text{IC}_{50}$ | Effect | Apoptosis                          |                                    |
|--|---------------------------------------|--------|------------------------------------|------------------------------------|
|  |                                       |        | Early apoptotic cells (%)          | Late apoptotic cells (%)           |
| Cisplatin (25)                             | —                                     |        | 5.9 $\pm$ 1.0                      | 5.3 $\pm$ 1.5                      |
| Cisplatin (25) + Riluzole (10)             | 0.70 $\pm$ 0.08                       | S      | 10.6 $\pm$ 1.3<br><i>P</i> = 0.021 | 17.6 $\pm$ 3.3<br><i>P</i> = 0.016 |
| Cisplatin (25) + SKA-31 (5)                | 0.64 $\pm$ 0.11                       | S      | 12.5 $\pm$ 3.9                     | 10.1 $\pm$ 2.4                     |
| Cisplatin (25) + TRAM-34 (25)              | 2.66 $\pm$ 0.78                       | A      | 13.8 $\pm$ 3.6<br><i>P</i> = 0.016 | 8.7 $\pm$ 1.6                      |
| Cisplatin (25) + E4031 (7)                 | 0.68 $\pm$ 0.07                       | S      | 8.0 $\pm$ 0.3                      | 13.2 $\pm$ 3.4<br><i>P</i> = 0.042 |
| Cisplatin (25) + Riluzole (10) + E4031 (7) | 0.47 $\pm$ 0.05                       | S      | ND                                 | ND                                 |
| Cisplatin (25) + SKA-31 (5) + E4031 (7)    | 0.69 $\pm$ 0.14                       | S      | ND                                 | ND                                 |
| Oxaliplatin (60) + Riluzole (10)           | 0.98 $\pm$ 0.01                       | S      | ND                                 | ND                                 |
| Oxaliplatin (60) + SKA-31 (5)              | 0.71 $\pm$ 0.05                       | S      | ND                                 | ND                                 |
| Oxaliplatin (60) + TRAM-34 (25)            | 3.36 $\pm$ 0.34                       | A      | ND                                 | ND                                 |
| Oxaliplatin (60) + E4031 (7)               | 0.83 $\pm$ 0.01                       | S      | ND                                 | ND                                 |

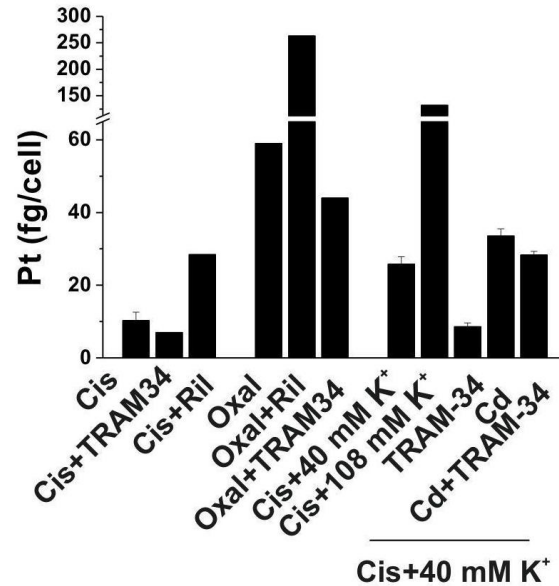
Abbreviation: ND = not determined.  $\text{CI} > 1$ , antagonism (A);  $\text{CI} = 1$ , additivity (Ad);  $\text{CI} < 1$ , synergy (S). HCT-116 cells were exposed to Cisplatin or Oxaliplatin in combination with Riluzole, SKA-31, TRAM-34 and E4031 for 24h as described in Pillozzi et al, 2011. All the drugs were used at drug concentrations indicated in the first column. Data are means  $\pm$  s.e.m. of three independent experiments, each carried out in triplicate. CI values were calculated using the CalcuSyn software Version 2 (Biosoft). For statistical analysis, Student's t-test was applied.

# Cisplatin uptake into resistant cells depend on $K_{Ca}3.1$ channel activity, and is potentiated by either $K_{Ca}3.1$ activators or hERG1 inhibitors

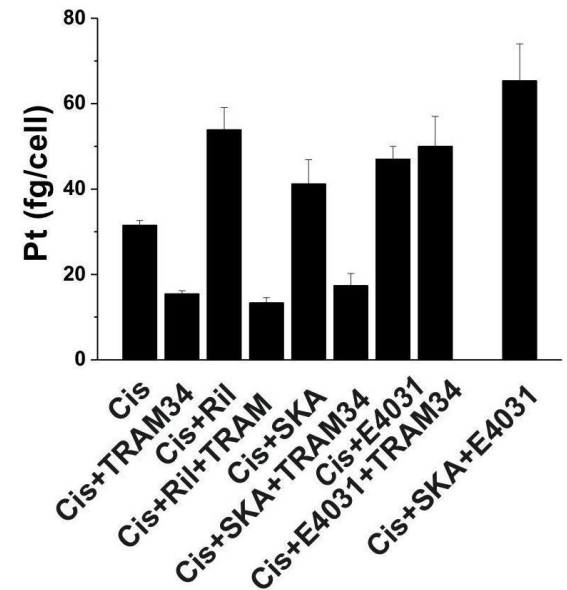
**A**



**B**



**C**



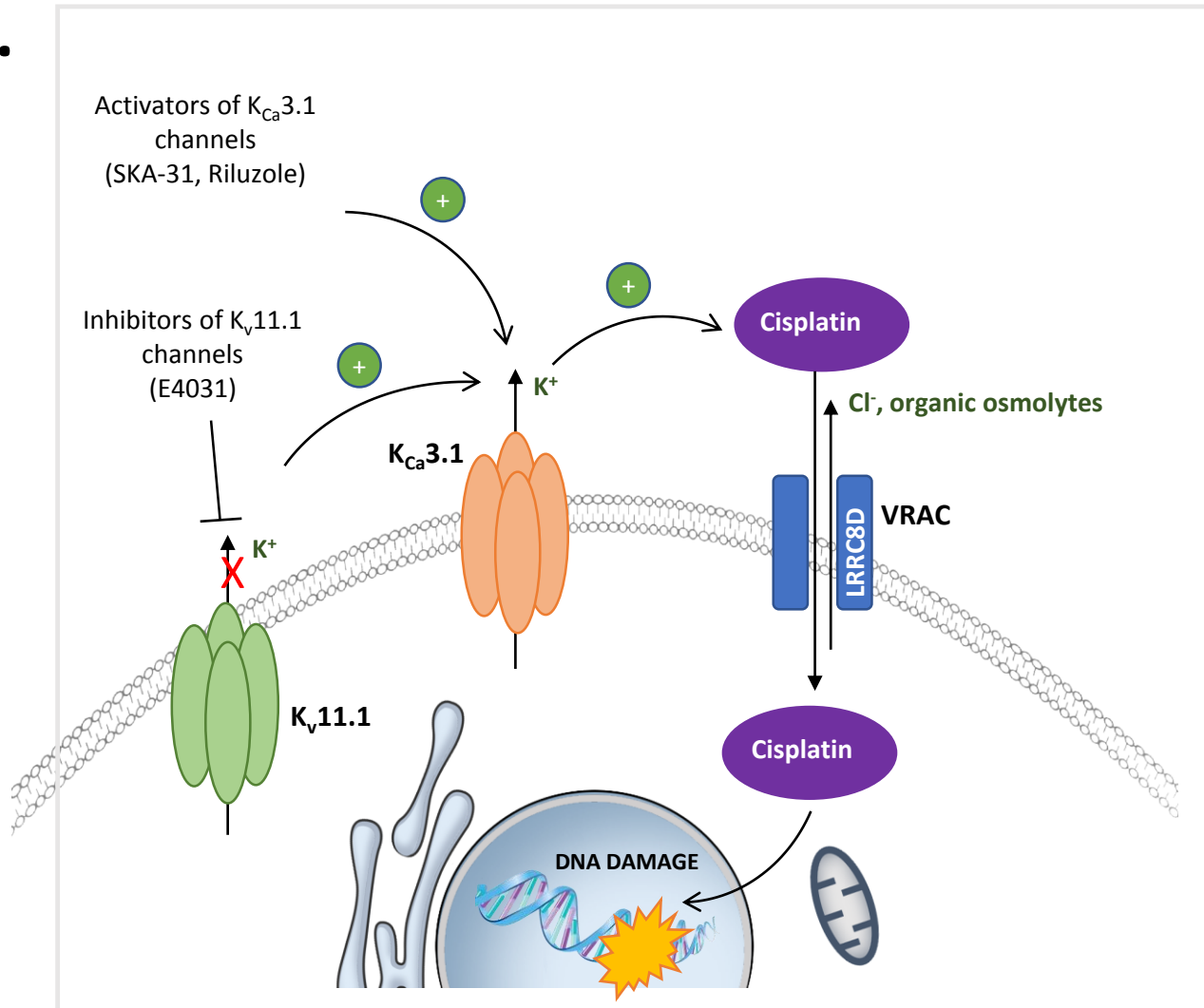
**Table 2. Summary of the effects of K<sup>+</sup> channel modulators (Riluzole, SKA-31, TRAM-34 and E4031) on different biological processes of HCT-116 cells**

| Drug     | V <sub>REST</sub> | Cisplatin       |                |           |                       |
|----------|-------------------|-----------------|----------------|-----------|-----------------------|
|          |                   | Platinum uptake | Cell viability | Apoptosis | Cell cycle            |
| Riluzole | Hyperpolarisation | ↑               | ↓ (S)          | ↑↑        | ↑↑ % of cells in G2/M |
| SKA-31   | Hyperpolarisation | ↑               | ↓ (S)          | ↑↑        | ↑ % of cells in G2/M  |
| TRAM-34  | Depolarisation    | ↓               | (A)            | ↑↑        | ↑↑ % of cells in G2/M |
| E4031    | Depolarisation    | ↑               | ↓ (S)          | ↑↑        | ↑ % of cells in G2/M  |

Abbreviations: ↓ = decrease, ↑ = increase, ↑↑ = strong increase, (A) = antagonism, (S) = synergy. V<sub>REST</sub> was determined in cells treated with the single K<sup>+</sup> channel modulators alone; Platinum uptake, cell viability, apoptosis and cell cycle data are relative to treatments in combination with Cisplatin (25 μM). Experimental data and concentrations used are from Table 1B, Figures 2–4 and Supplementary Table S7.

The activation of  $K_{Ca}3.1$  modulates the VRAC-dependent uptake of Cisplatin (Jentsch et al, 2016).

Blocking hERG1 increases the uptake of Cisplatin, which relies on the activity of  $K_{Ca}3.1$  channels.

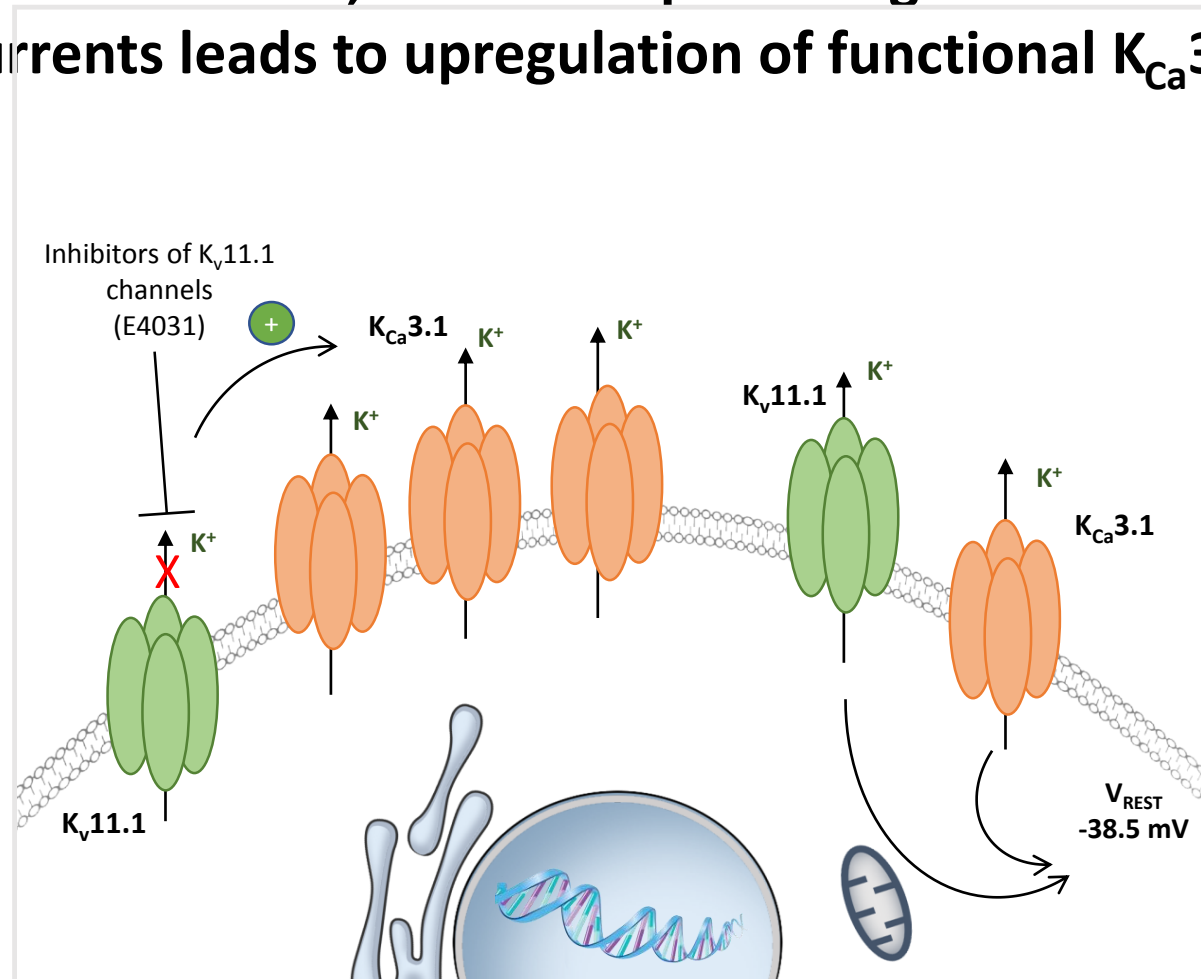




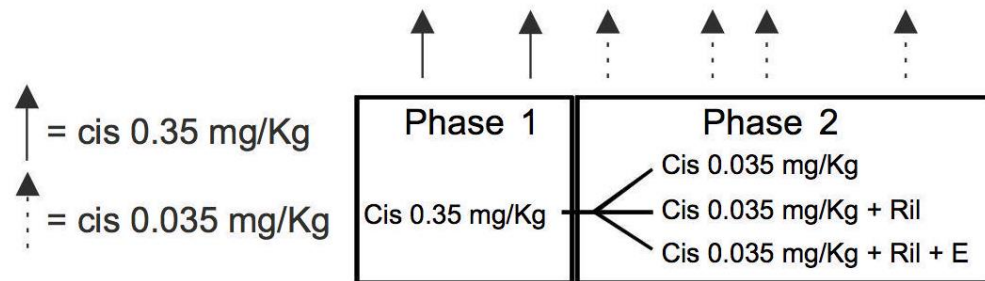
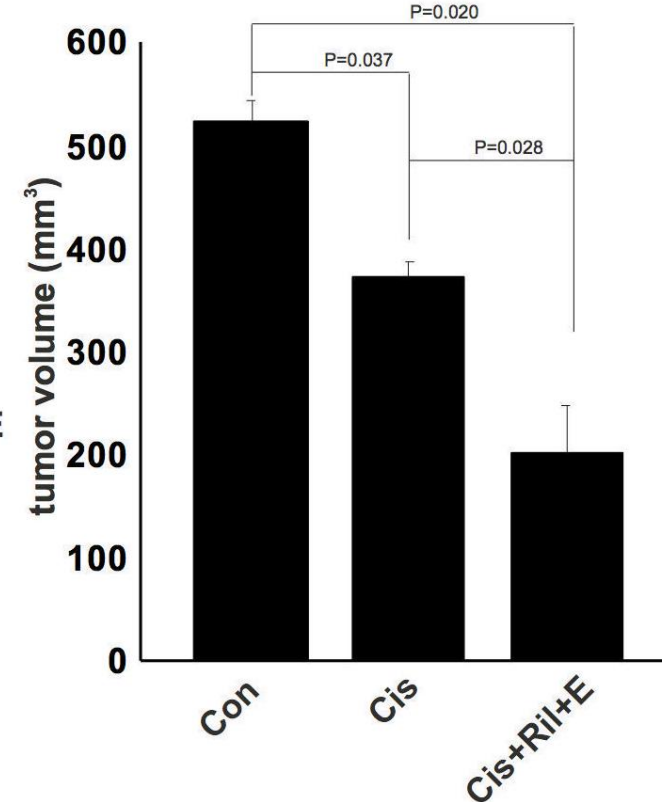
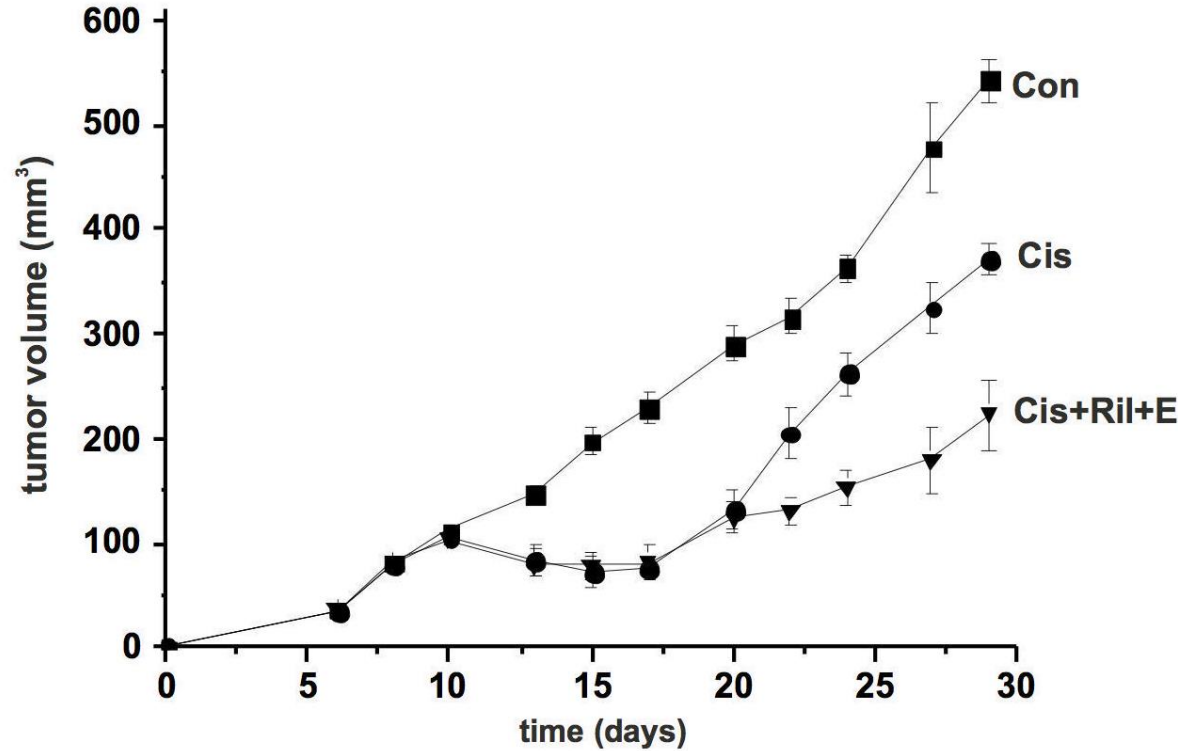
Cisplatin-resistant cells exhibit higher functional expression of  $K_{Ca}3.1$  and hERG1 channels, compared with Cisplatin-sensitive cells.

The two channels are functionally related in these cells:

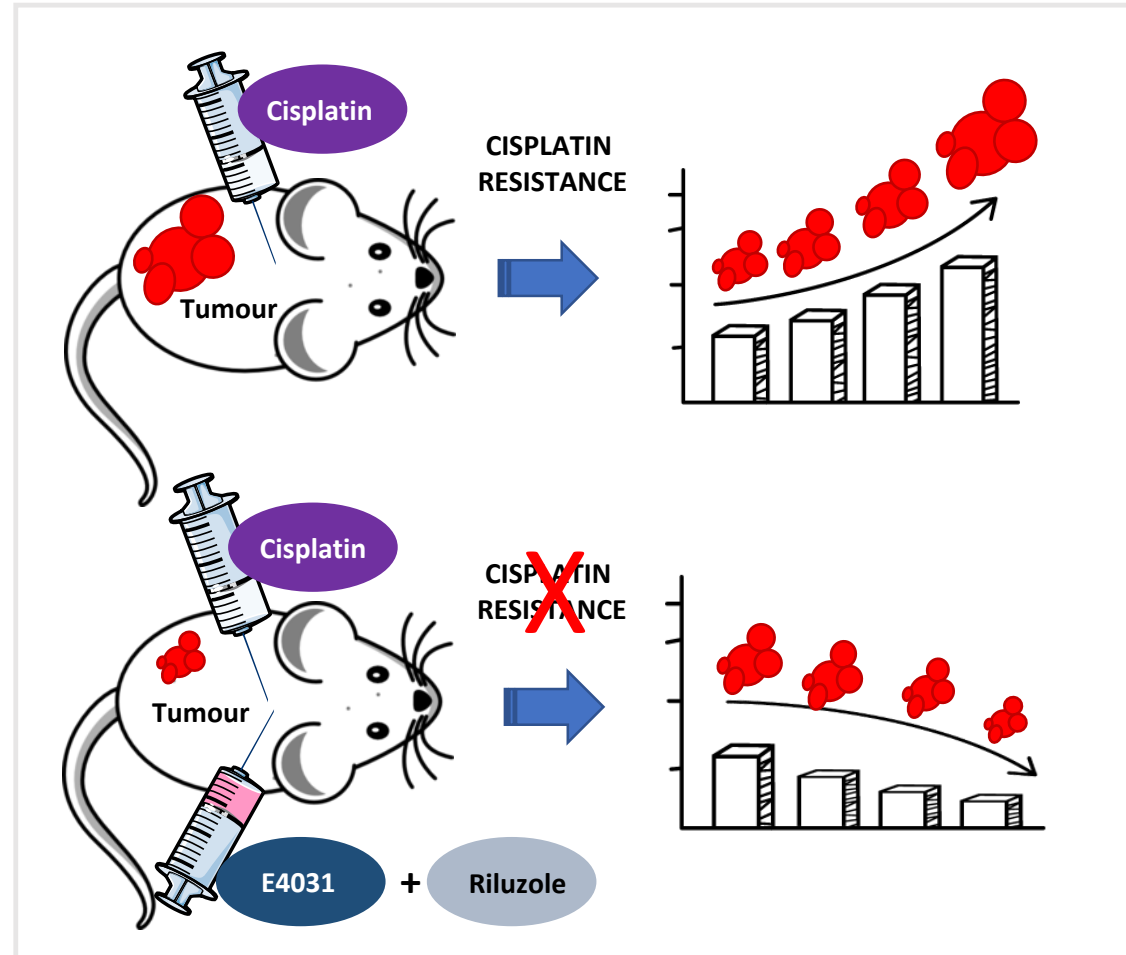
- (1) they set  $V_{REST}$  to more hyperpolarised values;
- (2) their expression is coordinated, one compensating for the other: prolonged (24h) inhibition of hERG1 currents leads to upregulation of functional  $K_{Ca}3.1$  channels.



The concomitant activation of  $K_{Ca}3.1$  and inhibition of hERG1 potentiates the pro-apoptotic activity of Cisplatin, *in vivo*, hence contributing to overcome Cisplatin resistance.



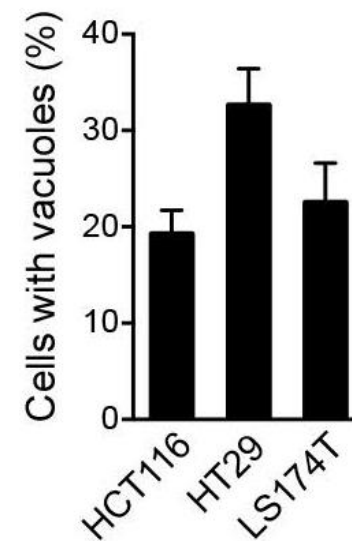
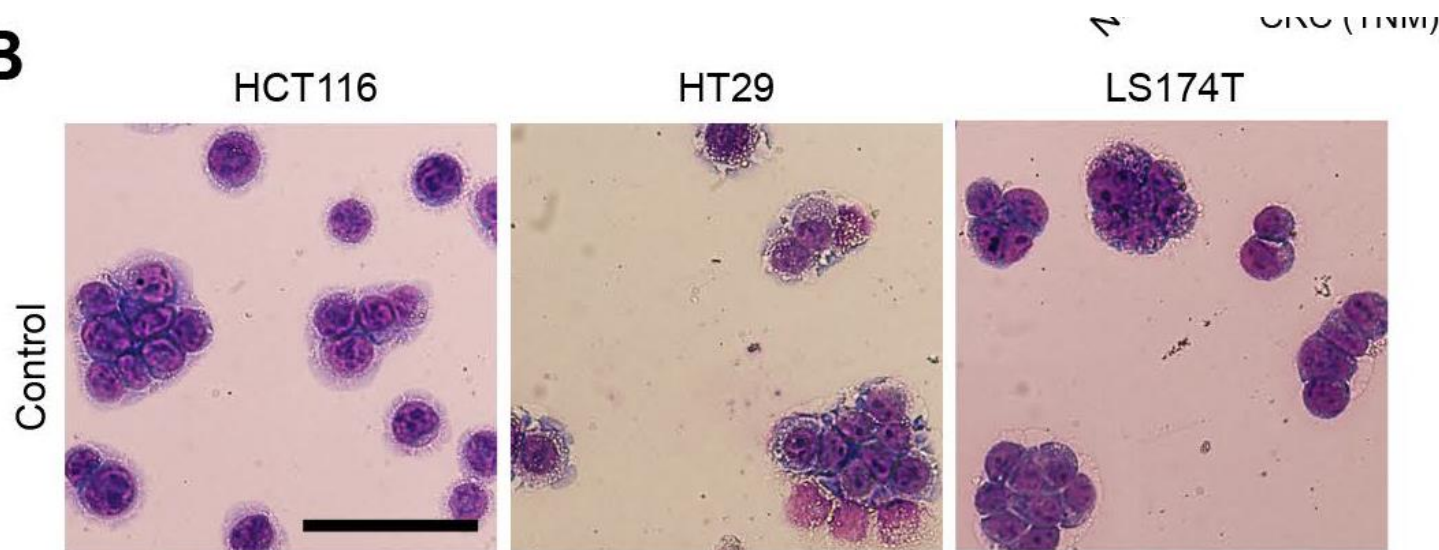
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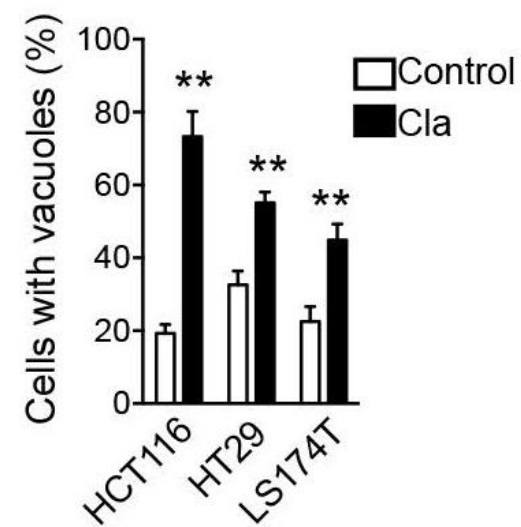
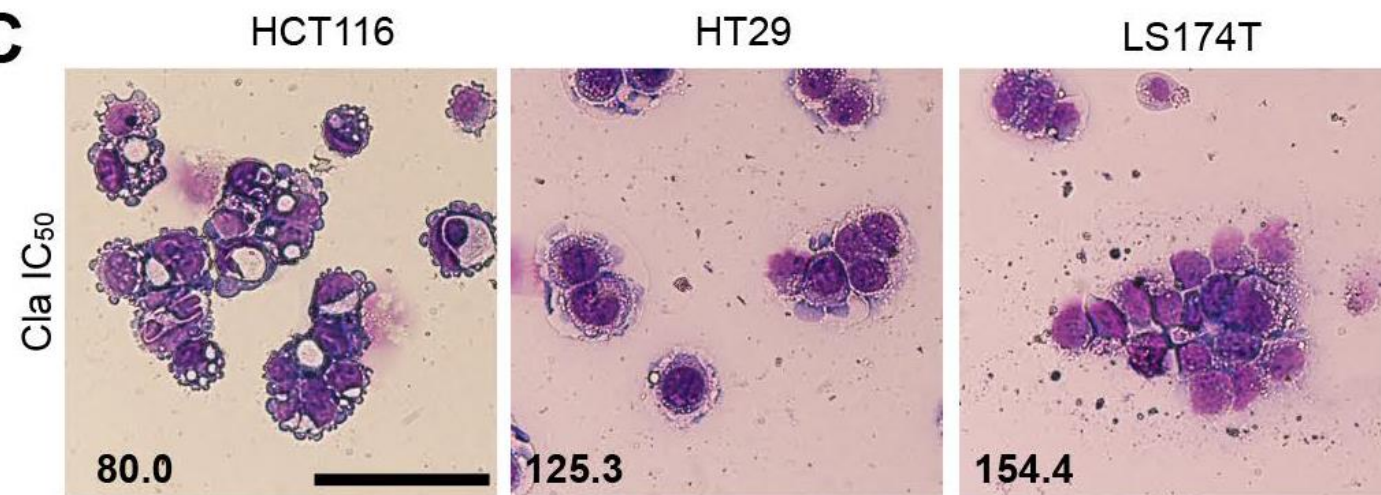
**(2) Effects of Clarithromycin on hERG1:  
sinergy with 5-FU**  
(unpublished)

# Clarithromycin triggers autophagy in CRC cells

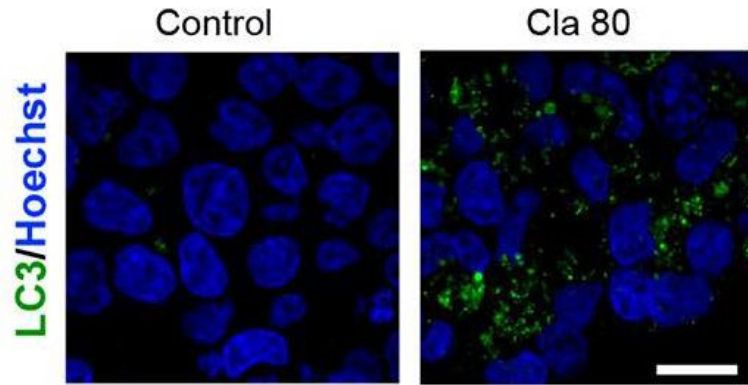
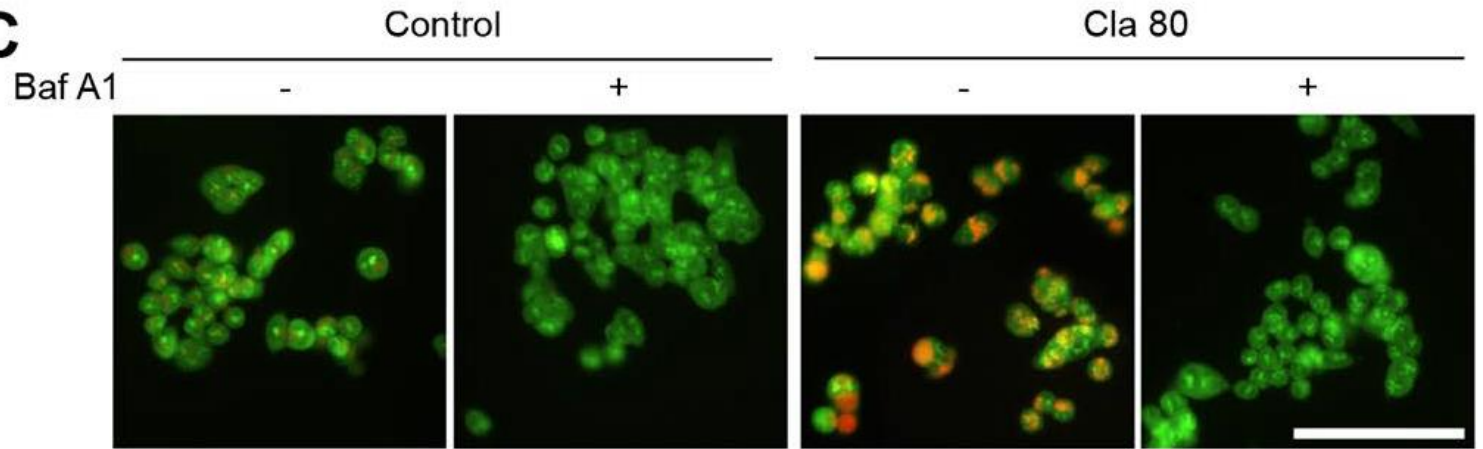
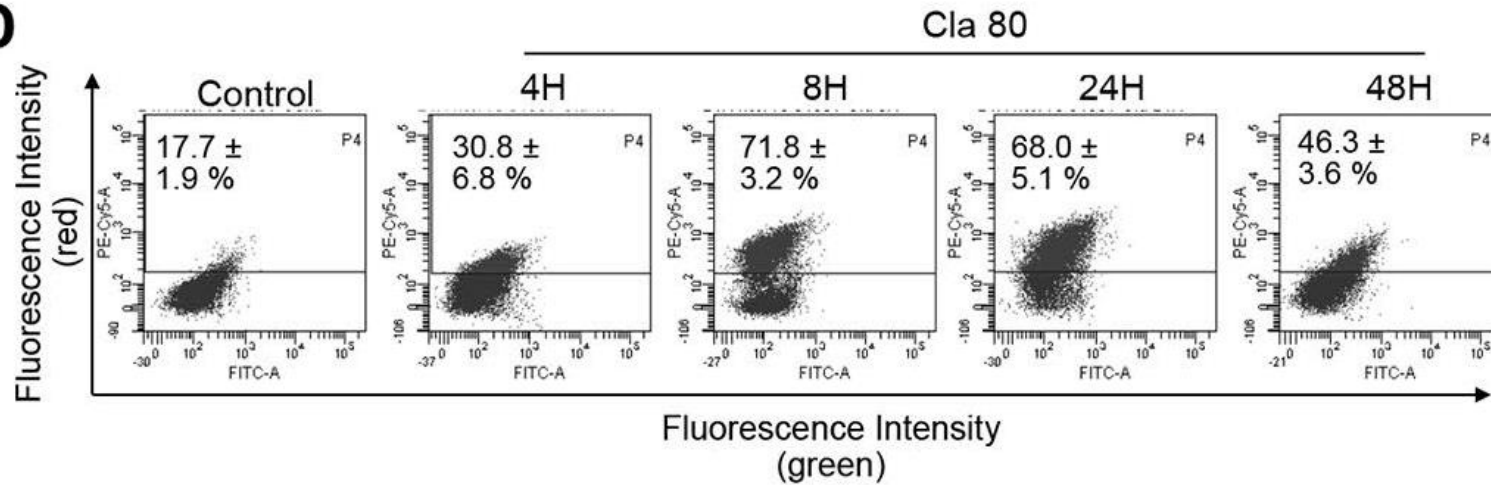
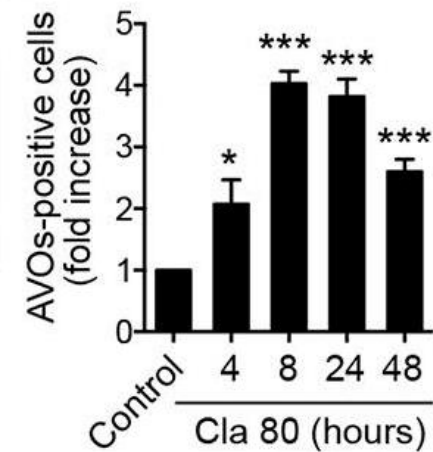
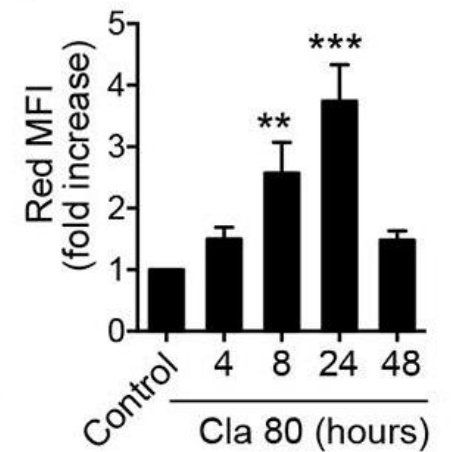
**B**



**C**

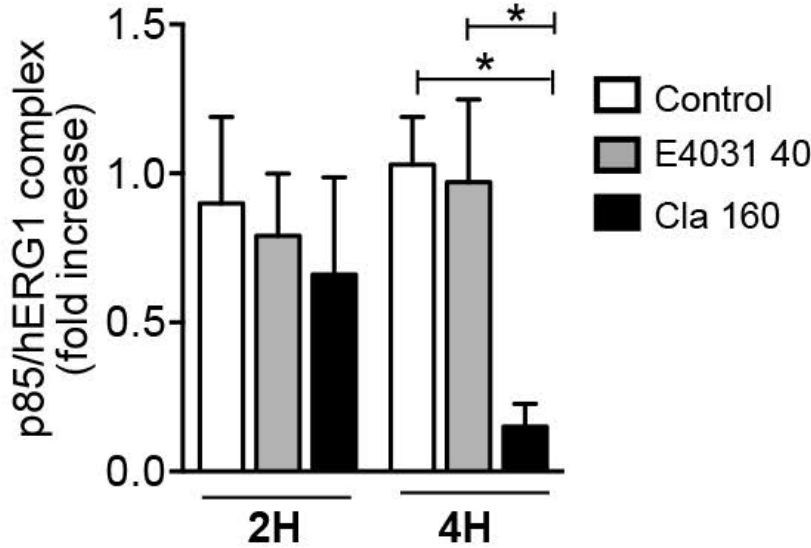
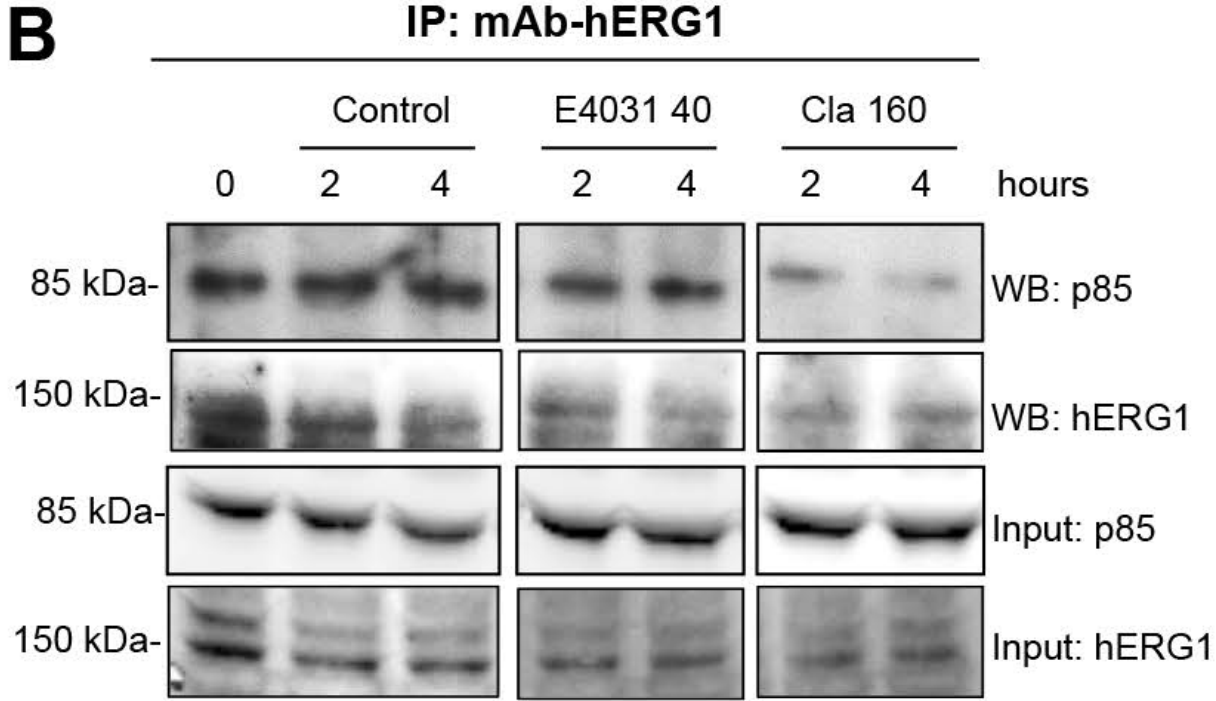


# Clarithromycin triggers autophagy in CRC cells

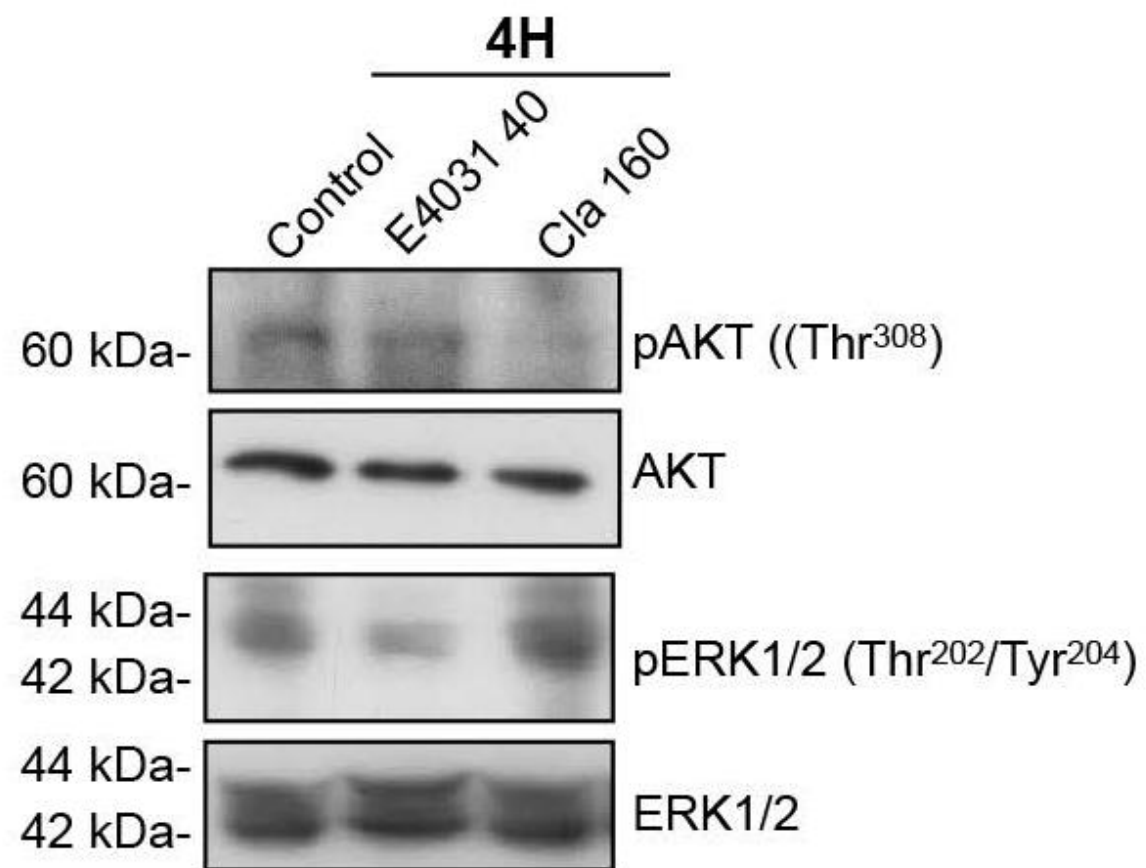
**B****C****D****E****F**

# Chlarithromycin dissociates the hERG1/PI3k (p85 subunit) complex

**B**

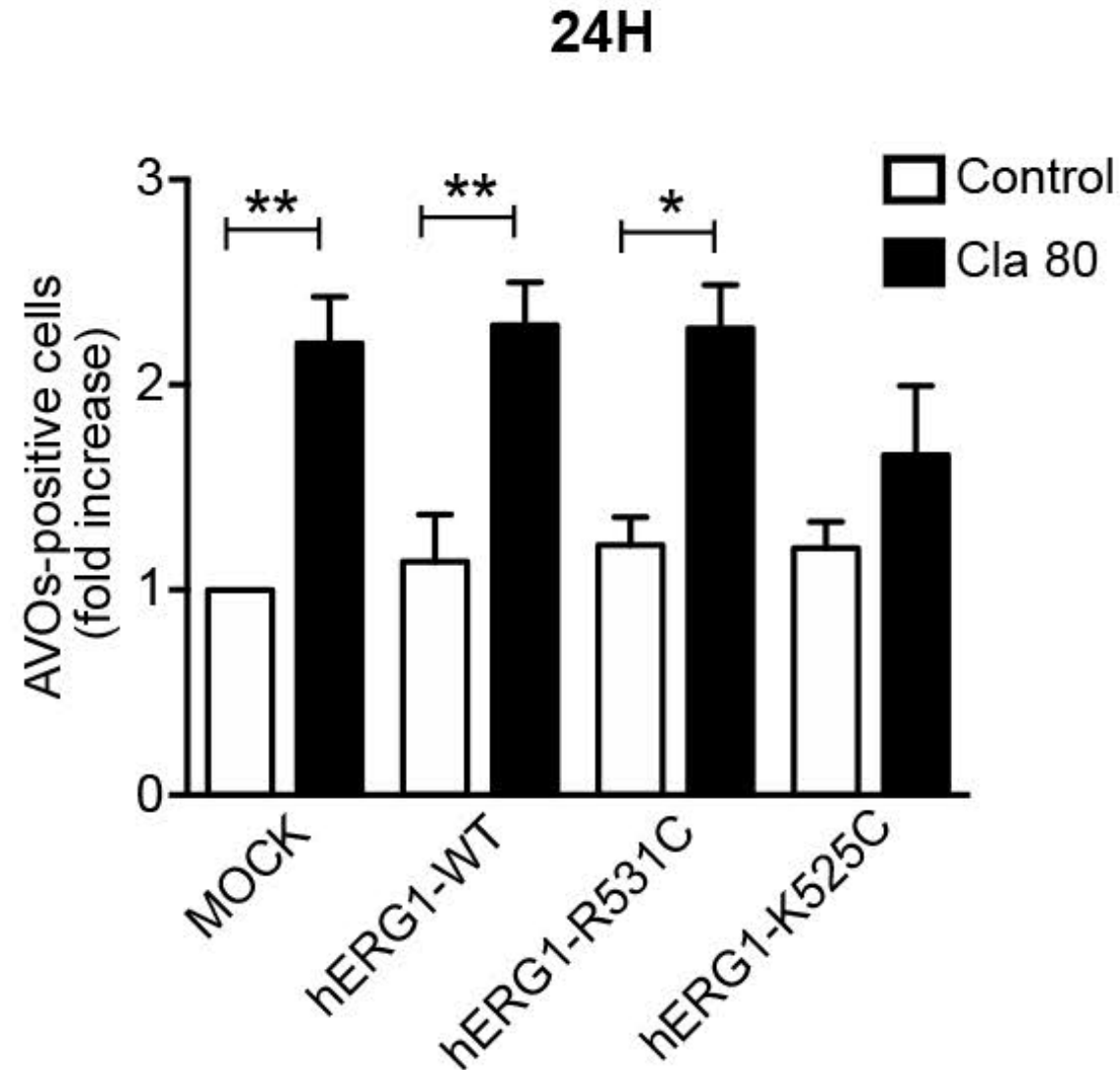


# Different effects of E4031 and Clarithromycin on Akt phosphorylation

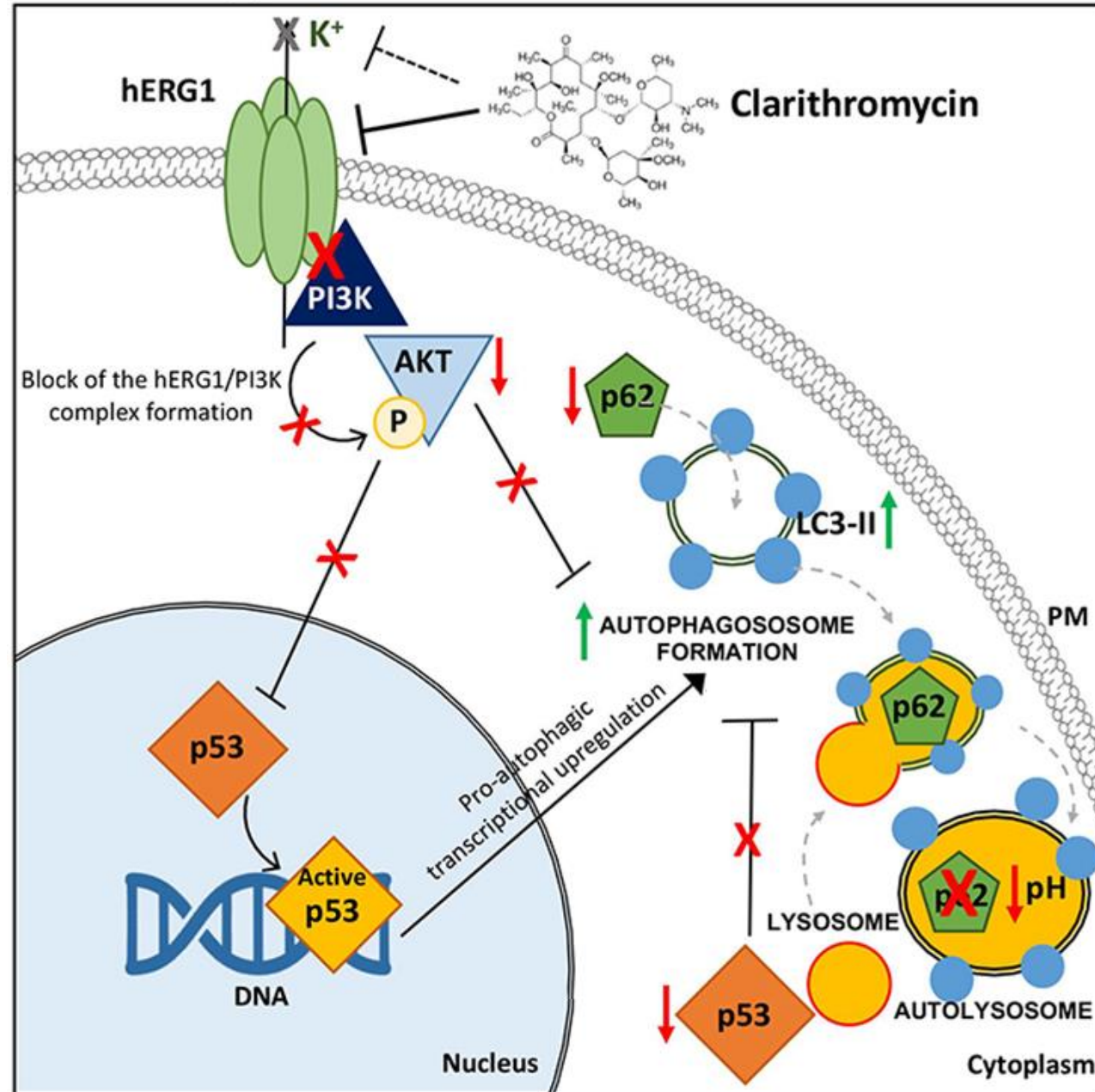




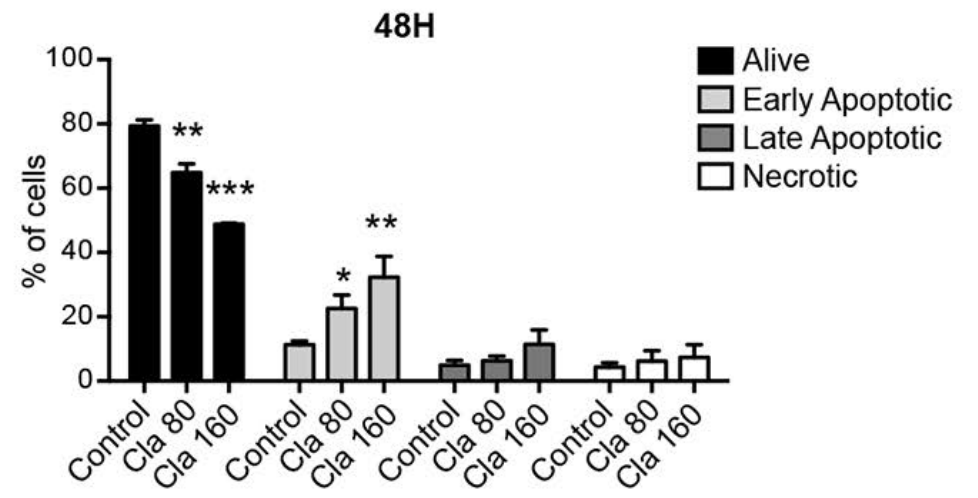
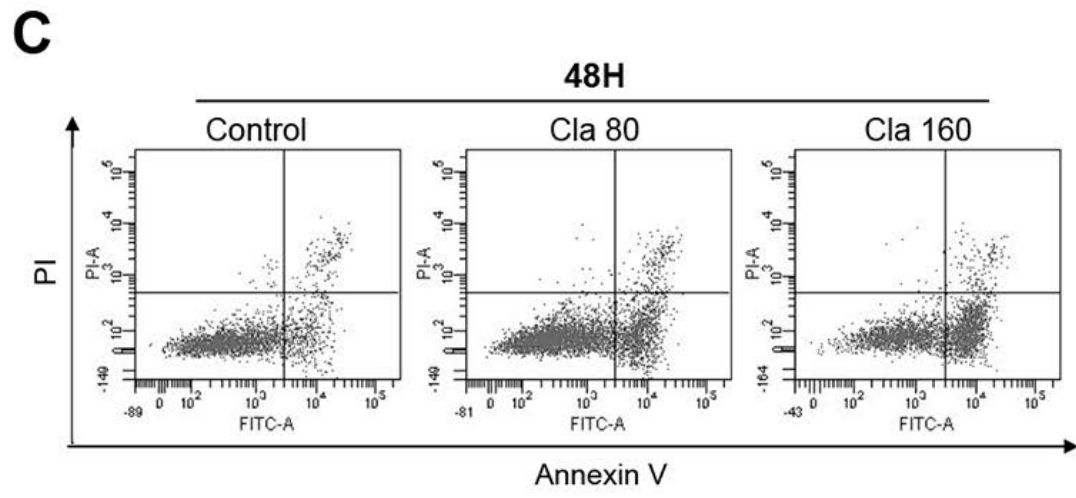
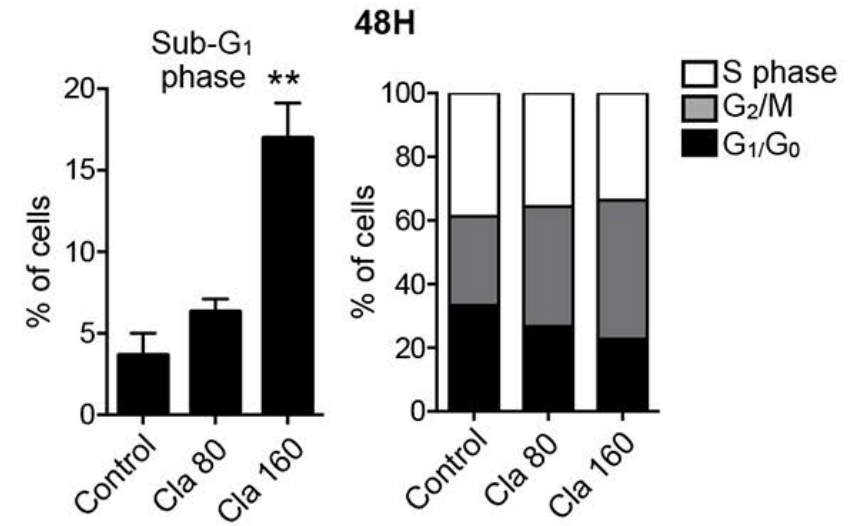
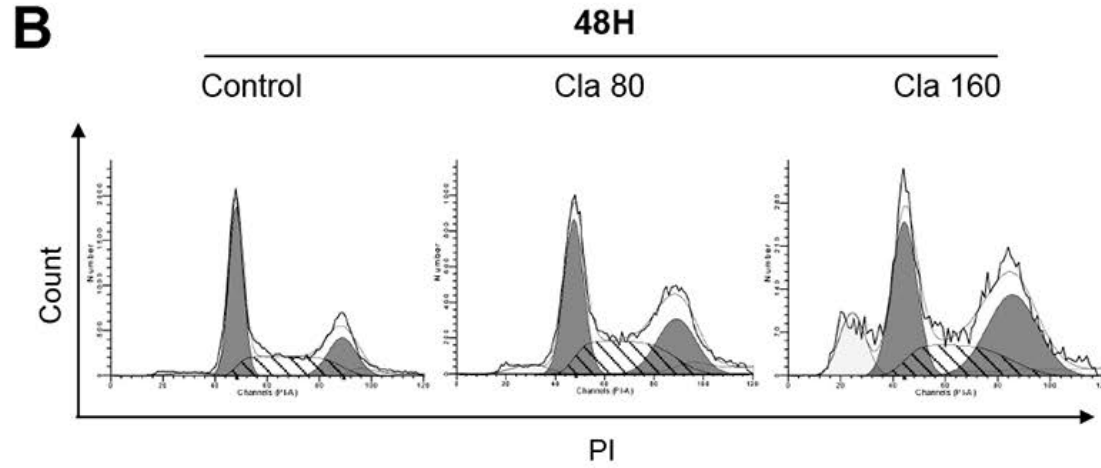
# Clarithromycin effects depend on the conformational state of hERG1 channels , occurring when the channel is preferentially in the closed state



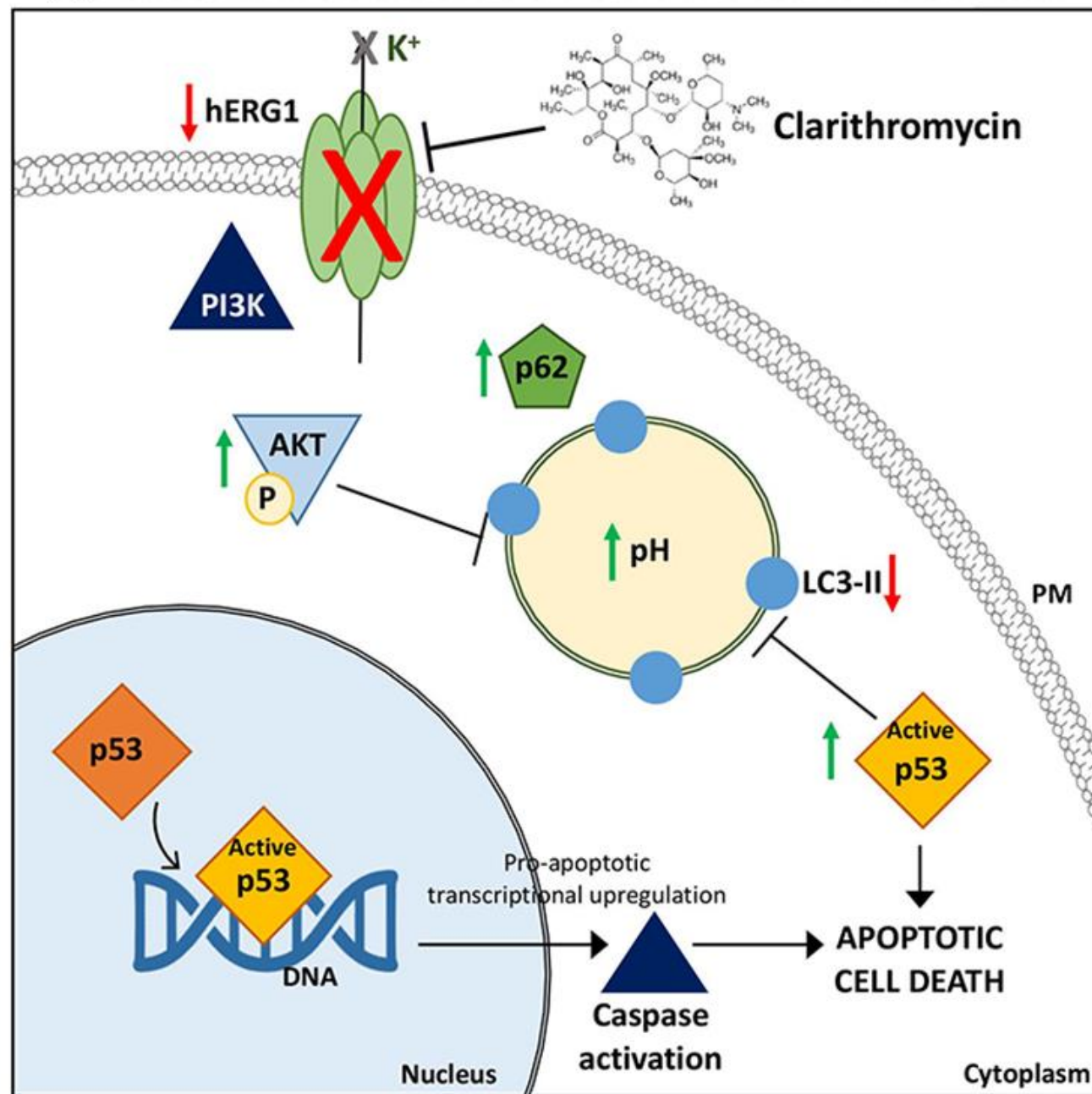
## (B) ACCUMULATION OF AUTOLYSOSOMES



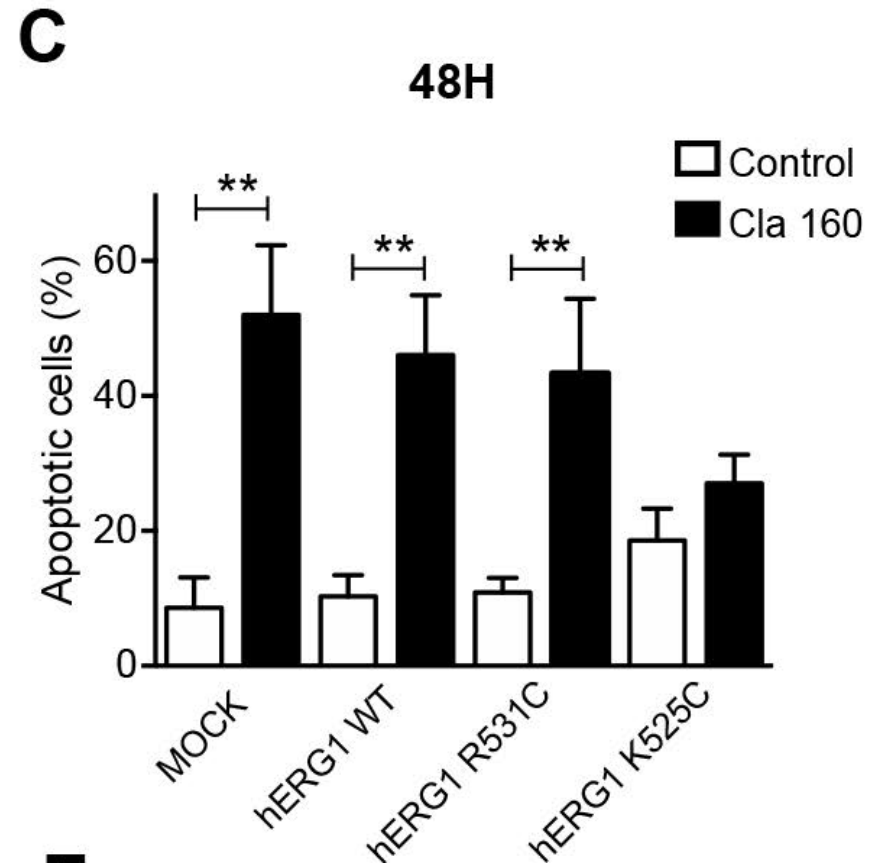
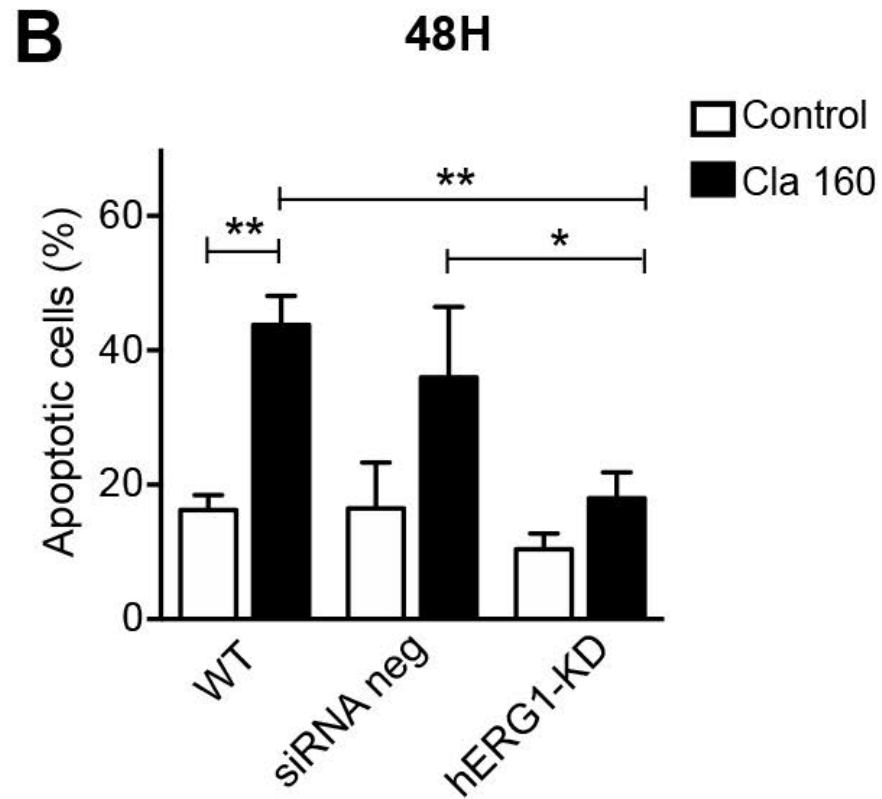
# Clarithromycin triggers apoptotic cell death



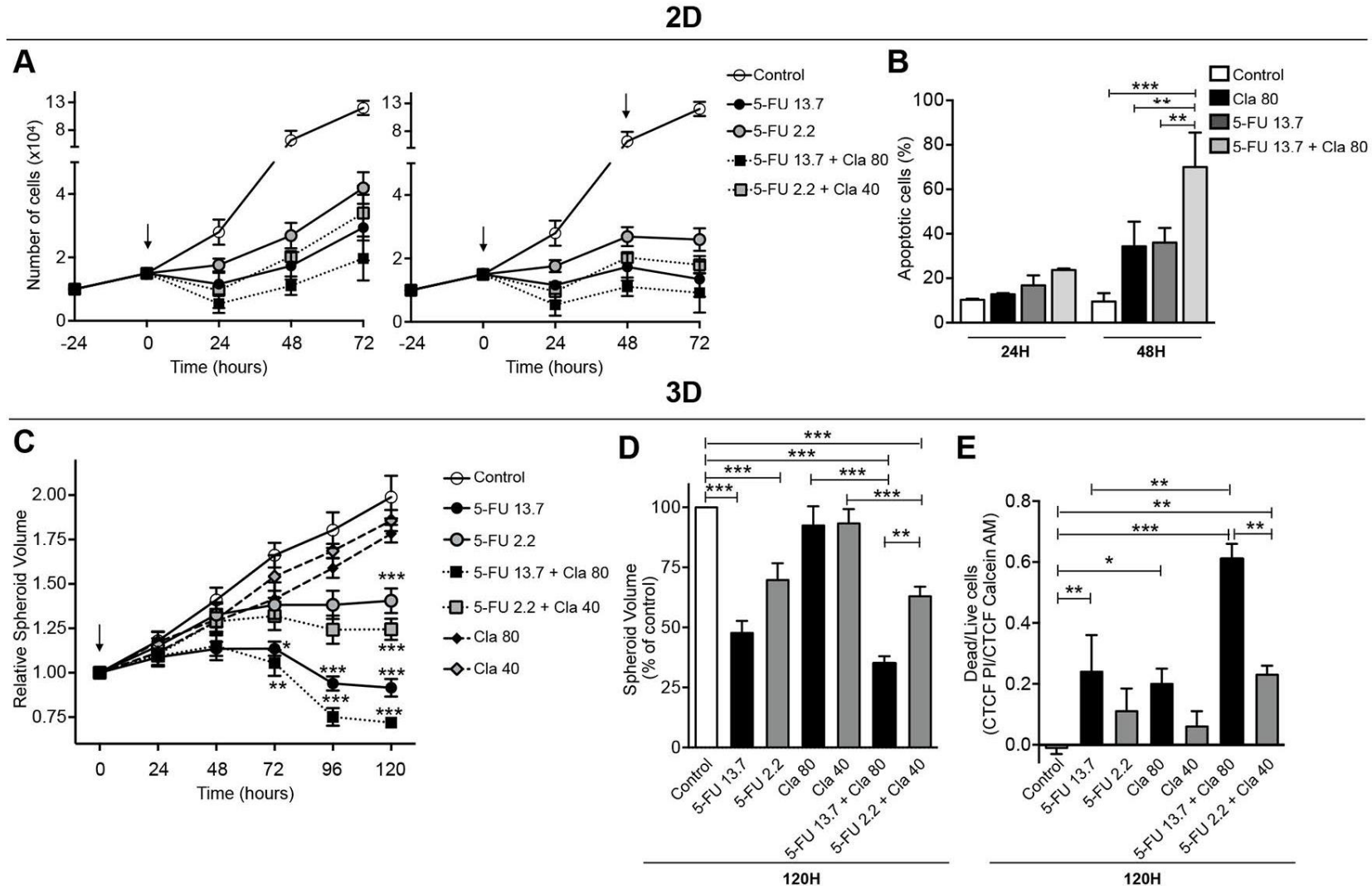
(C) EXHAUSTION OF AUTOPHAGY AND APOPTOTIC CELL DEATH



# The hERG1 conformational state determines (the closed state favours) the pro-apoptotic effects of Clarithromycin



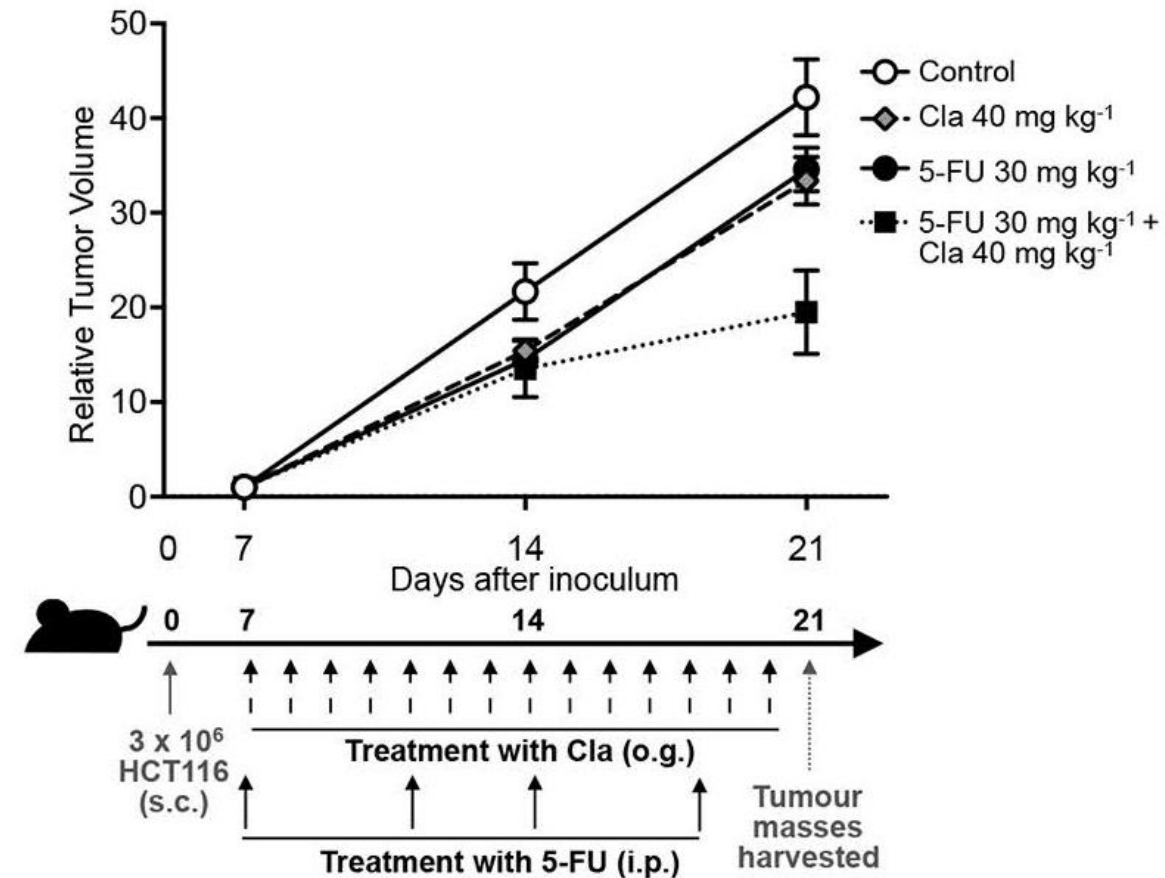
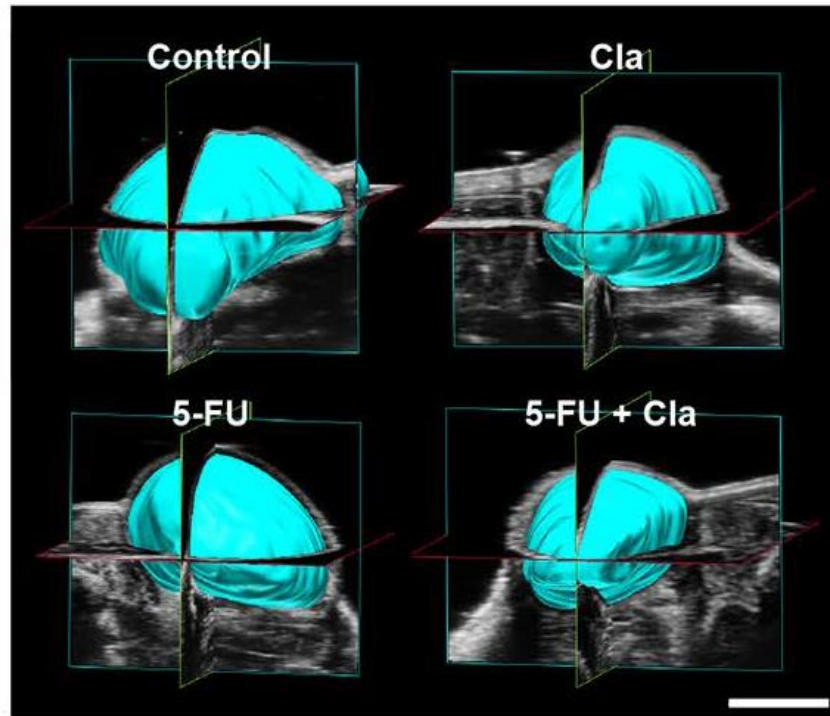
# Clarithromycin has a synergic, anti-proliferative, effect with 5-FU

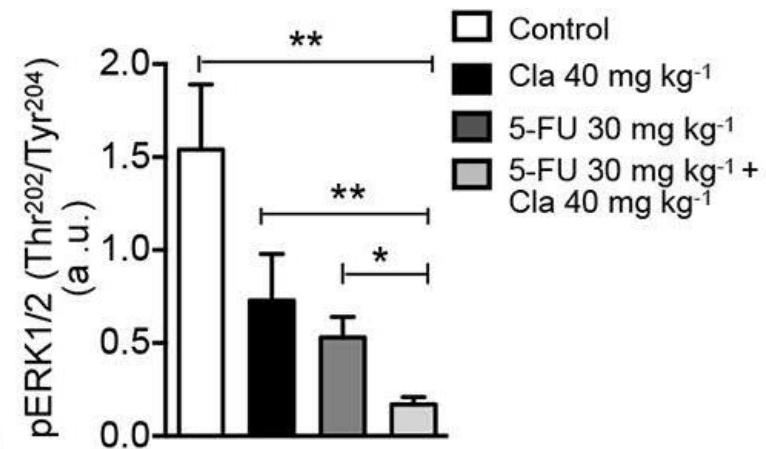
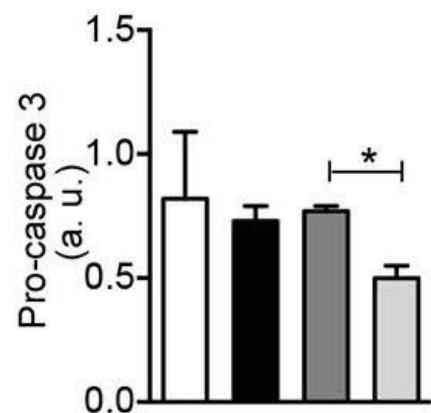
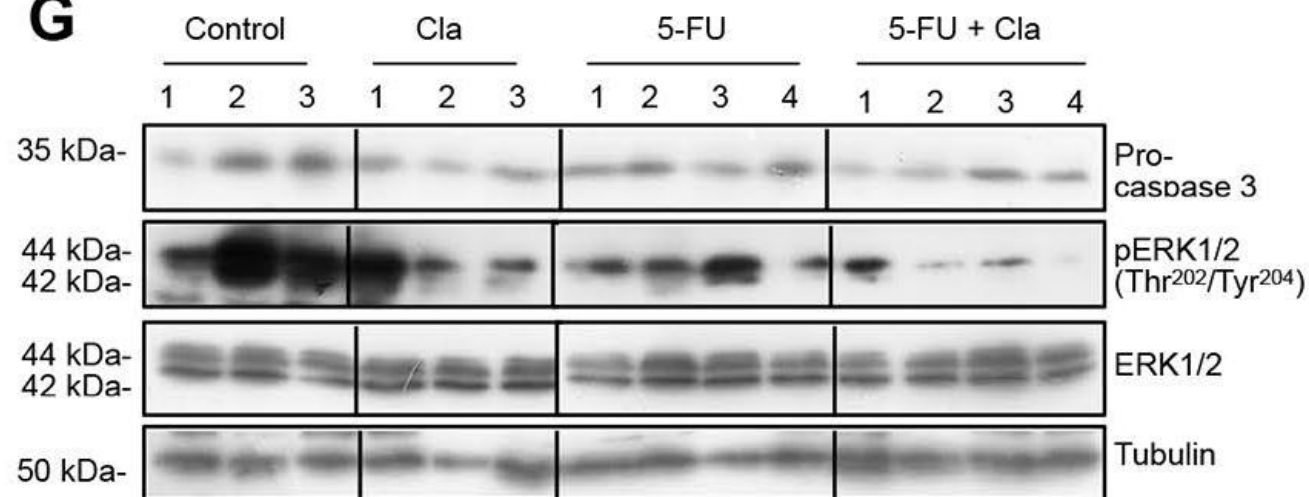
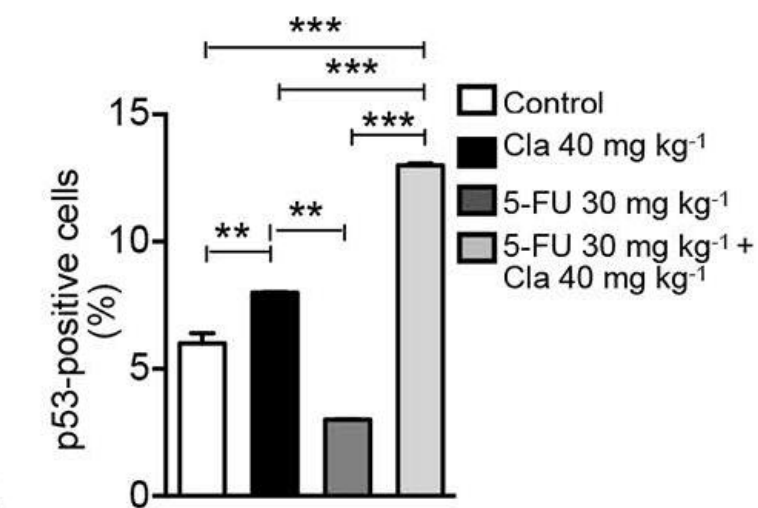
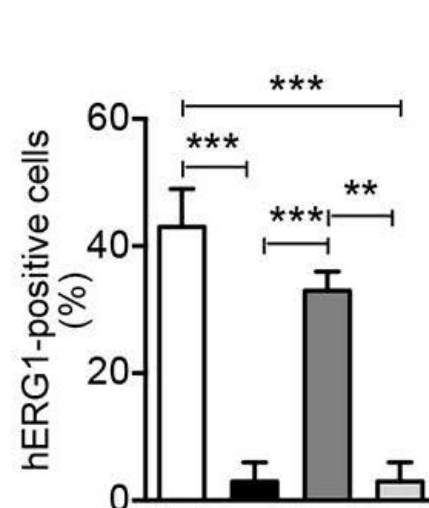
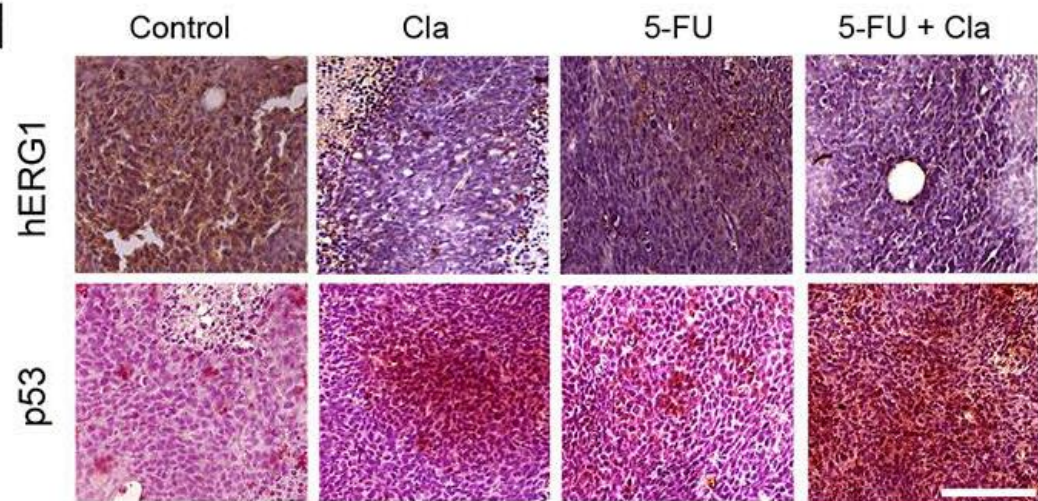


# ...overcoming 5-FU resistance in vivo, in a preclinical xenograft mouse model

in vivo

F



**G****H**



# The hERG1/ $\beta$ 1 integrin complex in CRC

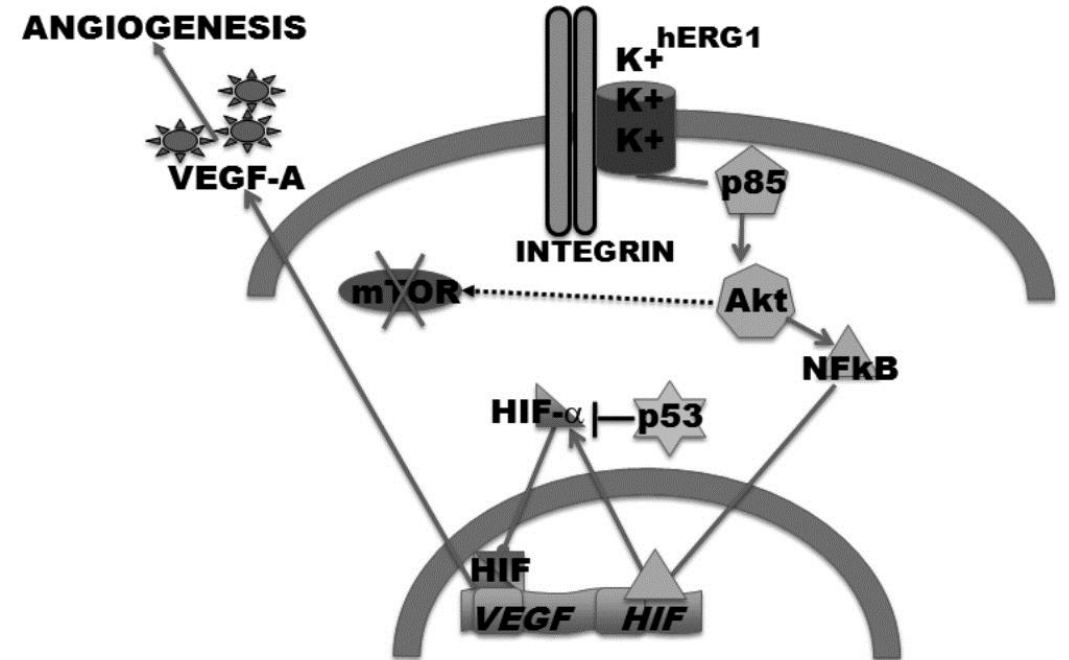


OPEN

hERG1 channels modulate integrin signaling to trigger angiogenesis and tumor progression in colorectal cancer

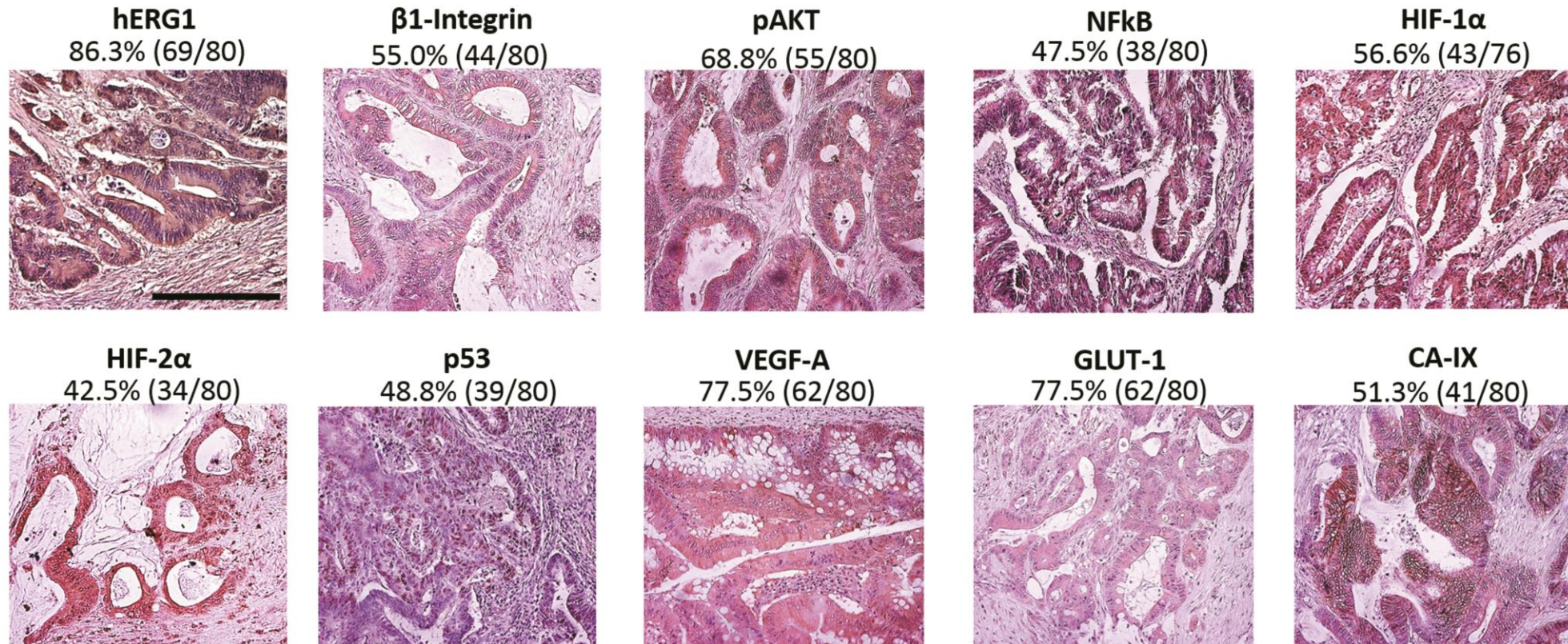
SUBJECT AREAS:  
COLORECTAL CANCER  
INTEGRIN SIGNALLING  
TUMOUR ANGIOGENESIS  
ION CHANNEL SIGNALLING

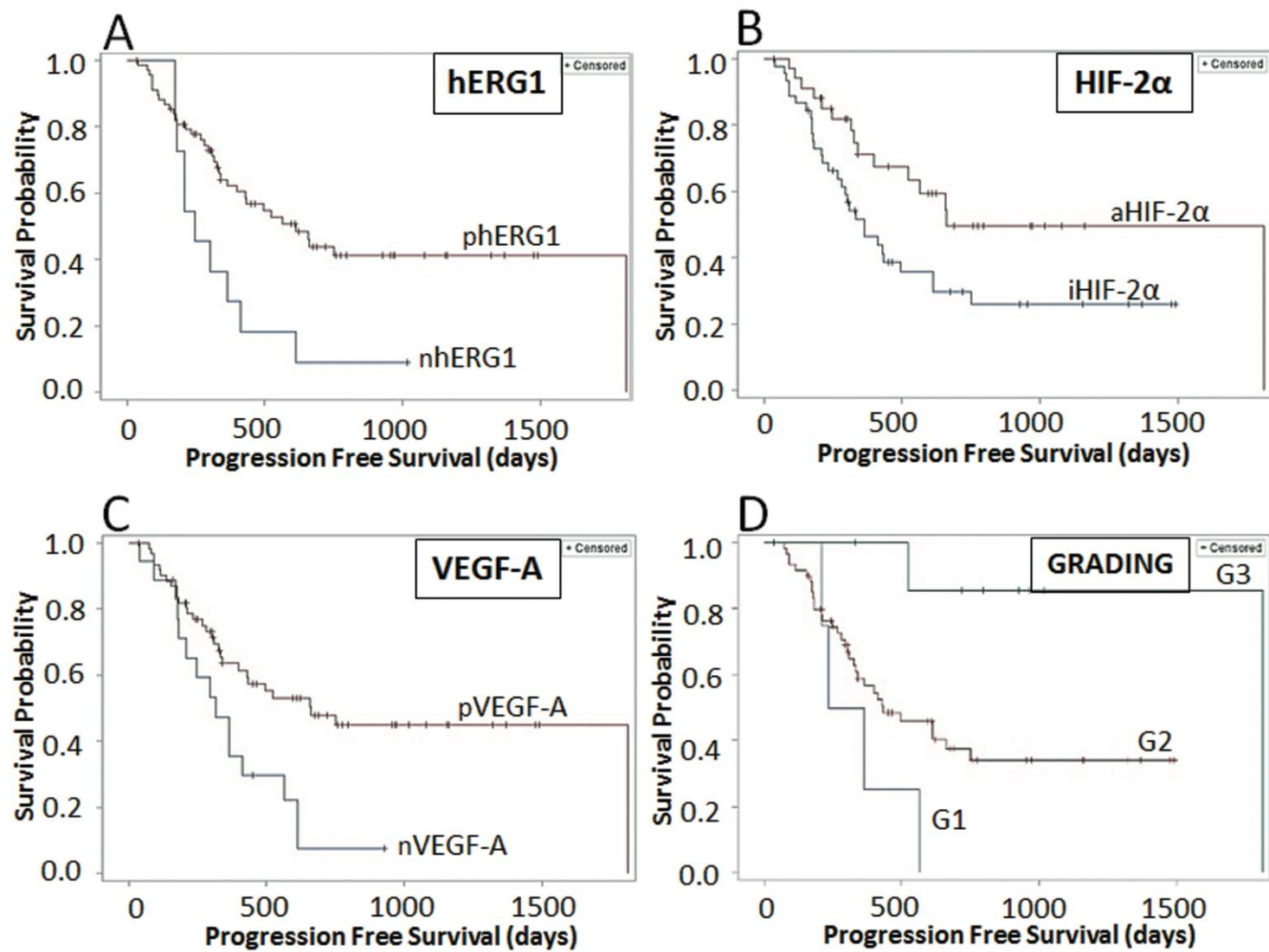
Olivia Crociani<sup>1</sup>, Francesca Zanieri<sup>1</sup>, Serena Pillozzi<sup>1</sup>, Elena Lastraioi<sup>1</sup>, Matteo Stefanini<sup>1</sup>, Antonella Fiore<sup>1</sup>, Angelo Fortunato<sup>1</sup>, Massimo D'Amico<sup>1</sup>, Marika Masselli<sup>1</sup>, Emanuele De Lorenzo<sup>1</sup>, Luca Gasparoli<sup>1</sup>, Martina Chiu<sup>2</sup>, Ovidio Bussolati<sup>2</sup>, Andrea Becchetti<sup>2</sup> & Annarosa Arcangeli<sup>1</sup>



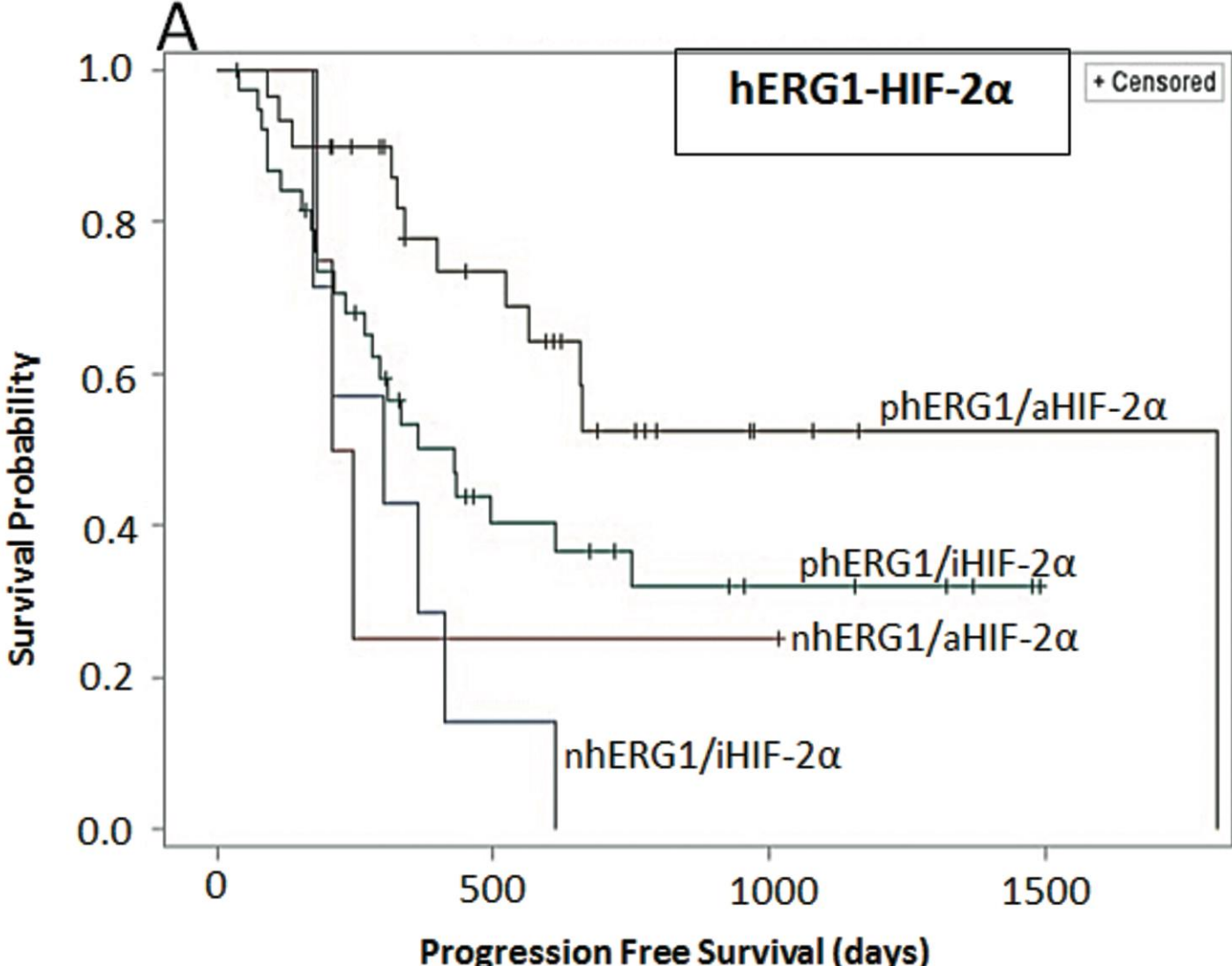
# Expression of hERG1 and several angiogenesis-related proteins in metastatic CRC samples from patients treated in first line with Bevacizumab

**FIGURE 1**





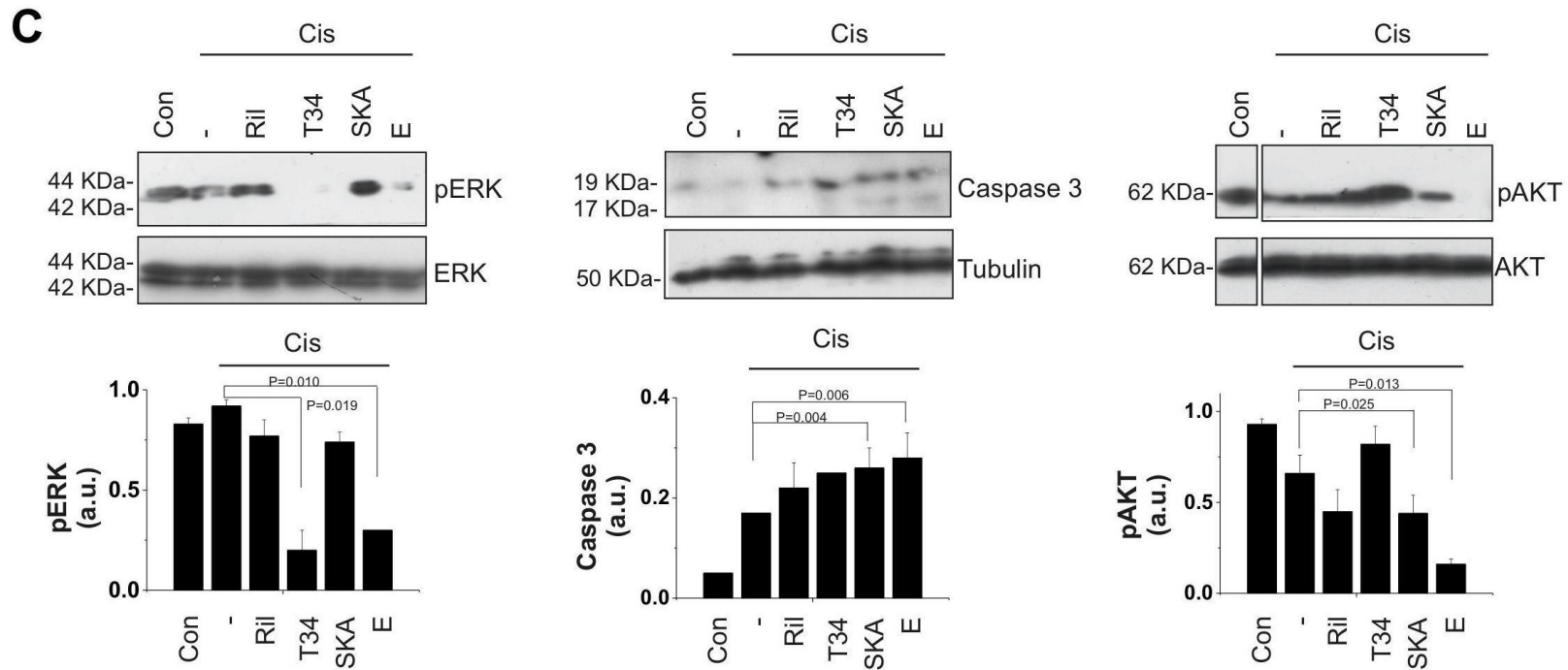
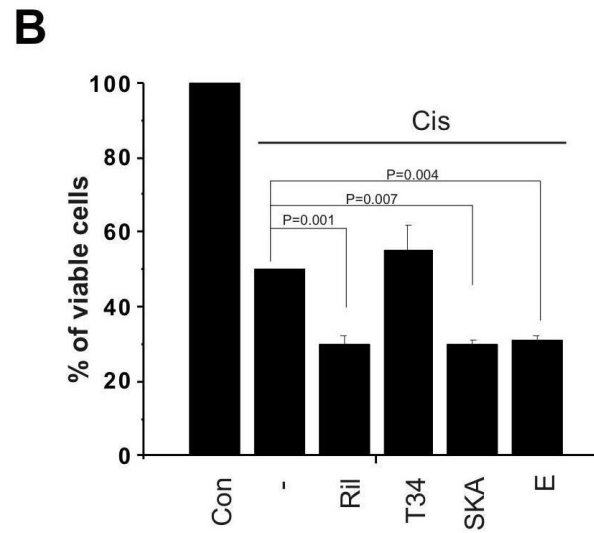
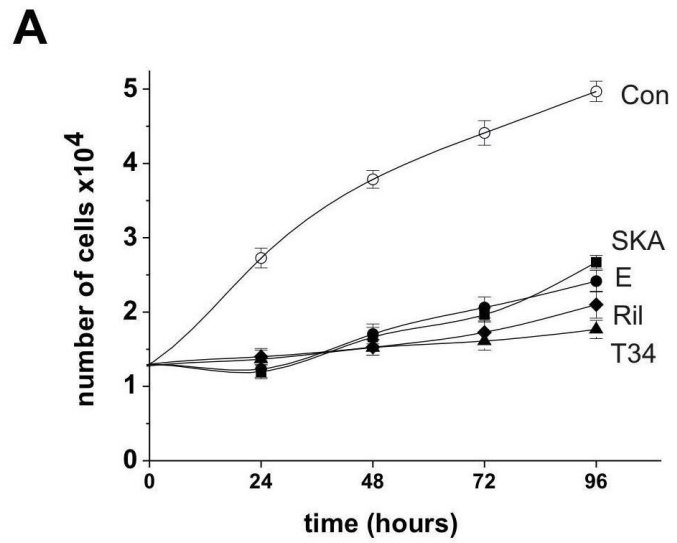
# hERG1 and HIF-2 $\alpha$ expression have a significant positive impact on PFS



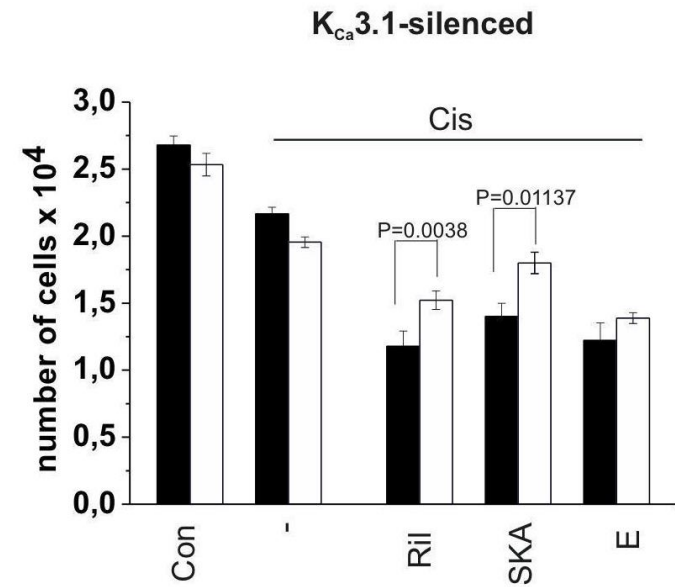
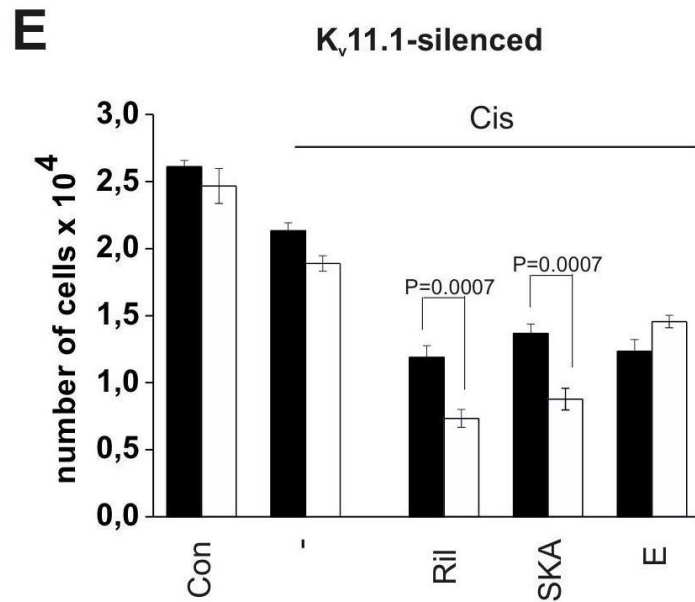
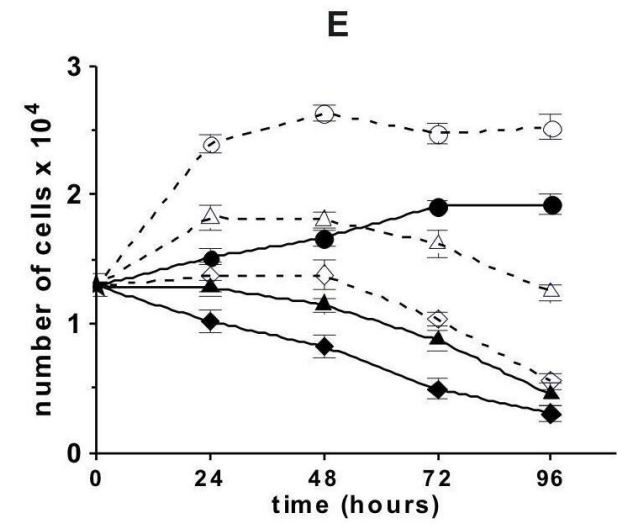
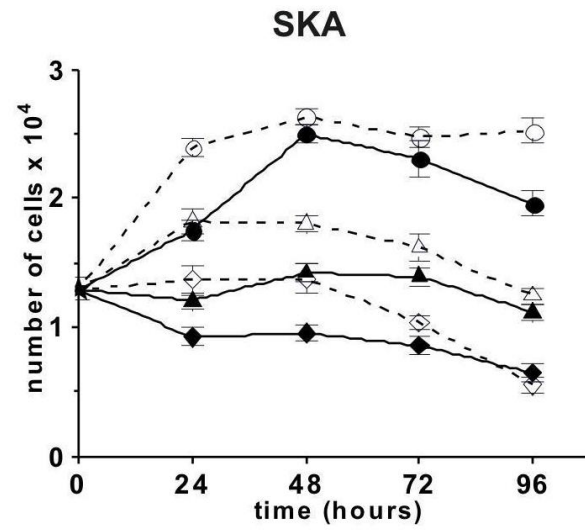
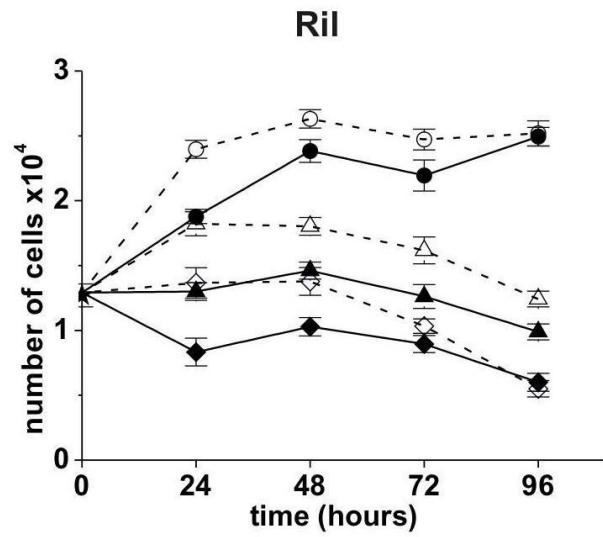
**Table 1A. IC<sub>50</sub> values and effects on apoptosis and cell cycle distribution of Cisplatin, Riluzole, SKA-31, TRAM-34 and E4031 in HCT-116 and HCT-8 cells**

|                | IC <sub>50</sub> (μM) | Concentration of the drug (μM) | Apoptosis              |                        | Cell cycle            |                        |                        |
|----------------|-----------------------|--------------------------------|------------------------|------------------------|-----------------------|------------------------|------------------------|
|                |                       |                                | Early apoptosis (%)    | Late apoptosis (%)     | G0/G1 (%)             | S (%)                  | G2/M (%)               |
| <b>HCT-116</b> |                       |                                |                        |                        |                       |                        |                        |
| Control        | —                     |                                | 1.0 ± 0.6              | 0.8 ± 0.5              | 27.7 ± 7.7            | 55.5 ± 3.7             | 16.9 ± 6.4             |
| Cisplatin      | 25.2 ± 2.1            | 25                             | 5.9 ± 1.0<br>P= 0.001  | 5.3 ± 1.5<br>P= 0.009  | 48.0 ± 4.5<br>P=0.009 | 39.2 ± 2.5<br>P= 0.018 | 12.8 ± 3.3             |
| Riluzole       | 9.5 ± 1.0             | 10                             | 13.6 ± 3.7<br>P= 0.004 | 10.3 ± 4.1<br>P= 0.028 | 51.6 ± 6.0<br>P=0.004 | 26.1 ± 8.8<br>P= 0.021 | 22.3 ± 3.1             |
| SKA-31         | 5.3 ± 0.3             | 5                              | 7.0 ± 0.9<br>P= 0.000  | 3.5 ± 0.9<br>P= 0.015  | 55.3 ± 2.4<br>P=0.006 | 31.4 ± 7.7<br>P= 0.028 | 13.3 ± 5.7             |
| TRAM-34        | 24.4 ± 1.8            | 25                             | 9.6 ± 2.1<br>P= 0.001  | 6.6 ± 0.9<br>P= 0.000  | 50.5 ± 3.2            | 34.2 ± 5.0             | 15.4 ± 7.6             |
| E4031          | 6.6 ± 1.6             | 7                              | 5.2 ± 1.2<br>P= 0.005  | 4.1 ± 1.0<br>P= 0.010  | 51.4 ± 4.3<br>P=0.010 | 26.4 ± 4.1<br>P= 0.012 | 22.3 ± 6.1             |
| <b>HCT-8</b>   |                       |                                |                        |                        |                       |                        |                        |
| Control        | —                     |                                | 0.8 ± 0.3              | 1.2 ± 0.3              | 30.7 ± 2.4            | 53.8 ± 3.3             | 15.5 ± 1.9             |
| Cisplatin      | 8.7 ± 1.4             | 9                              | 5.5 ± 1.4<br>P= 0.008  | 13.4 ± 7.2<br>P= 0.018 | 46.9 ± 1.3<br>P=0.019 | 43.4 ± 1.9<br>P= 0.002 | 9.7 ± 3.1<br>P= 0.041  |
| Riluzole       | 12.9 ± 0.7            | 13                             | 3.6 ± 0.9<br>P= 0.008  | 4.0 ± 1.2<br>P= 0.035  | 8.5 ± 3.4<br>P=0.003  | 6.7 ± 4.3<br>P= 0.000  | 84.6 ± 5.6<br>P= 0.000 |
| SKA-31         | 46.9 ± 1.4            | 45                             | 3.0 ± 0.4<br>P= 0.001  | 10.2 ± 5.0<br>P= 0.011 | 47.5 ± 3.4<br>P=0.021 | 39.9 ± 8.7<br>P= 0.046 | 12.6 ± 10.0            |
| TRAM-34        | 20.1 ± 1.1            | 20                             | 3.2 ± 1.2<br>P= 0.012  | 2.7 ± 1.0<br>P= 0.019  | 62.9 ± 3.2<br>P=0.026 | 28.9 ± 4.5<br>P= 0.026 | 8.2 ± 7.6              |
| E4031          | 13.3 ± 1.3            | 13                             | 2.8 ± 1.9              | 2.8 ± 0.7<br>P= 0.015  | 25.2 ± 0.4<br>P=0.012 | 57.1 ± 2.2             | 17.7 ± 2.4             |

IC<sub>50</sub> values were determined after 24 h of treatment by the Trypan Blue exclusion test, using the Origin Software. Apoptosis and cell cycle distributions were evaluated by treating the cells with the drug concentrations indicated in the third column for 24 h. The percentage of cells in early (Annexin + /PI - cells) and late apoptosis (Annexin + /PI + cells) was determined by Annexin/PI assay as detailed in the Materials and Methods section. Cell cycle distribution was assessed by flow cytometry after staining the cells with propidium iodide (PI) and is indicated as the percentage of cells in the different cell cycle phases. Data are means ± s.e.m. of three independent experiments, each carried out in triplicate. For statistical analysis, Student's t-test was applied.



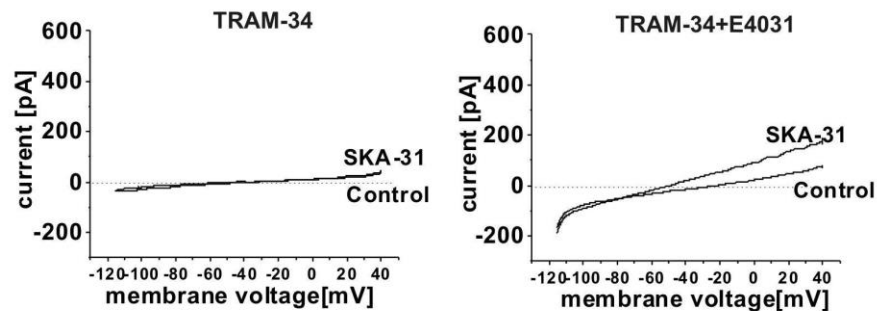
**D**



# hERG1 blockade leads to increased $K_{Ca}3.1$ expression and thereby stimulated Cisplatin uptake.

D

|                | Number of cells with active $K_{Ca}3.1$ current/total cells (%) | Slope fold variation | Current density (pA/pF) |
|----------------|---|----------------------|-------------------------|
| Control        | 16/18 (89%)   | $3.4 \pm 0.6$        | $22.1 \pm 2.4$          |
| E4031          | 14/18 (78%)   | $5.1 \pm 0.6$        | $42.2 \pm 2.5$          |
| TRAM-34        | 0/11 (0%)   | -                    | -                       |
| TRAM-34 +E4031 | 5/7 (71%)   | $1.7 \pm 0.2$        | $3.2 \pm 0.8$           |



E

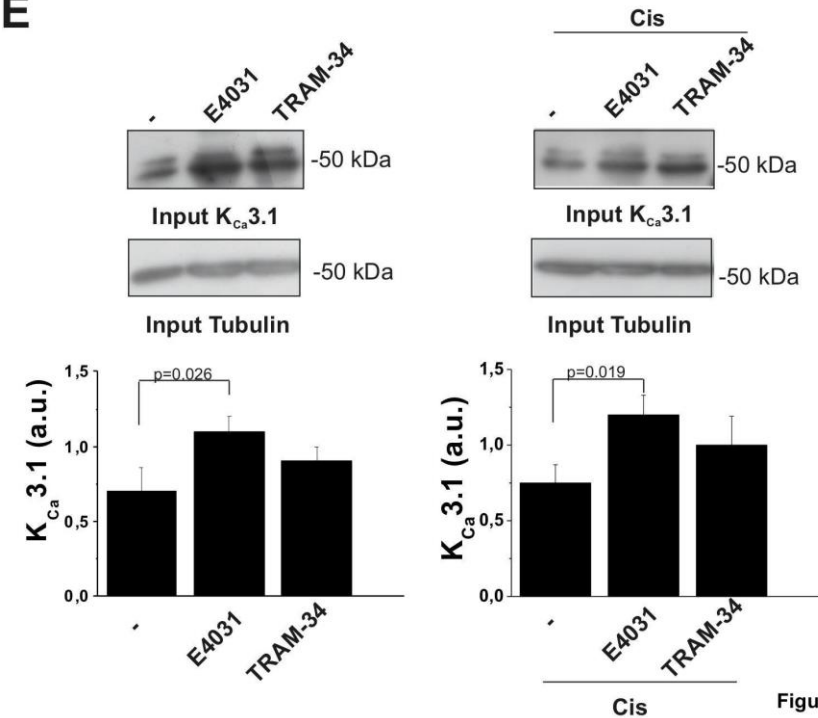
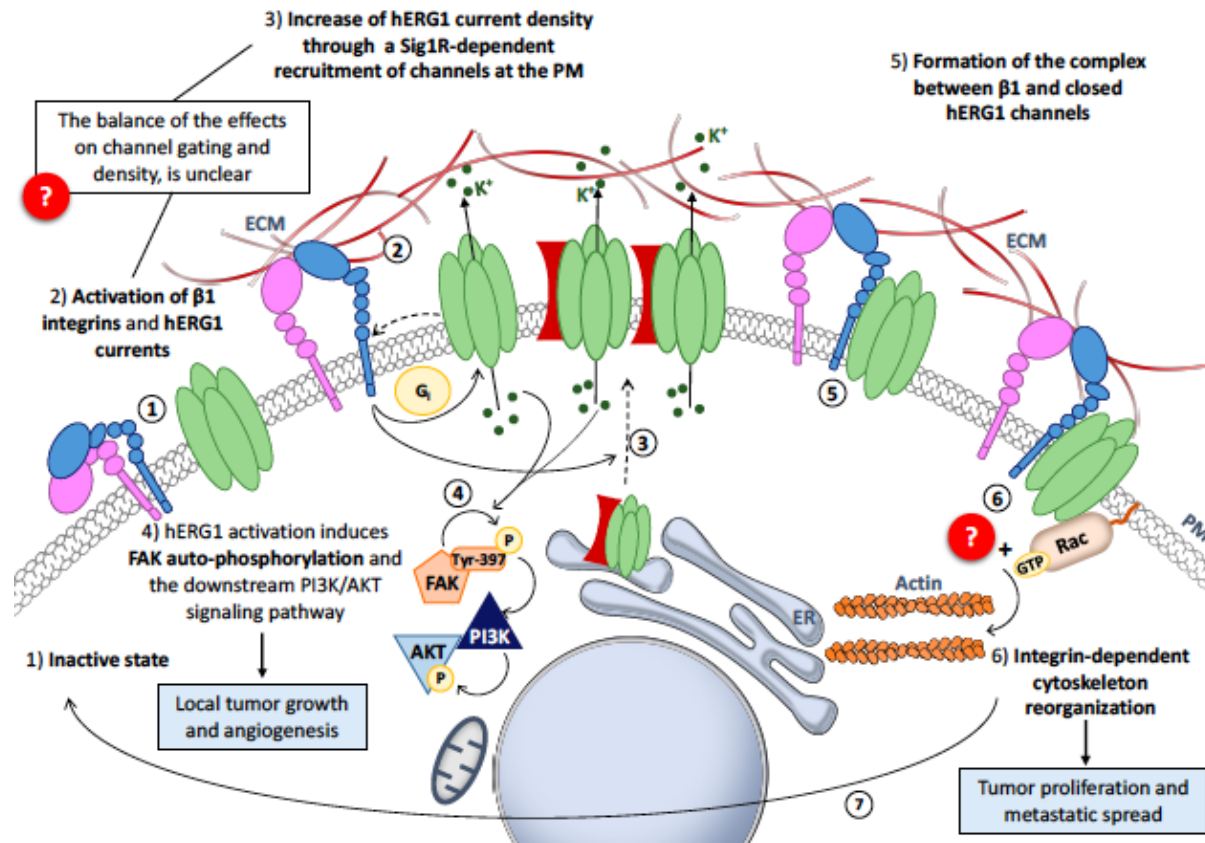


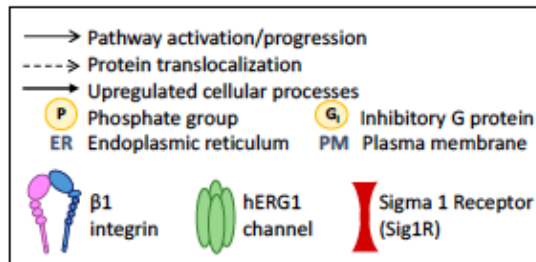
Figure 4



# hERG1 / $\beta 1$



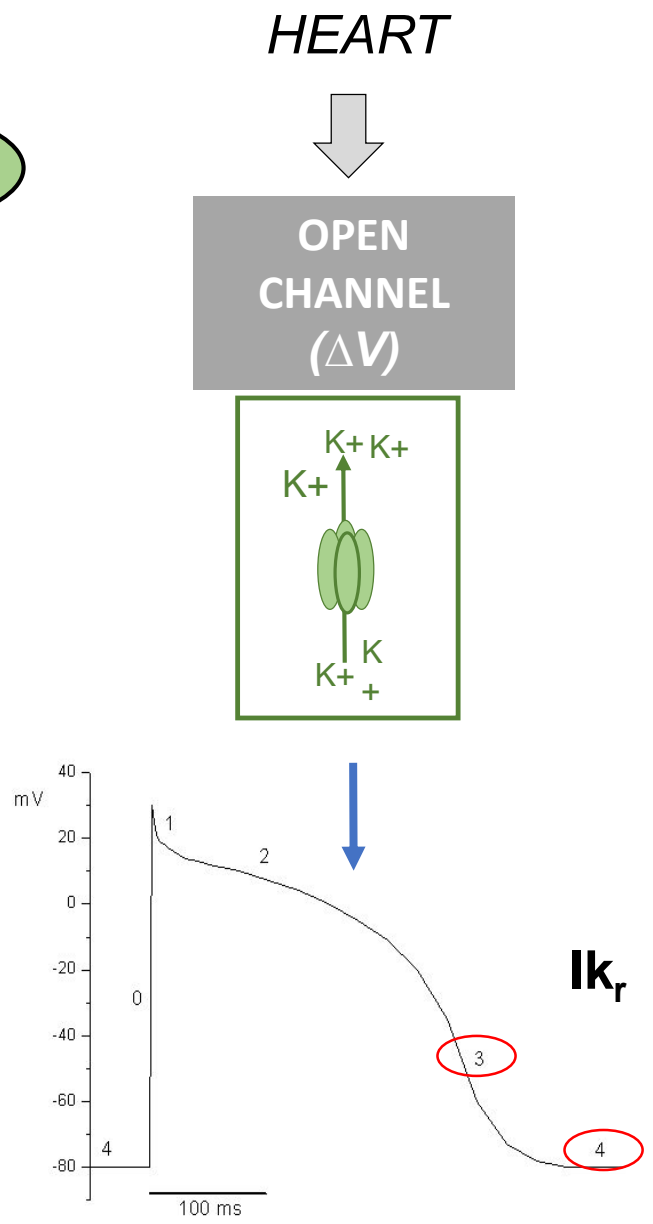
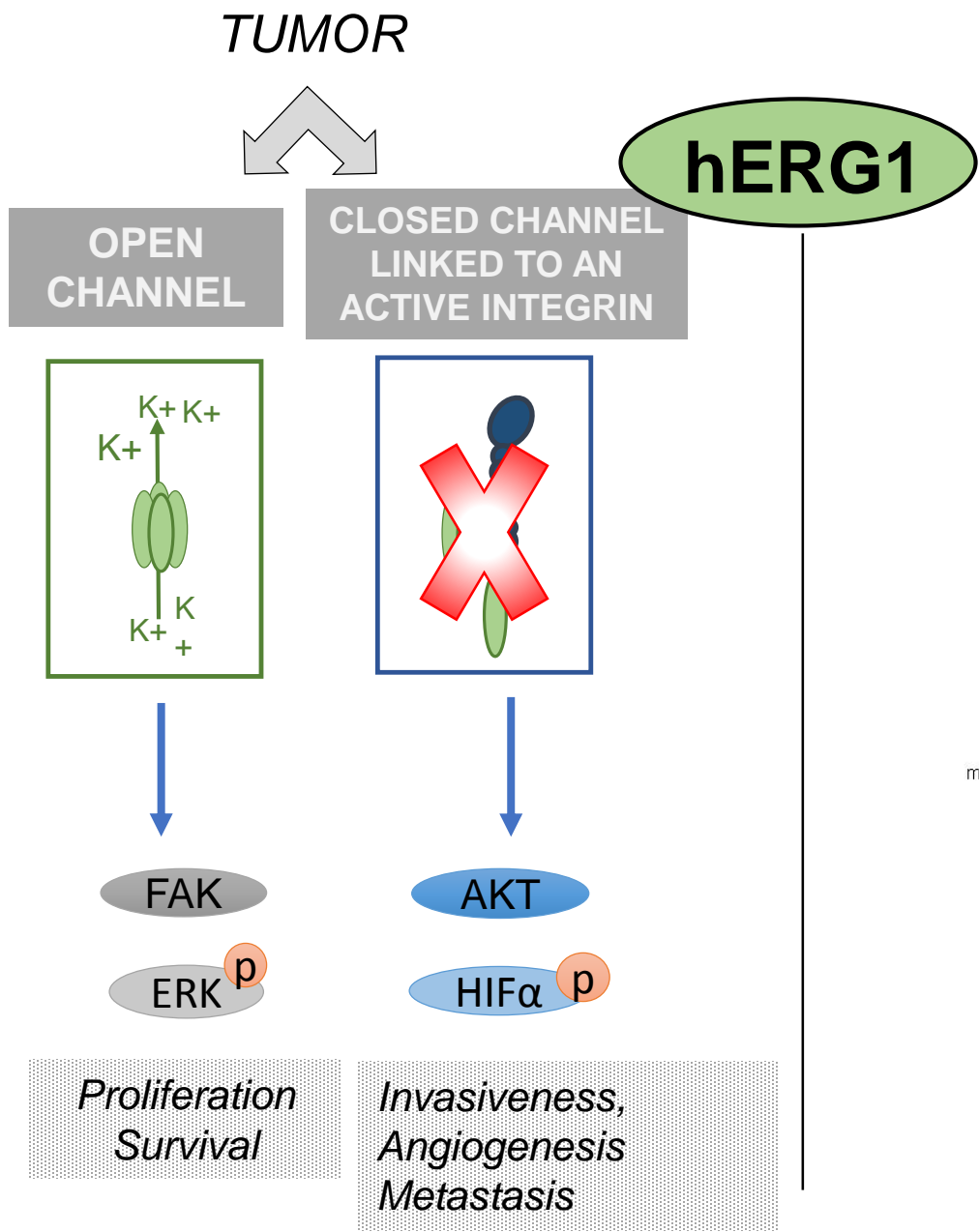
hERG1/ $\beta 1$  activates cancer-specific intracellular pathways (PI3K/AKT, HIF 1/2  $\alpha$ , small GTPases (Rac1), f-actin remodeling)



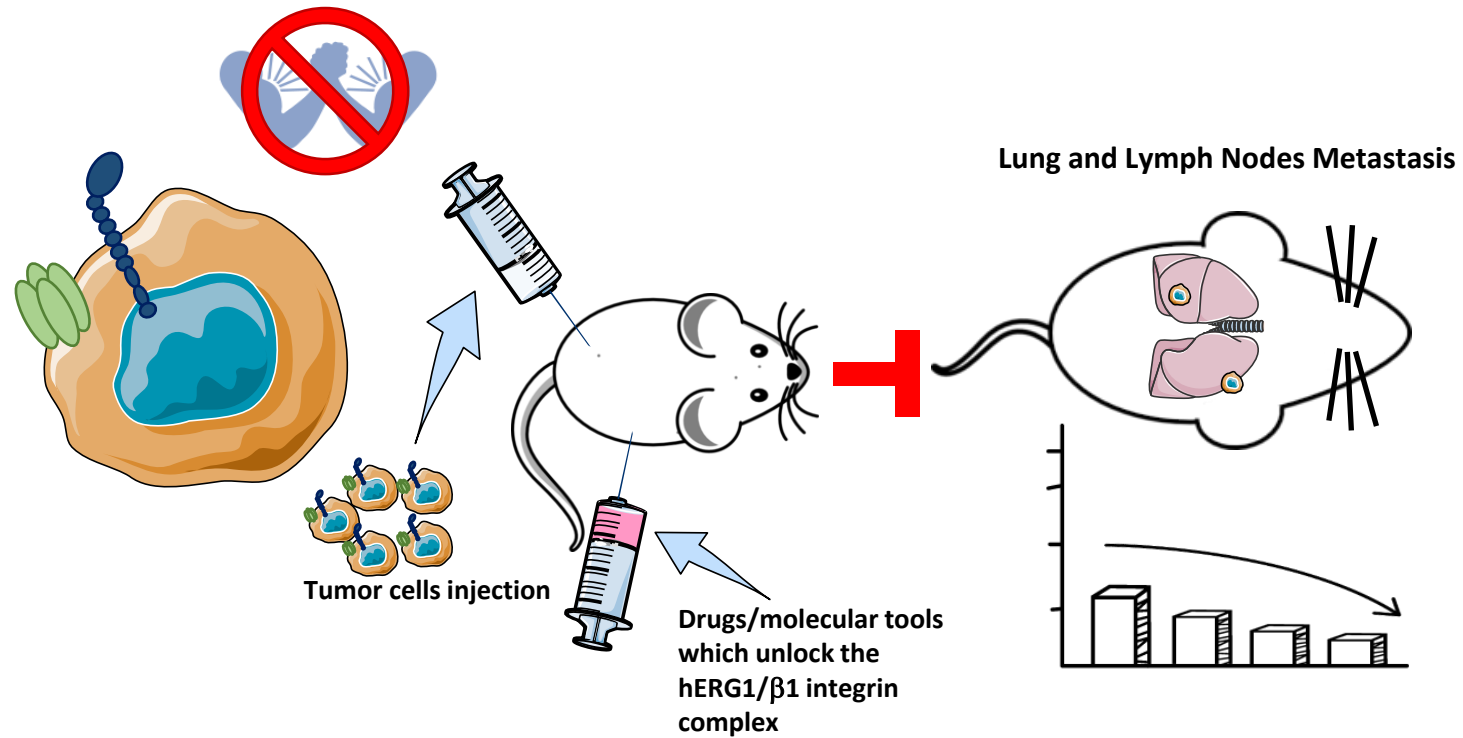
Becchetti A et al., Science Signaling, 2017

Becchetti A, Petroni G and Arcangeli A, Trends Cell Biol., 2019

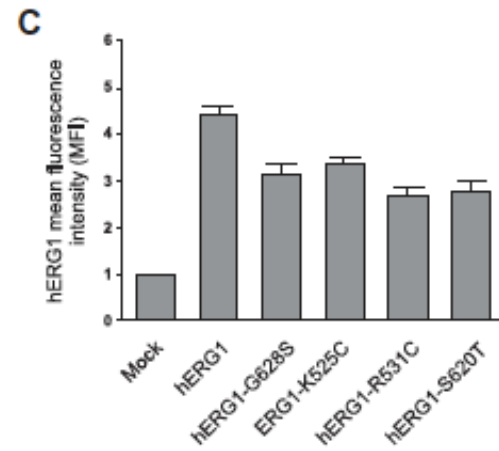
<https://www.youtube.com/watch?v=eqj2NXzvmpps>



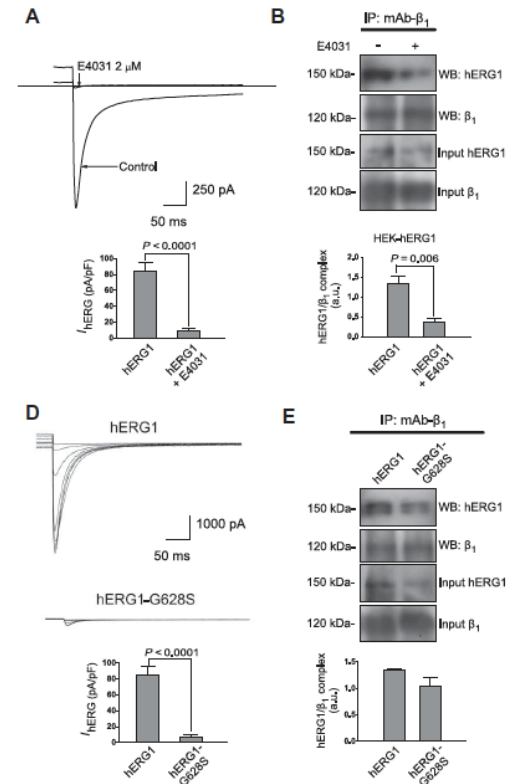
# Disrupting the hERG1/ $\beta$ 1 integrin complex inhibits tumor metastasis



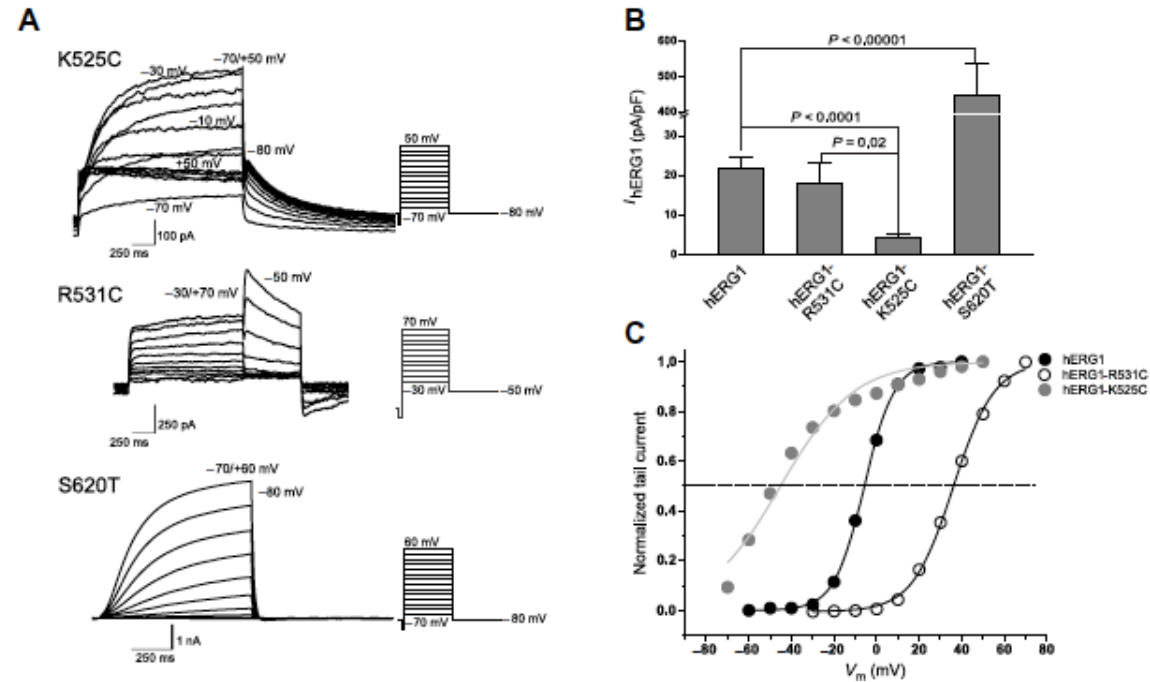
# The *hERG1* conformational state determines (the closed state favours) integrin association



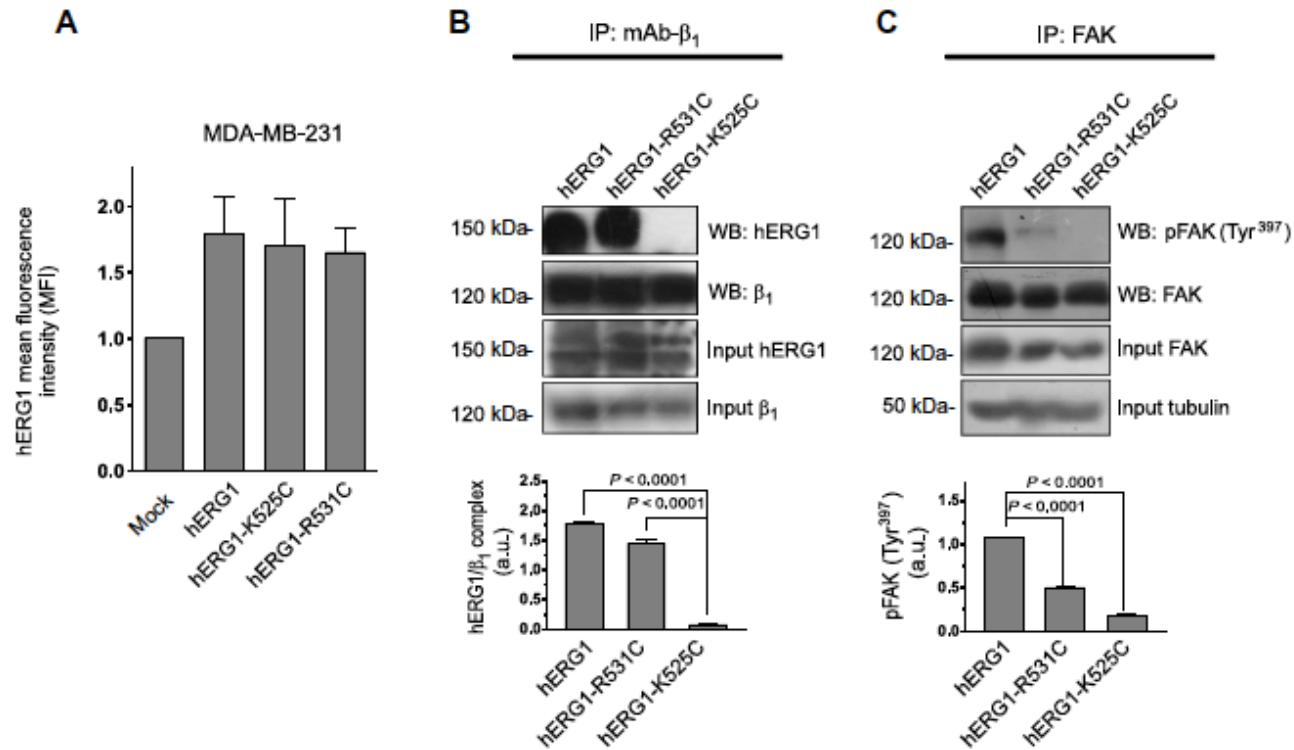
*hERG1* mutants:  
 G628S: non conductive  
 S620T: non inactivating  
 K525C: S4 (voltage sensor) mutant\*  
 R531C: S4 (voltage sensor) mutant\*  
 \*=alterations of gating



The hERG1 conformational state determines (the closed state favours) integrin association



The hERG1 conformational state determines (the closed state favours) integrin association: MDA-MB-231 breast cancer cells



*The hERG1 conformational state determines (the closed state favours) integrin association.....and tumor metastasis (MDA-MB-231 breast cancer cells)*

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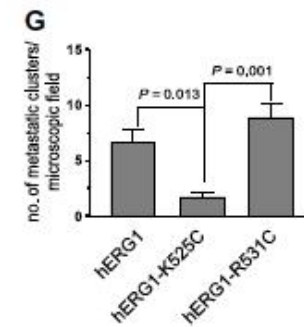
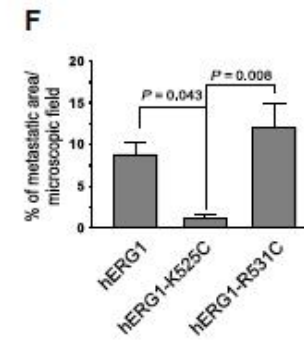
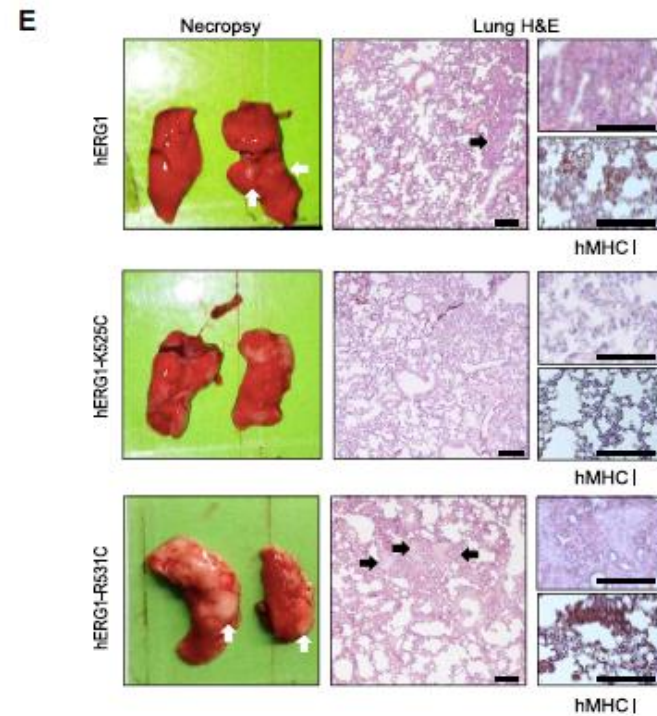
**D**

|  | MDA-MB-231 | hERG1        | hERG1-K525C  | hERG1-R531C  |
|--|------------|--------------|--------------|--------------|
| <b>Local tumor growth</b>                      |            |              |              |              |
| Number of tumor masses (%)                     |            | 9/10 (90%)   | 10/10 (100%) | 9/10 (90%)   |
| Median tumor volume (mm <sup>3</sup> )         |            | 150 (19–300) | 122 (33–300) | 212 (33–300) |
| <b>Metastases</b>                              |            |              |              |              |
| <b>Inguinal lymph nodes</b>                    |            |              |              |              |
| Number of mice with macroscopic metastases (%) |            | 2/5 (40%)    | 0/5 (0%)     | 3/5 (60%)    |
| <b>Lung</b>                                    |            |              |              |              |
| Number of mice with macroscopic metastases (%) |            | 2/5 (40%)    | 0/5 (0%)     | 4/5 (80%)    |



*The hERG1 conformational state determines (the closed state favours) integrin association.....and tumor metastasis (MDA-MB-231 breast cancer cells)*

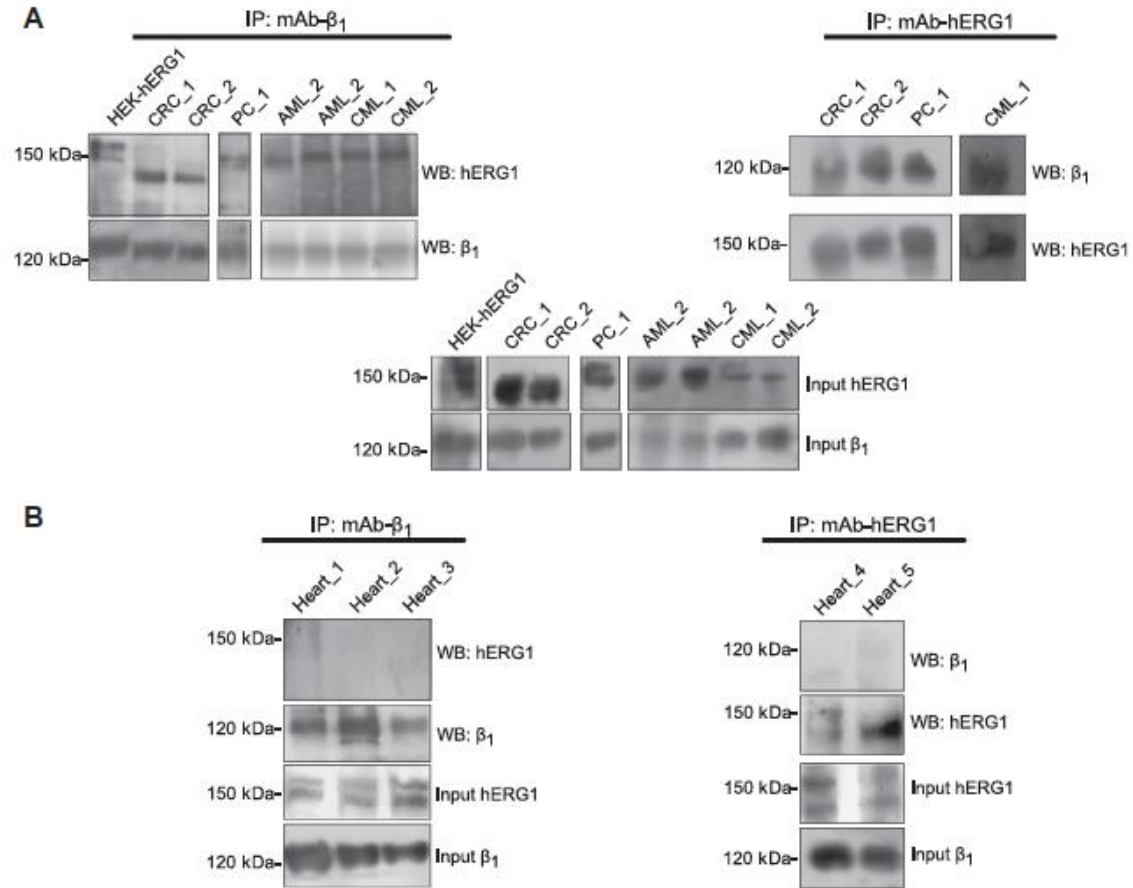
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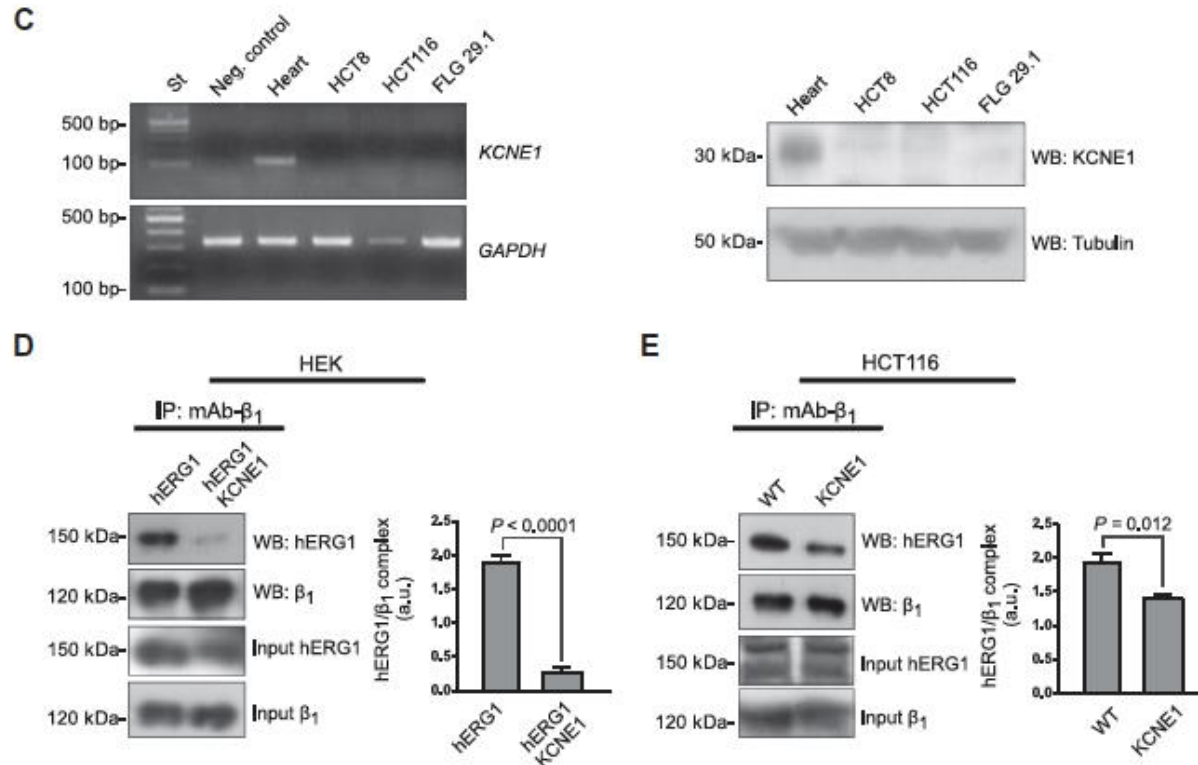


# The hERG1/β<sub>1</sub> complex occurs in tumour cells, but not in the heart

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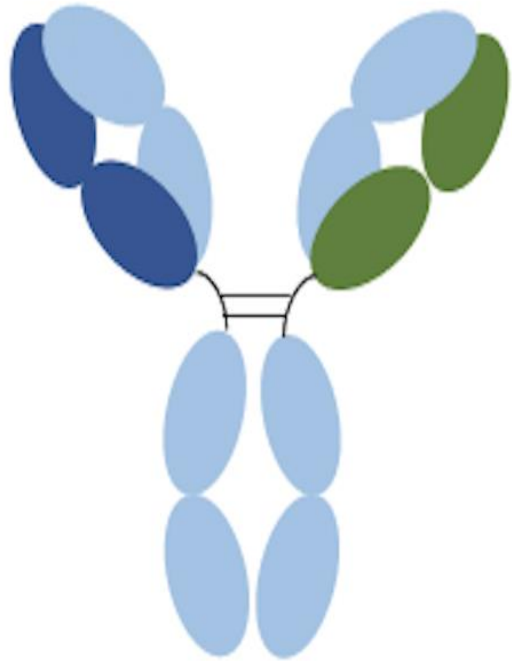
... because tumour cells do not express “canonical”  
(KCNE1) beta subunits



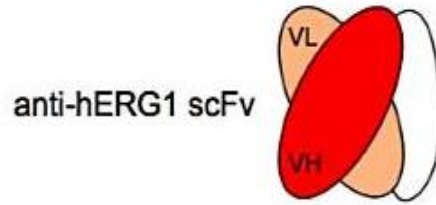
# PIPELINE DEVELOPMENT STRATEGY

DESIGNING NEXT GENERATION ANTIBODY MOLECULES

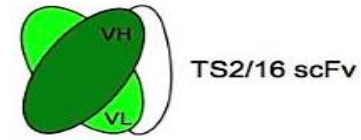
FROM BIG MONOCLONAL ANTIBODY TO SMALL BISPECIFIC DIABODY



**Anti-hERG1 Monoclonal Antibody**  
(M.W. 160 kDa)



**Anti-hERG1 Single-Chain Variable Fragment (Sc-FV)**

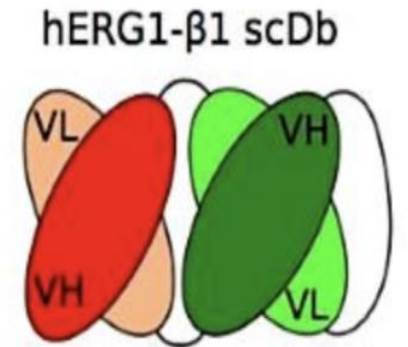


**Anti-integrin beta1 Single-Chain Variable –Fragment (Sc-fV)**



**Anti hERG1/integrin beta1 BISPECIFIC Antibody (DIABODY)**  
(M.W. 50 kDa)

**THERAPEUTIC ANTIBODY**  
**MCKAA2017THE01**



TUMOR

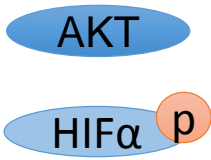
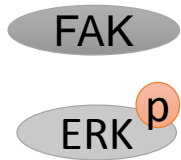
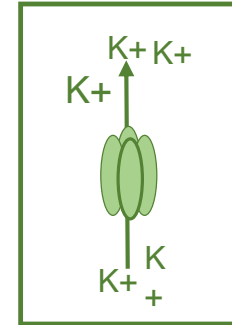
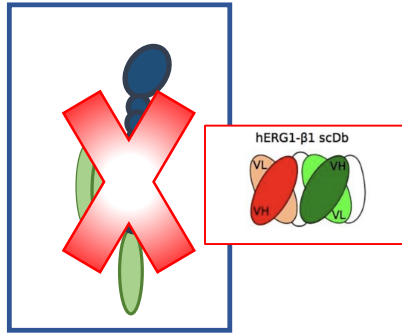
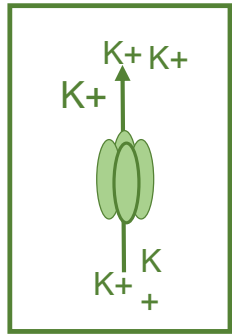
HEART

hERG1

OPEN CHANNEL

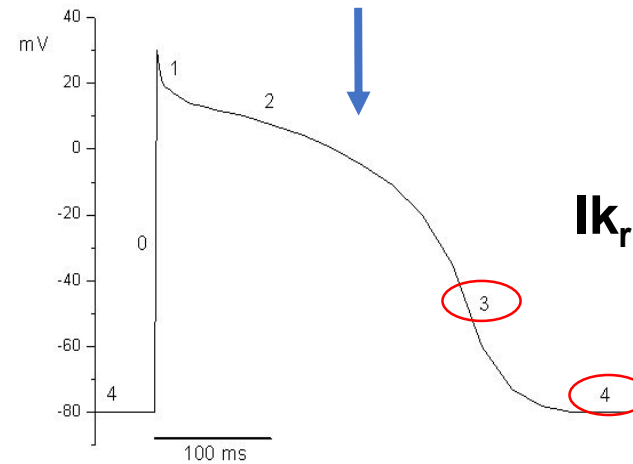
CLOSED CHANNEL LINKED TO AN ACTIVE INTEGRIN

OPEN CHANNEL ( $\Delta V$ )



Proliferation  
Survival

Invasiveness,  
Angiogenesis  
Metastasis



# CONCLUSIONS

- ❖ Ion channels are relevant in cancer (biomarkers!)
- ❖ Ion channels can exert both conductive and non conductive effects in cancer cells
- ❖ hERG1 has both conductive (ion flux-mediated) and non conductive (once bound to integrin receptors in the closed state) activities
- ❖ hERG1 mediates tumour progression (e.g. proliferation, invasion, angiogenesis, metastasis....)
- ❖ hERG1 can be considered a novel cancer biomarker
- ❖ The hERG1/beta1 integrin complex can be considered a therapeutic target in cancer and can be targeted through newly developed bispecific antibodies

# Acknowledgements:



**Prof.A.  
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Bicocca, Italy



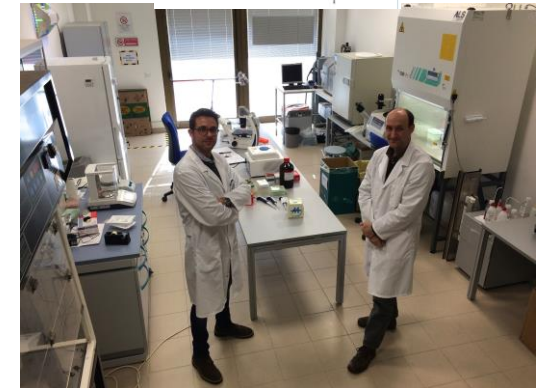
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MCK THERAPEUTICS



**ASSOCIAZIONE ITALIANA  
PER LA RICERCA SUL CANCRO**  
AIRC *Con la ricerca, contro il cancro*



Supported By:

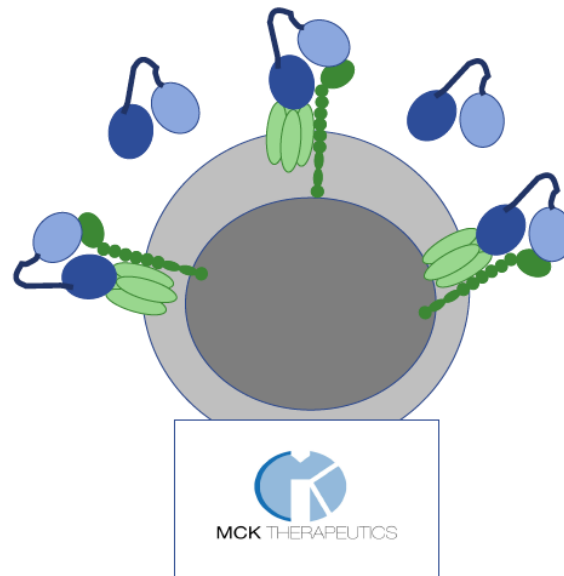
**ASSOCIATION  
FOR INTERNATIONAL  
CANCER RESEARCH**



# MCKAA2017THE01

**MCKAA2017THE01** is a highly innovative **BISPECIFIC SINGLE CHAIN DIABODY** molecule able to **target the hERG1/ $\beta$ 1 biomarker in pancreatic cancer.**

The DIABODY MCKAA2017THE01 has a double antigenic binding sites within the pancreatic cancer-specific antigen hERG1/ $\beta$ 1, **while DOES NOT bind hERG1 in cardiac myocytes**



Besides MCKAA2017THE01

# Novel companion diagnostics linked to MCKAA2017THE01





# MCK THERAPEUTICS APPROACH TO PRECISION CARE OF PANCREATIC CANCER

## DEVELOPING A FULL SET OF PRODUCTS FOR THERAPY, SCREENING & EARLY DETECTION OF PDAC

### DIAGNOSTIC COMPANIONS

### THERAPEUTICS

**MCKAA2018DIA03**

**IN VIVO MOLECULAR  
IMAGING**

**MICROMETASTASIS**

**PANCREATIC CANCER**

**MCKAA2017THE01**

**MCKAA2017DIA01**

**IN VITRO (IHC)**

**MCKAA2017DIA02**

**IN VITRO (IF)**

**SINGLE/COMBINATION  
TREATMENT AGENT**

**EARLY/ADVANCED  
(METASTATIC) STAGES**



# Acknowledgements:



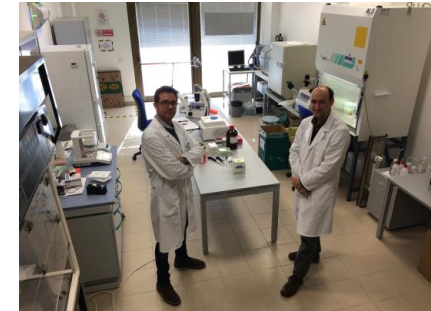
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**MCK** THERAPEUTICS

IMPROVING HEALTH & QUALITY OF LIFE OF PATIENTS WITH  
HUGE UNMET MEDICAL NEEDS